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TOS SCIENCE DESIGN FOR VECTORS AND PATHOGENS

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Change Record

REVISION	DATE	ECO #	DESCRIPTION OF CHANGE
A	09/24/2014	ECO-02246	Initial release
B	06/15/2018	ECO-05596	<ul style="list-style-type: none">• Minor format edits• corrected protocol names in reference documents• updated NEON TOS site number from 60 to 47• added testing rationale for mosquitoes• updated sampling schedule for ticks (MODIS) and threshold for high intensity sampling (data added as appendix)• updated list of tick-borne pathogens being tested, removed mention of high-throughput sequencing and gut microbiota, and added limiting testing to only nymphal ticks• added Guanica tick plot establishment parameters• updated to indicate selection of submandibular as the blood collection method• updated small mammal sampling frequency to reflect current design
C	04/06/2022	ECO-06790	<ul style="list-style-type: none">• Revised logo• Update to reflect change in terminology from relocatable to gradient sites



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1 DESCRIPTION

1.1 Purpose

NEON design documents are required to define the scientific strategy leading to high-level protocols for NEON subsystem components, linking NEON Grand Challenges and science questions to specific measurements. Many NEON *in situ* measurements can be made in specific ways to enable continental-scale science rather than in ways that limit their use to more local or ecosystem-specific questions. NEON strives to make measurements in ways that enable continental-scale science to address the Grand Challenges. Design Documents flow from questions and goals defined in the NEON Science Strategy document and inform the more detailed procedures described in Level 0 (LO; raw data) protocol and procedure documents, algorithm specifications, and Calibration/Validation (CalVal) and maintenance plans.

1.2 Scope

This document defines the rationale and requirements for vector and pathogen sampling in the NEON Science Design.

1.3 Acknowledgements

The design for each module was reviewed by and refined with input from technical working groups consisting of academic and public health researchers with relevant expertise:

Tick and tick-borne pathogens working group (2011 – 2016): Drs. Brian Allan, Ben Beard, Lorenza Beati, Dustin Brisson, Maria Diuk-Wasser, Rebecca Eisen, Holly Gaff, Sarah Hamer, Nicholas Ogden, Rick Ostfeld, Joe Piesman, Daniel Sonenshine, Andrea Swei, Michael Yabsley

Mosquito working group (combined for abundance/diversity/phenology and mosquito-borne pathogen designs) **(2011 – 2016):** Drs. Chris Barker, Roberto Barrera, Mark Blackmore, William Bradshaw, Desmond Foley, Howard Ginsberg, Mary Hayden, Chris Holzapfel, Steve Juliano, Laura Kramer, Shannon LaDeau, Chet Moore, Roger Nasci, Bill Reisen, Harry Savage

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2 RELATED DOCUMENTS AND ACRONYMS

2.1 Applicable Documents

Applicable documents contain information that is applied in the current document. Examples are higher level requirements documents, standards, rules and regulations.

AD[01]	NEON.DOC.000001	NEON Observatory Design
AD[02]	NEON.DOC.000913	NEON TOS Science Design for Spatial Sampling
AD[03]	NEON.DOC.014045	TOS Protocol and Procedure: Tick and Tick-borne Pathogen Sampling
AD[04]	NEON.DOC.014049	TOS Protocol and Procedure: Mosquito Sampling
AD[05]	NEON.DOC.000481	TOS Protocol and Procedure: Small Mammal Sampling
AD[06]	NEON.DOC.002652	NEON Level 1, Level 2 and Level 3 Data Products Catalog
AD[07]	NEON.DOC.000910	NEON TOS Science Design for Mosquito Abundance, Diversity, and Phenology
AD[08]	NEON.DOC.000915	NEON TOS Science Design for Small Mammal Abundance and Diversity
AD[09]	NEON.DOC.014015	NEON Bioarchive: Concept of Operations

2.2 Reference Documents

Reference documents contain information complementing, explaining, detailing, or otherwise supporting the information included in the current document.

RD[01]	NEON.DOC.000008	NEON Acronym List
RD[02]	NEON.DOC.000243	NEON Glossary of Terms

2.3 Acronyms

All acronyms used in this document are defined in RD[01].



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3 INTRODUCTION

3.1 Overview of the Observatory

The National Ecological Observatory Network (NEON) is a continental-scale ecological observation platform for understanding and forecasting the impacts of climate change, land use change, and invasive species on ecology. NEON is designed to enable users, including scientists, planners and policy makers, educators, and the general public, to address the major areas in environmental sciences, known as the Grand Challenges (**Figure 1**). NEON infrastructure and data products are strategically aimed at those aspects of the Grand Challenges for which a coordinated national program of standardized observations and experiments is particularly effective. The open access approach to the Observatory’s data and information products will enable users to explore NEON data in order to map, understand, and predict the effects of humans on the earth and understand and effectively address critical ecological questions and issues. Detailed information on the NEON design can be found in AD[01] and AD[02].

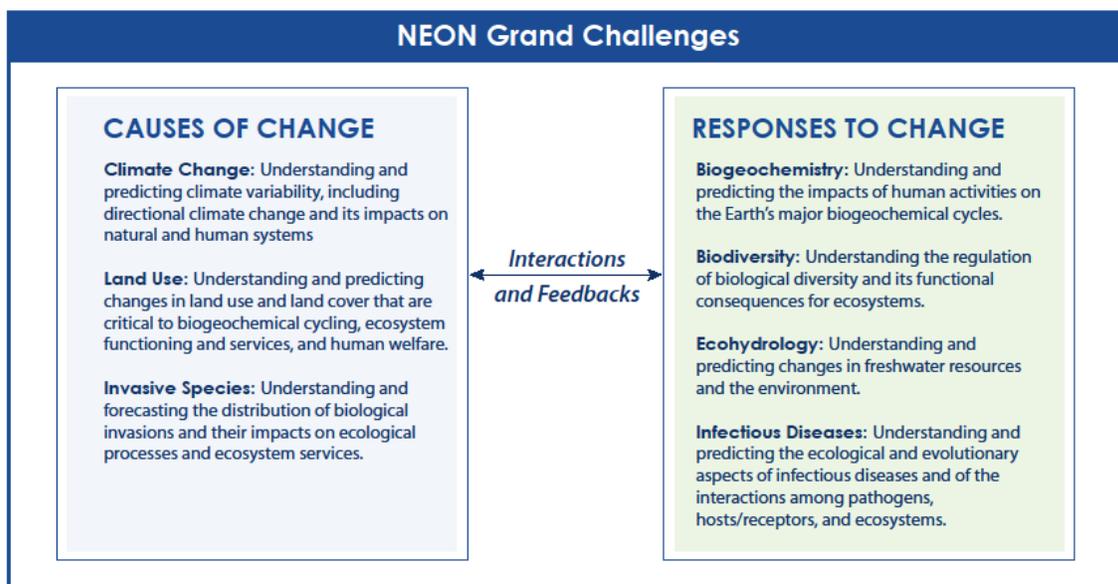


Figure 1. The seven Grand Challenges defined by the National Research Council (2001).

3.2 Components of the Observatory

There are five components of the Observatory, the Airborne Observation Platform (AOP), Terrestrial Instrument System (TIS), Aquatic Observation System (AOS), Aquatic Instrument System (AIS), and Terrestrial Observation System (TOS). Collocation of measurements associated with each of these components will allow for linkage and comparison of data products. For example, remote sensing data provided by the Airborne Observation Platform (AOP) will link diversity and productivity data collected on individual plants and stands by the Terrestrial Observation System (TOS) and flux data captured by instruments on the tower (TIS) to that of satellite-based remote sensing. For additional information on these systems, see Keller et al. 2008, Schimel et al. 2011.



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3.3 The Terrestrial Observation System (TOS)

The NEON TOS will quantify the impacts of climate change, land use, and biological invasions on terrestrial populations and processes by sampling key groups of organisms (sentinel taxa), infectious disease, soil, and nutrient fluxes across system interfaces (air, land, and water) (AD[01], AD[02]). The sentinel taxa were selected to include organisms with varying life spans and generation times, and wide geographic distributions to allow for standardized comparisons across the continent. Many of the biological measurements will enable inference at regional and continental scales using statistical or process-based modeling approaches. The TOS sampling design captures heterogeneity representative of each site to facilitate this inference when possible. Plot and organism-scale measurements will also be coordinated with the larger-scale airborne measurements, which provide a set of synergistic biological data products at the regional scale. The standardization of protocols across all sites is key to the success of NEON (and its novelty) and must be maintained at all sites through time. Thus, although specific techniques may be required at some sites (e.g., due to different vegetation types), protocols have been developed to ensure data comparability. Details of these design elements can be found in individual design documents available through the NEON website (www.neonscience.org).

The TOS Science Designs define the scientific strategies leading to high-level sampling designs for NEON sentinel taxa, terrestrial biogeochemistry, and infectious disease, linking NEON Grand Challenges and science questions to specific measurements (AD[01]). The TOS Spatial Sampling Design document describes the sampling design that collocates observations of the components of the TOS and allocates sampling locations throughout a site according to a stratified-random design (AD[02]). TOS Science Design documents were developed following input from the scientific community, including discipline-specific Technical Working Groups, and the National Science Foundation. Science Designs will be reviewed periodically to ensure that the data collected by NEON are those best suited to meet the requirements of the observatory (AD[01]), are (to the extent possible) consistent with standards used by the scientific community, and fit within the scope of NEON.



4 INTRODUCTION TO THE VECTOR AND PATHOGEN SAMPLING DESIGN

4.1 General Background

Parasites and pathogens (hereafter, pathogens) are important drivers of ecological and evolutionary changes in natural, agricultural, and urban ecosystems and have exerted significant effects on the demography and culture of human populations throughout history (Dobson and Grenfell 1995, Dobson and Carper 1996, Harvell et al. 1999, Swabe 1999, Daszak et al. 2000, Taylor et al. 2001, Hudson et al. 2002, Strange and Scott 2005, Alexander 2010, Brown and Gilfoyle 2010, McNeill 2010, Brooks and Hoberg 2013). In recent decades, the number of emerging and re-emerging pathogens and associated infectious diseases, many of which are vector- and reservoir-borne zoonoses of significant public health concern, has increased dramatically (Garnett and Holmes 1996, Dobson and Foutopoulos 2001, Taylor et al. 2001, Anderson et al. 2004, Morens et al. 2004, Woolhouse and Gowtage-Sequeria 2005, Jones et al. 2008, but see Rosenberg et al. 2013). This trend is believed to reflect both the growing epidemiological connectivity between natural and human-associated systems as well as anthropogenic environmental modification: changes in climate, land- and resource-use practices and patterns of human trade and travel have enormous potential to alter patterns of infection and disease dynamics (Coakley et al. 1999, Patz et al. 2000a, Gubler et al. 2001, Harvell et al. 2002, Patz et al. 2004, Weiss and McMichael 2004, Wolfe et al. 2005, Wolfe et al. 2007, Lafferty 2009, Rohr et al. 2011, Altizer et al. 2013, Perry et al. 2013). In some cases, increases in the frequency and severity of pathogen outbreaks, particularly those arising from cross-system infection events (e.g., zoonoses, anthroponoses), are predicted (Marano et al. 2007, Myers et al. 2013). In others, environmental change may result in pathogen losses, some of which will have important public health or economic consequences (Gomez and Nichols 2013). In both scenarios, changes in the abundance of vectors, reservoirs, or pathogens, and in the epizootiology (study of the causes and outcomes of diseases in animal populations) and epidemiology (study of diseases in human populations) of associated diseases, may have important implications for the health of human and livestock populations and the conservation of wildlife (Gubler 1998, Binder et al. 1999, Daszak et al. 2000, Cleaveland et al. 2001, Daszak et al. 2001, Strange and Scott 2005, Thompson et al. 2010). These changes could also have unanticipated effects on ecological communities at large, particularly when individuals of these species play influential roles in community-level interactions or ecosystem function (Mitchell and Power 2003, Hudson et al. 2006, Hatcher et al. 2014).

In light of these patterns and predictions, there is a clear need for increased pathogen surveillance efforts that bridge the historical divisions among human, domesticated animal, and wildlife diseases (e.g., epidemiology of human and domesticated animal diseases versus epizootiology and ecology of wildlife diseases). For instance, the field of conservation medicine and the OneHealth initiative are both predicated on a synthetic approach that emphasizes the linkages between environmental change and the health of human and wildlife populations (Daszak et al. 2004, Kaplan et al. 2009, Rock et al. 2009, Atlas et al. 2010, Coker et al. 2011, Aguirre et al. 2012). The importance of understanding the particular links between the dynamics of natural ecosystems and human health was highlighted as a critical priority by the National Research Council (National Research Council 2001, 2003) and the Millennium



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Ecosystem Assessment (Patz et al. 2005). Given the broad spatiotemporal extents over which many important changes in disease dynamics are likely to occur, the myriad factors that could underlie those changes, and the broad spatiotemporal scales over which some of those patterns (e.g., climate- and land use-driven) are likely to become apparent, the value of multifaceted surveillance efforts is increasingly recognized (Altizer et al. 2013). As the size and scope of surveillance efforts expands, appropriate sampling design and methodological standardization greatly facilitate comparisons across both datasets and scales. Although logistically challenging, such large-scale efforts are critical to characterize regional, continental, and multi-decadal patterns of disease dynamics. Insights gleaned from such projects hold promise for informing efforts to promote human health and wildlife conservation while furthering our fundamental understanding of the ecology and evolution of host-pathogen interactions (Kovats et al. 2001, Crowl et al. 2008).

In 2012, the United States National Science Foundation began funding the construction of the National Ecological Observatory Network (NEON) with the goal of creating the first continental-scale ecological monitoring system (Keller et al. 2008). Using standardized methods implemented at 47 sites for up to 30 years, NEON will provide insights into the effects of global change drivers (e.g., climate and land-use change) on the physical and ecological environment across multiple spatial and temporal scales (www.neonscience.org, Schimel et al. 2011). As part of NEON's terrestrial observation system (Kao et al. 2012), sampling of pathogens and associated vector and reservoir species will be conducted to elucidate the changing ecology of a suite of tick-, mosquito-, and rodent-borne pathogens. Among these are the etiological agents of important human diseases including Lyme borreliosis, Hantavirus pulmonary syndrome, and West Nile virus disease. The archiving of samples collected as part of this effort also has the potential to provide baselines for detecting future epizootiological shifts in other pathogens not currently known to be of importance to human or animal health. Measures of vector/reservoir abundance and pathogen prevalence will be made simultaneously with a suite of environmental and organismal information collected at NEON sites. These include a variety of measurements of abiotic environmental conditions, remotely-sensed data on vegetation and biogeochemistry, and phenological, demographic, and biodiversity information on sentinel taxa that include soil microbes, plants, insects, birds, and small mammals. To promote the advancement of open-access science, NEON data and archived physical samples will be freely available for additional analyses by members of the research community. Given its broad spatiotemporal scope, emphasis on methodological standardization, numerous and varied eco-environmental foci, and open-access policies, the NEON project will advance our current understanding of and ability to predict changes in host/pathogen interactions and associated disease dynamics in novel, inter-disciplinary, and collaborative ways.

NEON's vector and pathogen sampling broadly targets tick, mosquito, and small mammal (specifically, rodent) populations and associated pathogens. These vector/reservoir taxa were selected as sampling targets for two reasons. First, because of their physiology, ecology, and human associations, individuals and populations of these taxa are likely to respond quickly and measurably to changes in climate and land-use practices. Second, they play important roles in the transmission of a diverse suite of pathogens, many of which are of public health significance. The vector-borne framework and associated archiving of



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samples should enhance the long-term adaptability to accommodate previously uncharacterized and/or emerging pathogens. It also mirrors the organizational structure and operational approach employed by most public health agencies for vector-borne disease surveillance. The framework accommodates NEON's organizational focus on sentinel taxa and encompasses pathogens of interest identified in the early plans for the Observatory: West Nile virus and dengue viruses, *Borrelia burgdorferi* (primary etiological agent of Lyme borreliosis in North America), and Sin Nombre virus (an etiological agent of hantavirus pulmonary syndrome). Generally speaking, sampling will involve the collection of individuals of target vector/reservoir taxa to quantify their abundance, analysis of associated samples (tissues or whole organisms) to estimate the prevalence of infection by a particular suite of pathogens, and archive of some or all remaining samples for additional use by other members of the research community.

4.1.1 Ticks and Tick-Borne Pathogens

Ticks transmit a variety of pathogens, many of which are zoonotic and have considerable public health significance (Spach et al. 1993, Sonenshine 1994, Sonenshine and Roe 2014). In northern latitudes, tick-borne pathogens are responsible for the majority of cases of vector-borne diseases in humans (Randolph 2001) and Lyme borreliosis is the most frequently reported vector-borne disease in the United States (Centers for Disease Control and Prevention 2008). First recorded in the U.S. in 1975 (Steere et al. 1977), the reported number of human cases of Lyme borreliosis within the country has increased over 200% since 1992 (<http://www.cdc.gov/lyme/stats/index.html>, accessed on May 1, 2014). According to a recent analysis, up to 300,000 human infections may occur annually (Mead et al. 2013). Other zoonotic tick-borne pathogens in North America (e.g., *Anaplasma* spp., *Ehrlichia* spp., *Babesia* spp.) have exhibited similar patterns of emergence in terms of increases in prevalence and spatial extent over the past two decades (Childs and Paddock 2003, Doudier et al. 2010, Centers for Disease Control and Prevention 2012a). Tick-borne pathogens are associated with 13% of the infectious diseases currently classified by the CDC as nationally notifiable (<http://www.cdc.gov/nndss/>, accessed on May 1, 2014, Centers for Disease Control and Prevention 2010).

In addition to the public health significance of diseases associated with tick-borne pathogens, the physiology and ecology of ticks make them an ideal target for NEON sampling. Among arthropods, ticks are particularly sensitive to abiotic environmental conditions (Sauer and Hair 1986, Needham and Teel 1991, Stafford 1994, Dister et al. 1997, Jones and Kitron 2000, Teel et al. 2010, Sonenshine and Roe 2014) and associated physiological constraints make it highly likely that the demography and biogeography of ticks of many species, and the pathogens they transmit, will be affected by climate change (Estrada-Pena 2009, Gatewood et al. 2009, Diuk-Wasser et al. 2012, Leger et al. 2013, Medlock et al. 2013, Ogden et al. 2013). Further, the multi-host life cycles of ticks of most species increase their ecological connectivity and sensitivity to community-level perturbations associated with changes in human land- and resource-use patterns. These anthropogenic effects can manifest as direct alterations to the physical environment (Barbour and Fish 1993) or changes in community structure that include the abundance and diversity of available hosts (Childs and Paddock 2003, LoGiudice et al. 2003, Paddock and Yabsley 2007, Allan et al. 2010, Keesing et al. 2010).



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4.1.2 Mosquito-Borne Pathogens

The tremendous human health burden attributable to pathogens transmitted by mosquitoes is unsurpassed among vector-borne diseases. Globally it is estimated that in 2004 alone there were over 1.8 million human malaria deaths (Murray et al. 2012), and 96 million people are thought to experience disease associated with infection by dengue viruses each year (Bhatt et al. 2013). Moreover, mosquito-borne pathogens can also cause substantial reductions in populations of livestock and wildlife, with potentially important repercussions for human health, economic productivity, and the structure and function of ecological communities (e.g., Daubney et al. 1931, van Riper et al. 1986, Morris 1989, Scott and Weaver 1989, LaDeau et al. 2007, Paweska and van Vuren 2014).

Forecasts of potential ecological and public health consequences of climate change often focus on mosquitoes and the pathogens they transmit (Shope 1992, Reeves et al. 1994, Sutherst 2004). Although mosquitoes are found worldwide in all areas that are not permanently frozen, they are most consistently abundant in localities with tropical or moderately temperate climates where relatively warm and wet conditions prevail (although populations of some species in subarctic and alpine regions reach extremely high abundance during parts of the year). As a result, increases in temperature or precipitation at higher latitudes or elevations due to climate change may promote the range expansion of mosquitoes currently confined to tropical areas (Epstein et al. 1998, Patz et al. 2000b, Caminade et al. 2012, Eisen and Moore 2013, but see Reiter, 2001). This geographic spread could be facilitated by the incidental and periodic long-distance transport of mosquitoes that often occurs as part of human travel and international commerce (Lounibos 2002, Tatem et al. 2006). In North America and Europe, the potential introduction and establishment of dengue viruses, chikungunya virus, yellow fever virus, and Rift Valley fever virus are of particular concern (Gould and Higgs 2009, Weaver and Reisen 2010). Additionally, there is abundant evidence that changing climatic conditions will significantly affect mosquito demography and processes associated with the transmission of mosquito-borne pathogens (Mordecai et al. 2013). For example, changes in ambient temperature are predicted to alter mosquito vectorial capacity (Watts et al. 1987, Reisen et al. 2006, Paaijmans et al. 2012) and biting rates (Lardeux et al. 2008), and may in some cases catalyze host range shifts in arboviruses (Brault and Reisen 2013).

4.1.3 Rodent-Borne Pathogens

Small mammals, and rodents in particular, are common and influential members of most ecological communities and play central roles in the epidemiology and epizootiology of numerous pathogens. Rodentia is the most diverse order of the class Mammalia, including roughly 40% of extant species (Huchon et al. 2002). Rodents of some species are opportunistic foragers that have been synanthropic for thousands of years (e.g., Matisoo-Smith et al. 1998). Others inhabit the urban/wildland interface and may serve as epidemiological links between humans and infectious agents endemic to natural ecosystems (Kuenzi et al. 2001, Douglass et al. 2006). As a result of these intimate associations, and perhaps in part because of their shared mammalian pedigree, a variety of pathogens can be transmitted from rodents to humans (Meerburg et al. 2009). One of the most notable examples is the bacterium *Yersinia pestis*, the etiological agent of plague. The bacterium is usually transmitted to humans via the



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bite of a flea that has fed on an infected rodent (Gage and Kosoy 2005). Over the course of recorded history plague has been responsible for multiple human pandemics and tens of millions of fatalities (Gage et al. 2008). More recently, human infection by rodent-borne hantaviruses (Mills et al. 2010) has received considerable attention following an outbreak of hantavirus pulmonary syndrome (HPS) caused by Sin Nombre virus in the American southwest (Nichol et al. 1993, Centers for Disease Control and Prevention 2012b). Of the 17 hantaviruses identified in North America, six are known to cause HPS, which has a case fatality rate of roughly 36% (Mills et al. 2010, MacNeil et al. 2011). In addition to these highly publicized examples, rodents are known reservoirs for more than 35 bacterial and viral pathogens worldwide (<http://www.cdc.gov/rodents/index.html>, accessed May 1, 2014), many of which cause human diseases including Lyme borreliosis, typhus, babesiosis, and Rocky Mountain spotted fever (Meerburg et al. 2009).

Rodents and the pathogens they carry may also be model systems for investigating the effects of climate and land use changes on the epizootiology and epidemiology of zoonotic diseases. The often large sizes and high densities of rodent populations may favor the maintenance and spread of pathogens, and frequent contact between some rodent species and both domestic animals and human populations increases the likelihood of pathogen spillover or zoonotic transmission. Additionally, populations of many species of rodents that serve as reservoirs have high reproductive potential and turnover, attributes that promote rapid demographic responses to environmental changes with cascading effects on infection dynamics and the risk of human disease (Yates et al. 2002, Luis et al. 2010). Modifications of ecosystems by human activities, including urbanization and agricultural development, may also affect the structure of rodent communities in ways that alter the relative abundance of reservoir-competent species (Mills 2006, Clay et al. 2009).

4.2 NEON's Contribution

Data associated with NEON's vector and pathogen sampling programs will provide rare long-term datasets on pathogen dynamics at locations that span a range of anthropogenic impacts, from wildland areas to agricultural sites. These datasets will be additionally unique and valuable in that they will be generated using standardized methods across multiple sites that collectively span a variety of ecological systems. Datasets based on this spatially-replicated, long-term, methodologically standardized approach are exceedingly uncommon and should prove highly valuable in extending current understanding of the ecology of host/pathogen interactions and the effects of global change phenomena on associated transmission and disease dynamics. Vector and pathogen samples collected by NEON can further this understanding by providing the materials necessary for additional studies on topics including the biogeography, phylogeography, evolution, and coevolution of vectors, reservoirs, and pathogens.

4.3 Purpose and Scope

This document outlines the sampling design plans and associated rationale underlying NEON's three vector and pathogen sampling modules: tick-borne pathogens, mosquito-borne pathogens, and rodent-borne pathogens. It also provides cursory information about associated protocols. Associated details can



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be found in TOS Protocol and Procedure: Tick and Tick-borne Pathogen Sampling (AD[03]), TOS Protocol and Procedure: Mosquito Sampling (AD[04]), and TOS Protocol and Procedure: Small Mammal Sampling ([AD05]).



5 SAMPLING FRAMEWORK

The sampling designs for NEON’s tick-, mosquito-, and rodent-borne pathogen sampling modules are described below. Detailed information on the specific conception of and plans for each individual module is preceded by a discussion of the general design elements common across modules. Because sampling protocols flow from these designs, an understanding of the priorities and strategic decisions underlying the latter is integral to contextualizing when, where, and how the data and samples will be collected within NEON. This understanding should facilitate the use of NEON-generated resources and the replication (or modification) of NEON methods by other researchers or organizations attempting to integrate their work with or extend that being conducted by NEON.

5.1 Science Requirements

This science design is based on Observatory science requirements that reside in NEON’s Dynamic Object-Oriented Requirements System (DOORS). Copies of approved science requirements have been exported from DOORS and are available in NEON’s document repository, or upon request.

5.2 Data Products

Execution of the protocols that stem from this science design procures samples and/or generates raw data satisfying NEON Observatory scientific requirements. These data and samples are used to create NEON data products and are documented in the NEON Level 1, Level 2 and Level 3 Data Products Catalog (AD[06]).

5.3 Priorities and Challenges for Vector and Pathogen Sampling

5.3.1 Priorities and Challenges Common to All Three Sampling Modules

5.3.1.1 Prioritization of Time Series Data

Sampling designs emphasize a high frequency of intra-annual sampling to generate fine-scale time series data on changes in vector/reservoir abundance and pathogen presence/absence through time. When implemented over the relatively long lifespan of the observatory (up to 30 years at core sites), this approach should allow changes in sampled vector, reservoir, and pathogen populations to be characterized at two temporal scales. Of primary interest are interannual changes in the seasonal mean or maximum of vector/reservoir abundance and pathogen presence/absence or prevalence (**Figure 2A**). Given a sufficiently high sampling frequency, sampling should secondarily elucidate changes in aspects of interannual phenology of sampled populations (e.g., timing of onset and duration of seasonal cycles) (**Figure 2B**). The secondary focus is particularly important since the seasonal phenology of many vector, reservoir, and pathogen populations is expected to be sensitive to changes in climate and land-use practices (Altizer et al. 2013). Resulting phenological shifts could have profound implications for the maintenance, abundance, and spread of pathogens (Altizer et al. 2006, Fisman 2007).



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The emphasis on fine-scale time series data is driven largely by fundamental constraints on the spatial resolution of data generated by this sampling. Because NEON sites were not selected based on considerations of local vector or reservoir abundance or pathogen prevalence, not all sites will be productive in terms of sampling yields. Within sites, the number of sampling plots will generally be insufficient to characterize or measure changes in local habitat associations of targeted taxa. In light of these limitations, the designs seek to characterize changes in vector/reservoir abundance and pathogen presence/absence at the level of the site rather than of individual sampling plots. While plot-level field data will be available (e.g., number of adult ticks of species A collected at plot B within site C during sampling event D), data and samples should be aggregated across plots within sampling events at each site to make inferences at the site level (e.g., adult ticks of species A collected at all sampling plots within site C during sampling event D).

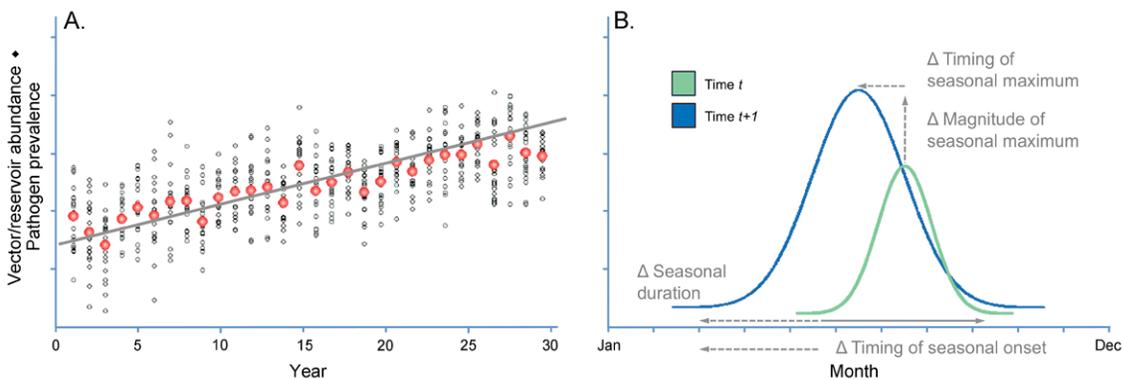


Figure 2. Hypothetical data illustrating detection of long-term (A) and seasonal (B) changes in vector/reservoir abundance and pathogen prevalence.

The allocation of sampling effort (e.g., number of sampling plots, frequency of sampling events) captured in module-specific sampling designs reflects the prioritization of high resolution time series data. In addition, designs attempt to incorporate considerations of anticipated resource availability, logistic constraints, and levels of effort commonly reported or deemed reasonable for surveys of tick-, mosquito-, and rodent-borne pathogens. Evaluating the adequacy of these plans through traditional power analyses is not wholly appropriate given that this approach is typically used to assess the design of studies motivated by one or a few specific research questions. Nevertheless, the module-specific sections below include results of power analyses evaluating the ability of proposed sampling plans to detect pathogens, and interannual trends in the seasonal mean prevalence of pathogens, at a NEON site. These analyses highlight just two of the many ways in which data collected by NEON vector/pathogen sampling could be used to test hypotheses about the dynamics of host/pathogen interactions and the epizootiology and epidemiology of associated diseases. More exhaustive statistical exploration of the power of NEON data when used in these various analyses is beyond the scope of this document.



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Initial NEON Science requirements associated with vector and pathogen sampling prescribed that the prevalence of infection by select pathogens (or immunological indications thereof) must be able to be quantified. This requires statistical power that is adequate relative to at least two criteria. First, sampling should be able to detect pathogens when they are present at biologically meaningful prevalence. Results of power analyses reported in the module-specific sections below provide quantitative estimates of detection probabilities under a variety of sampling scenarios. Second, sampling should be able to estimate prevalence with acceptable precision. In power analyses reported in the module-specific sections below, this is estimated as the ability to detect temporal changes in prevalence under a variety of sampling scenarios (notably, variation in trend magnitudes). A higher power to detect trends of smaller magnitudes is evidenced by smaller confidence intervals around the prevalence estimate at a given time point.

Because NEON will conduct tick- and rodent-borne pathogen testing through analysis of individual samples (i.e., not pooled), associated power analyses for trend detection used the negative binomial distribution. Due to the expectation of relatively low rates of infection, mosquito-borne pathogen testing will involve pooled samples (20-50 individual mosquitoes physically homogenized and the resulting homogenate tested for parasites). Because of this approach, power analyses for trend detection used the binomial distribution and the complementary log-log (CLL) link function, which accommodates for group testing within a GLM framework (Farrington 1992). Trend magnitudes were specified in terms of an annual increase in prevalence (values varied by module) and trend detection periods were set at either 10 or 25 years (approximations of the lifespans of NEON gradient and core sites, respectively). Year-zero infection prevalence was parameterized using values typically reported in the literature and/or commonly associated with the types of pathogens under consideration. Two combinations of type I error rate (α) and power ($1-\beta=1-p_{(\text{type II error})}$) were considered: a higher confidence scenario involving $\alpha=0.05$ and power=0.9 and a lower confidence scenario in which $\alpha=0.1$ and power=0.8. Calculations were performed both with and without a temporal autocorrelation term (e^{-1} at a half year). This value was chosen to represent a moderate magnitude of temporal correlation to provide contrast with respect to the case with temporal independence. While it is anticipated that there will be some temporal autocorrelation, the magnitude and lag will only be clarified through several years of sampling. Sampling frequencies considered in the analyses included those proposed at core and gradient sites for each sampling module as well as lower frequencies (e.g., one, two, three, and four times per year) to generate continuous power curves in accompanying figures. In these figures, green boxes are used to represent regions of sampling space where designs had sufficient power to detect trends of a specified magnitude. Unless otherwise noted in the figure captions, these boxes are bounded along the x-axis by sampling frequencies proposed for core and gradient sites and on the y-axis by anticipated levels of replication (i.e., number of field-collected samples or laboratory tests) associated with a single sampling event. Scripts used to perform these power calculations in R (R Core Team 2013) are available in Appendix A.

Prevalence as a metric, however, is likely not the most appropriate for such a large-scale endeavor as NEON, as (a) its interpretation is complicated by assumptions of homogenous mixing of hosts, vectors,



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and pathogens; (b) prevalence is extremely variable in space and time and, therefore, is not a reliable indicator of a site-level, multi-year evaluation of disease risk; and (c) it is one of multiple factors in disease risk, and is mathematically secondary to vector/host populations and behaviors (K. Tsao, pers. comm.). Therefore, methods detailed in Gu and Novak (2004) were used to also quantify the ability of the proposed designs simply to detect a pathogen when it is present at a site. Two approaches were used. First, the relationship between the number of samples tested and the probability of detecting a pathogen at various levels of infection prevalence was evaluated. Second, the relationship between infection prevalence and number of samples that would need to be tested to detect a pathogen with varying levels of statistical confidence was characterized.

5.3.1.2 Selection of Sampling and Testing Methods

Designs prioritize field sampling methods that meet three criteria. First, the methods must provide an effective means of collecting individuals of targeted vector/reservoir taxa. Second, utilization of well-established, widely employed methods should promote use of NEON data by the research community and the integration of these and similar data collected by other groups including private and academic researchers, local and state-level organizations, and federal agencies such as the U.S. Centers for Disease Control and Prevention (CDC) or U.S. Department of Agriculture (USDA). As an example, the CDC miniature CO₂ light trap has been used for decades in locations around the world to collect a variety of mosquitoes important in arbovirus transmission (Sudia and Chamberlain 1962, Newhouse et al. 1966, Pfuntner 1979). Analogous methods for the collection of ticks and small mammals include drag sampling (Milne 1943) and mark-recapture live trapping, respectively. Third, because standardization within the Observatory is emphasized to facilitate comparability of data across sites and through time, methods should be applicable across the spectrum of site-specific environmental conditions and ecology of sampled populations at NEON sites. While designs prioritize methods that can be used under a wide range of circumstances, they also reflect the reality that site-specific modifications may be desirable or necessary in some cases. For example, the primary design may be augmented through use of additional and supplementary methods (e.g., trap types) to increase sampling success at sites where vector/reservoir species are rare or not effectively sampled by methods employed in the primary design.

The majority of post-collection sample processing, including most taxonomic identification of vector/reservoir samples and all pathogen testing, will be performed on a contract basis by experts at external facilities. To increase cost efficiency, testing methods that can simultaneously detect more than one pathogen species are prioritized over the use of multiple pathogen-specific tests (e.g., family-specific assay for mosquito-borne flaviviruses rather than separate tests specific for West Nile virus, St. Louis encephalitis virus, and dengue viruses). Broadly reactive screens will be followed up with more specific assays to identify the pathogen(s) present in samples that test positive.

5.3.1.3 Selection of Sampling Plots

A fixed plot design will be used for vector/reservoir and pathogen field sampling. Selecting the locations of sampling plots at each site is complicated by the fact that in many cases, individuals of target taxa are likely



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to exhibit clustered patterns of distribution in space. This aggregation can arise from factors such as abiotic tolerances, specific habitat associations, and/or patterns of host-mediated dispersal. The productivity of sampling would be increased by locating at least a portion of sampling plots in or near areas of aggregation (e.g., adaptive cluster sampling, Thompson 1990, Brown et al. 2013). Identifying these within-site locations in a systematic way that can be applied across the observatory, however, is complicated by at least three issues. First, site-specific data on these locations are generally unavailable for most sites prior to the start of NEON sampling. Second, inferring patterns of local spatial distribution using regional data or expertise is problematic since the quality and availability of these resources vary considerably among sites and regions. Finally, because populations of target species often exhibit considerable interannual fluctuations in size, multiple years of within-site sampling would be required to empirically characterize patterns of site-specific distribution and/or local habitat associations with confidence.

Given these complications in systematically gathering accurate information on the within-site distribution of individuals of target taxa, sampling plots will be established at random locations within sites. This approach has a number of general strengths. First, because the goal of the sampling is to make site-level inferences about local vector, reservoir, and pathogen populations, the random distribution of sampling plots within sites allows for spatially unbiased site-level estimates of parameters of interest. Second, given that local patterns of distribution of individuals of target species may change over the lifespan of the observatory, plot locations based on current patterns of distribution might not be optimal in future years. A strategy of randomized plot distribution would be more robust to these changes. Finally, using the same approach to selecting plot locations as other NEON TOS sampling modules should facilitate statistical analyses and modeling involving multiple NEON data streams.

The design of each sampling module specifies a standard number of sampling plots per site. As detailed in the NEON TOS Science Design for Spatial Sampling (AD[02]), a stratified-random approach will be used to select plot locations within the dominant vegetation types ($\geq 5\%$ total cover) at each site. The number of sampling plots per type will be proportional to the percent cover of each type. Data collected at these plots during the first few years of sampling will be used to statistically evaluate whether the proposed number of plots within sites is sufficient to characterize parameters of interest with desired confidence. If it is determined that more data and/or samples are needed to achieve this confidence, additional sampling plots will be added as resources and logistics permit. Changes in the location of fixed plots can be made in consultation with members of the relevant technical working group(s) and the Observatory Director.

5.3.2 Priorities and Challenges for Mosquito-Borne Pathogen Sampling

The mosquito-borne pathogen sampling design was developed in conjunction with the design of sampling for mosquito abundance and diversity (for additional details see the NEON TOS Science Design for Mosquito Abundance, Diversity, and Phenology, AD[07]). Because these two sampling efforts differ fundamentally in their objectives, they would be optimized using different strategies if designed independently. Pathogen-related sampling seeks to collect and test large numbers of particular mosquito species to quantify rare phenomena (infection by pathogens). In contrast, abundance and



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diversity sampling should involve spatiotemporally and taxonomically broad sampling to characterize the community of mosquitoes present at a site. Although a combined sampling plan driven by considerations of pathogen-related sampling would not be suitable for robust sampling of mosquito abundance and diversity, a plan based upon the sampling priorities of abundance and diversity sampling would also collect some of the mosquitoes needed for pathogen testing. The sampling design therefore represents a combination of the two sampling efforts into a unified approach (hereafter, mosquito sampling) that is driven largely by priorities of abundance and diversity sampling and that can be augmented as needed to meet requirements of pathogen-related sampling.

5.3.3 Priorities and Challenges for Rodent-Borne Pathogen Sampling

As with mosquito sampling, rodent-borne pathogen sampling occurs coincidentally with NEON small mammal abundance and diversity sampling (for additional details see the NEON TOS Science Design for Small Mammal Abundance and Diversity, AD[08]). The proposed sampling design for the former is structured in part by design priorities and considerations that are central to the latter (e.g., number and location of sampling plots within sites), referred to hereafter as small mammal sampling. The inherent variability of small mammal communities and populations across time and space, their relatively low densities in many locations (relative to ticks and mosquitoes), and the skill and experience required by field staff to consistently collect quality blood samples also pose challenges to the goal of acquiring sufficient sample sizes for generating pathogen prevalence estimates through time.



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6 SAMPLING DESIGN FOR VECTOR AND PATHOGEN SAMPLING

6.1 Sampling Design for Tick and Tick-Borne Pathogen Sampling

Sampling for ticks and tick-borne pathogens (hereafter, tick sampling) will broadly target hard ticks (family Ixodidae). Six species are of particular interest for tick-borne pathogen testing: *Ixodes scapularis* (black-legged tick), *I. pacificus* (western black-legged tick), *Amblyomma americanum* (lone star tick), *A. maculatum* (Gulf coast tick), *Dermacentor andersoni* (Rocky mountain wood tick), and *D. variabilis* (American dog tick). Collectively, ticks of these species transmit a large and taxonomically-diverse suite of pathogens, many of which are zoonotic and of public health concern (<http://www.cdc.gov/ticks/diseases/>, accessed on May 1, 2014, Spach et al. 1993). Ticks of other species collected through this sampling will be not be tested for pathogens but will be archived and available to the scientific community. Pathogen testing will broadly target bacteria, including *Anaplasma phagocytophilum*, *Borrelia burgdorferi*, *Borrelia lonestari*, *Borrelia mayonii*, *Borrelia miyamotoi*, *Ehrlichia chaffeensis*, *Ehrlichia ewingii*, *Francisella tularensis*, and *Rickettsia rickettsia*, as well as the eukaryote *Babesia microti*.

Additional details of the implementation of NEON tick sampling can be found in TOS Protocol and Procedure: Tick and Tick-borne Pathogen Sampling (AD[03]).

6.1.1 Sampling Methods

6.1.1.1 Sample Collection in the Field

Ticks will be sampled using the dragging method, which is arguably the most commonly used method to sample ticks and is particularly effective for ticks of species that exhibit questing behavior (i.e., sit-and-wait) (Milne 1943, Falco and Fish 1992). Among tick sampling methods, dragging also most closely approximates the human risk of picking up hard ticks from the environment. During drag sampling, a cloth of standardized size is pulled along the ground at a slow pace. The cloth is periodically examined for attached ticks, which are typically removed with forceps or tape. Ticks that become attached to the clothing of sampling personnel during drag sampling can also be collected when the drag cloth is examined. The distance covered during a drag is generally standardized and recorded for use in calculating tick density. In instances where thick vegetation prevents continuous drag sampling, the flagging method can be used as an alternative or in conjunction with dragging (Ginsberg and Ewing 1989). Flag sampling essentially involves using the drag cloth held in the hand and slowly waved over or underneath vegetation rather than pulled along the ground (Rulison et al. 2013).

While their use is not currently planned, two additional methods of tick sampling could be considered to supplement drag sampling as resources permit. Collection of ticks using dry-ice baited CO₂ traps is well suited to sample ticks of species such as *A. americanum* that exhibit active hunting behavior and locate hosts by following carbon dioxide plumes (Garcia 1962, Falco and Fish 1989, Kinzer et al. 1990). Ticks attracted to a CO₂ trap are captured on tape attached to the edge of the trap's base platform. A drag/flag cloth can be used to collect additional ticks from the ground and vegetation in the vicinity of



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the trap. If used, CO₂ traps would use a vented 1.9L insulated cooler containing approximately 1.5kg of dry ice and be deployed in the center of each sampling plot for 24 hours following the completion of drag and/or flag sampling during a sampling event.

The removal of ticks from vertebrate hosts is another commonly used method of sampling ticks (Luckhart et al. 1992, Clark et al. 1998, Kollars et al. 2000). Results of sampling using this method can provide important insights into the host associations of ticks and by extension, the transmission cycles of associated pathogens (Clark et al. 2001, Eisen et al. 2004). Deer are final hosts for ticks of many species, and researchers often remove ticks from deer killed by sportsmen (i.e., at hunter check stations) or from road kill (Luckhart et al. 1992, Kollars et al. 2000). Because NEON will not be sampling deer, this is not a methodological option. Although NEON will conduct regular sampling of small mammals, including species that are important hosts for ticks and play significant roles as reservoirs of many tick-borne pathogens (Donahue et al. 1987, Ostfeld et al. 1996), removal of ticks from captured small mammals is not currently planned. This decision is driven by the need to limit small mammal handling time in the face of other prioritized data and sample collection requirements. Design-related details on tick sampling from rodents can be found in the rodent-borne pathogen section of this document. Any associated protocol details will be captured in TOS Protocol and Procedure: Small Mammal Sampling (AD[05]).

In 2012, tick sampling using the drag method was prototyped at all three sites in NEON Domain 03: Ordway Swisher Biological Station (core), Jones Ecological Research Center (gradient), and Disney Wilderness Preserve (gradient). Results demonstrate the utility of drag sampling to elucidate spatial variation in tick abundance within and among sites. The lone star tick (*A. americanum*) is known to be established throughout central and northern Florida and southern Georgia and often reaches levels of extremely high abundance where it occurs (Springer et al. 2014). Based on this regional information it could be assumed that individuals of this species would be abundant at all three NEON sites in Domain 03. While results of tick sampling at the core site in June 2012 confirmed a high abundance of ticks, concurrent sampling of nearly 16,000m² (collectively) at the two associated gradient sites resulted in the collection of only five ticks total (**Table 1**). These results may indicate that local conditions at these sites are not conducive to the maintenance of large tick populations. Regular vegetation burning at both gradient sites likely contributed to this outcome, demonstrating the impact of land-use practices on tick demography. Results of iterative sampling that will occur during Observatory observations could shed light on the temporal duration of such demographic effects (i.e., rates of recovery of tick populations after controlled burns at a site).

Ticks were found in all three vegetation types sampled at Ordway Swisher Biological Station, where results provided evidence supporting spatial clustering of ticks among vegetation types. Ticks were one to two orders of magnitude more abundant in deciduous closed tree canopy habitats compared to evergreen open tree canopy and perennial graminoid grassland habitats (**Table 1**). This pattern was apparent in spite of the greater sampling effort (number of plots and average distance covered during drag and walking sampling) associated with the latter two vegetation types. Further, although sample



sizes were low, ticks were present in every plot sampled within deciduous closed tree canopy plots. In contrast, ticks were absent in 85% and 60% of sampled plots located in evergreen open tree canopy and perennial graminoid grassland habitats, respectively.

Table 1. Results of year-one spatial sampling of ticks in NEON Domain 03, spring and summer 2012.

Site	Vegetation type	# plots sampled	Tick abundance per plot (mean±stdev)	Total # ticks* collected	Max # ticks* per plot	# plots with no ticks*	Total area sampled (m ²)
Disney Wilderness Preserve	Evergreen open tree canopy	9	0.3±0.95	3	3	8	2,700
	Mixed evergreen / deciduous open tree canopy	1	0.0±0.0	0		1	300
	Perennial graminoid grassland	7	0.0±0.0	0		7	2,100
	Total	17	0.2±0.7	3	3	16	5,100
Jones Ecological Research Center	Annual graminoid or forb	6	0.0±0.0	0		6	1,958
	Evergreen open tree canopy	5	0.4±0.89	2	2	4	5,973
	Mixed evergreen / deciduous open tree canopy	4	0.0±0.0	0		4	1,481
	Perennial graminoid grassland	4	0.0±0.0	0		4	1,401
	Total	19	0.1±0.5	2	2	18	10,813
Ordway Swisher Biological Station	Deciduous closed tree canopy	3	20.0±17.1	60	39	0	811
	Evergreen open tree canopy	14	0.2±0.6	3	2	12	6,156
	Perennial graminoid grassland	5	1.2±1.8	6	4	3	1,615
	Total	22	3.1±8.7	69	39	15	8,582
Overall	58	1.2±5.4	74	39	49	24,495	

* only nymphal and adult ticks considered

6.1.1.2 Sampling Event

An event of tick sampling at a site will involve drag sampling around the perimeter of each of six 1600m² square sampling plots per site. During a sampling event, a 1m² piece of white, cotton flannel cloth will be pulled along the ground at a slow pace and examined at five- to ten-meter intervals along the drag path (**Figure 3**). Ticks found attached to the drag cloth and field staff clothing will be removed using forceps (nymphal and adult ticks) or masking tape, packing tape, or reusable lint rollers (larval ticks, Savage et al. 2013). The distance of each drag will be recorded for calculations of tick density. Flagging will be used along any portions of the sampling path where thick vegetation prevents continuous drag sampling.

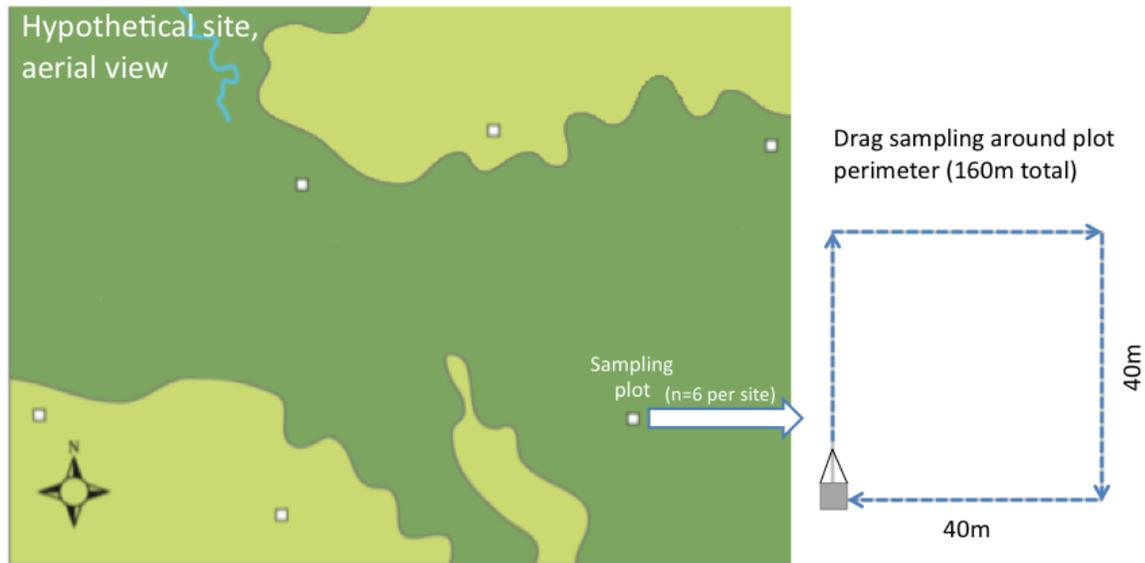


Figure 3. Schematic illustrating an event of tick sampling.

In the field, collected nymphal and adult ticks will be transferred into labeled vials containing 95% ethanol. Larvae collected using tape or reusable lint rollers will be submerged in ethanol to separate (Savage et al. 2013). Samples will be transported on ice in portable coolers to a NEON domain lab. For each sampling event/plot combination, collected ticks will be enumerated by life stage by field staff and then transferred into cold storage at $<4^{\circ}\text{C}$ (ideally -20°C).

6.1.1.3 Taxonomic Identification of Samples

NEON field staff will not perform any taxonomic identification of collected ticks. Collected ticks will be sent to one or more external facilities for taxonomic identification and pathogen testing. Nymphal and adult ticks will be identified to species based on visual examination of external morphology (e.g., Cooley and Kohls 1944, Cooley and Kohls 1945, Keirans and Litwak 1989) and enumerated by species and life stage for each sampling event/plot combination (e.g., number of nymphs of species A collected at plot B within site C during sampling event D). If resources permit, uncertain identifications may be verified as necessary or desired through examination by a secondary ID facility or using genetic methods such as DNA barcoding (Hebert et al. 2003, Pons et al. 2006, Mukherjee et al. 2014). Larval ticks will not be identified but will be counted and archived.

6.1.1.4 Pathogen Testing

Following identification, ticks will be combined by species, life stage, site, and event (e.g., all nymphs of species A collected at site C during sampling event D) for pathogen testing. A subset of species-identified nymphal ticks will be tested for infection by pathogens. Nymphs are preferable for testing since adult ticks are generally not contributors to the cycle of transmission and humans are more likely to be infected with a pathogen from a nymphal tick than an adult tick (Levi et al. 2015). A minimum of 10 and maximum of 100 nymphal ticks per species will be analyzed for each sampling event. Actual sample sizes



for pathogen testing will depend on collection success and analytical costs. Ticks will be tested individually using methods that allow for the detection of a wide range of pathogens including individuals in the genera *Anaplasma*, *Borrelia*, *Ehrlichia*, *Francisella*, and *Rickettsia*.

Species-identified but untested nymphal and adult-stage ticks, all unidentified larval, nymphal and adult-stage ticks, and products generated during pathogen testing (e.g., genomic extractions, PCR products) will be archived in existing facilities. Archiving plans, including details on accessing samples for private investigation, are detailed in the NEON Bioarchive Concept of Operations (AD[09]).

6.1.2 Spatial Distribution of Sampling

As described in the general introduction and in the NEON TOS Science Design for Spatial Sampling (AD[02]), a stratified random approach will be used to select six tick sampling plots located within the dominant vegetation types ($\geq 5\%$ total cover) at each site. The number of plots per type will be proportional to the percent cover of each type. Once a set of long-term sampling plots is selected, NEON will maintain a fixed plot design, sampling those plots irrespective of changes in the presence or abundance of ticks and/or associated pathogens.

Site-specific deviations

The stratified-random design is a key feature of the Terrestrial Observation Sampling component of the Observatory. However, difficulties in sampling ticks using the drag/flag method have prevented standardized plot establishment at the Guanica site (GUAN) in Puerto Rico. Namely, the density of vegetation at most of the randomly selected locations would not allow a drag cloth or flag to remain intact. Alternative plot establishment guidelines were therefore approved for use at GUAN; these alternative plot establishment rules are non-random and represent transect paths within the TOS sampling boundary where tick sampling is feasible, in accordance with the following guidelines.

1. Required: A minimum area of 80 m² (with an ideal target of 160 m²) can be sampled using either the flagging or dragging method. If necessary, the area may consist of multiple segments, each preferably within 10 meters of another segment.
2. Strongly preferred: The majority (50% or more) of the area is located within:
 - 300 m from the center of a mammal plot,
 - 300 m from the center of a bird plot, OR
 - 300 m from the center of a Distributed Base Plot

Ideally the area will be in range of all three plot types, but the order reflects priority (mammals are first preference). Ideally both this criterion AND the Distance from other TOS plots criterion will be met, but if this is not possible, this criterion takes higher priority.

3. Preferred: Does not cross roads or frequently-used paths to other plots. If no other options are available, plots may be located linearly along the edge of a dirt road or path, where vegetation or leaf litter can be sampled.



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Other than these changes in plot establishment, the tick sampling process remains the same. This includes the fixed nature of these locations until sampling is no longer possible. The efficacy of this alternative sampling method will be evaluated after several years of data collection to determine whether other sampling options might be preferable.

6.1.3 Temporal Distribution of Sampling

The goals of the NEON sampling are to characterize the site-specific species composition and relative abundance of ticks of concern to public health and to capture a sufficient number of such ticks (if present) to adequately characterize the presence and absence of pathogens in the resident tick community. The activity of Ixodid ticks is known to vary significantly throughout the year in temperate zones in response to varying environmental conditions (e.g., temperature and moisture; Gray 1998). Repeated sampling throughout a sampling window is therefore advisable to increase the probability of capturing the peak activity times of ticks and therefore increase sample sizes.

Sampling Window

Sampling will occur during the growing season when temperatures and tick activity are increased. During this period, a given sampling event will only be performed if the high temperature on the day prior to planned sampling was $>0^{\circ}\text{C}$ and the mean high temperature in the five days prior to planned sampling was $>7^{\circ}\text{C}$. These temperatures represent conservative thresholds below which ticks are generally not active (Duffy and Campbell 1994, Clark 1995, Vail and Smith 1998). Sampling will be postponed whenever the ground is wet (e.g., shortly after a rain event or when dew is heavy) and avoided during the hottest and/or driest periods of the day (relative to typical temperatures at any given site).

Adjustments to the Sampling Window

Prior to 2016, tick sampling was scheduled from March through December at all sites, if the moisture and/or temperature thresholds described above were met. However, challenges in staffing and resource availability prompted an effort to constrain the sampling window in an ecologically-informed manner. For this effort, NASA's MODIS (Moderate Resolution Imaging Spectroradiometer) EVI (Enhanced Vegetation Index) phenology data from the most recently available decade (2001-2009; <https://modis.gsfc.nasa.gov/data/dataproduct/mod13.php>) were used to define the growing season on a site-specific basis. For each site, the average Julian dates of increasing green-up and the mid-point between decreasing greenness and minimum greenness are used to define the beginning and end of the sampling season, respectively (AD[03]). On average, these criteria yield an average of 6 bouts per site (versus the 10 bouts per site that might be implemented during a March to December timeframe) at all sites except those in Domains 04 and 20. These domains feature tropical habitats with greening data suggestive that year-round sampling could be appropriate. Given budget constraints coupled with the lack of tick captures to date, a 4-bout window (6 months; sampling occurring every 6 weeks) is being



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used at Domain 04. Tick collections are not permitted in Domain 20. Sampling dates will be updated using the most recent MODIS data every 5-10 years.

In 2017, field operations staff implemented the revised sampling windows, while continuing to monitor temperatures and delay deployment if the mean high temperature in the five days prior to planned sampling remains persistently below 7°C or below 0°C the day before sampling. Likewise, sampling may conclude earlier than the MODIS-based dates if temperatures fall persistently below these thresholds. As a safeguard against possible discrepancies between MODIS readings and on-the-ground greening and temperatures, field staff notify NEON Science if the timing suggested by temperature thresholds would result in initiating or completing sampling more than one month off from the MODIS-derived dates. This information, along with the data collected at each site, may be used to fine tune sampling windows.

Sampling Frequency

Sampling at each site will begin at a frequency of one sampling event every six weeks. This frequency will be maintained until ticks are collected during a sampling event, a threshold that will trigger an increase in sampling frequency to one event every three weeks. This frequency will be maintained at the site for one year after a documented tick capture. If ticks are not captured for one calendar year, the site will revert to sampling every six weeks until ticks are captured at the site. In this way, it is expected that a small number of sites will oscillate in their sampling frequency if there is very occasional tick presence at the site.

From 2014 to 2017, the capture of a single tick triggered more frequent ‘high-intensity’ sampling every three weeks. In 2018, data from 2016 and 2017 were used to evaluate this threshold (Appendix A). Sites with very low captures (i.e., <20 per year) remained consistently low through time, suggesting that a higher threshold to trigger high intensity sampling would not result in missing a period of high captures. The threshold to trigger high intensity sampling was therefore increased to collection of five or more ticks as a cost saving measure, with the caveat that sampling results would continue to be evaluated and the threshold revised downward, if needed.

6.1.3.1 Power Analyses Evaluating Sampling Frequencies

Evaluation of power to detect prevalence trends

For power analyses to evaluate the ability of sampling and testing to detect long-term trends in tick-borne pathogen infection prevalence, year-zero prevalence values were set to 5% (rare pathogen) or 15% (common pathogen) based on results of studies of *B. burgdorferi* infection in *I. scapularis* populations (Piesman et al. 1986, Ginsberg et al. 2004, Ogden et al. 2007, Diuk-Wasser et al. 2012). Sampling event frequencies of one, two, three and four times per year, as well as every three or six weeks, were considered. Given plans to test between 10 and 100 ticks per species per sampling event, a conservative sample size threshold of 40 ticks tested for pathogens per sampling event was used. Hence, a sampling frequency was deemed adequate if 40 or fewer tested ticks per event were sufficient to detect a pathogen prevalence trend of a given magnitude. Note that while these calculations assume



that sample sizes for pathogen testing will be dictated mostly by analytical costs and statistical considerations, ticks may be uncommon at many sites and/or during some periods, and the number available to be tested for pathogens may fall below 40 per event.

When power analyses were run with a temporal autocorrelation term associated with the prevalence data, the proposed design was only capable of detecting annual prevalence increases of $\leq 1.0\%$ for a common pathogen at a core site (i.e., over 25 years). In this scenario, the design can detect an annual increase of 0.75% with higher confidence ($\alpha=0.05$, power=0.9) (Figure 4A). In the absence of temporal autocorrelation in the data, the design can detect an annual increase of 0.25% with high confidence at sampling frequencies of one event every three and six weeks. At these sampling frequencies, the design can detect an annual increase of 0.50% for a rare pathogen at a core site with higher confidence (Figure 4B), and an annual increase of 0.25% with lower confidence ($\alpha=0.1$, power=0.8) but only at a sampling frequency of one event every three weeks. The power of the design was weaker at gradient sites (i.e., over 10 years). There, for common pathogens and at a sampling frequency of one event every six weeks, annual increases of 0.75% (Figure 4C) and 1.0% can be detected with lower and higher confidence, respectively. For rare pathogens, an annual increase of 1.0% can only be detected with lower confidence at a sampling frequency of one event every three weeks (Figure 4D).

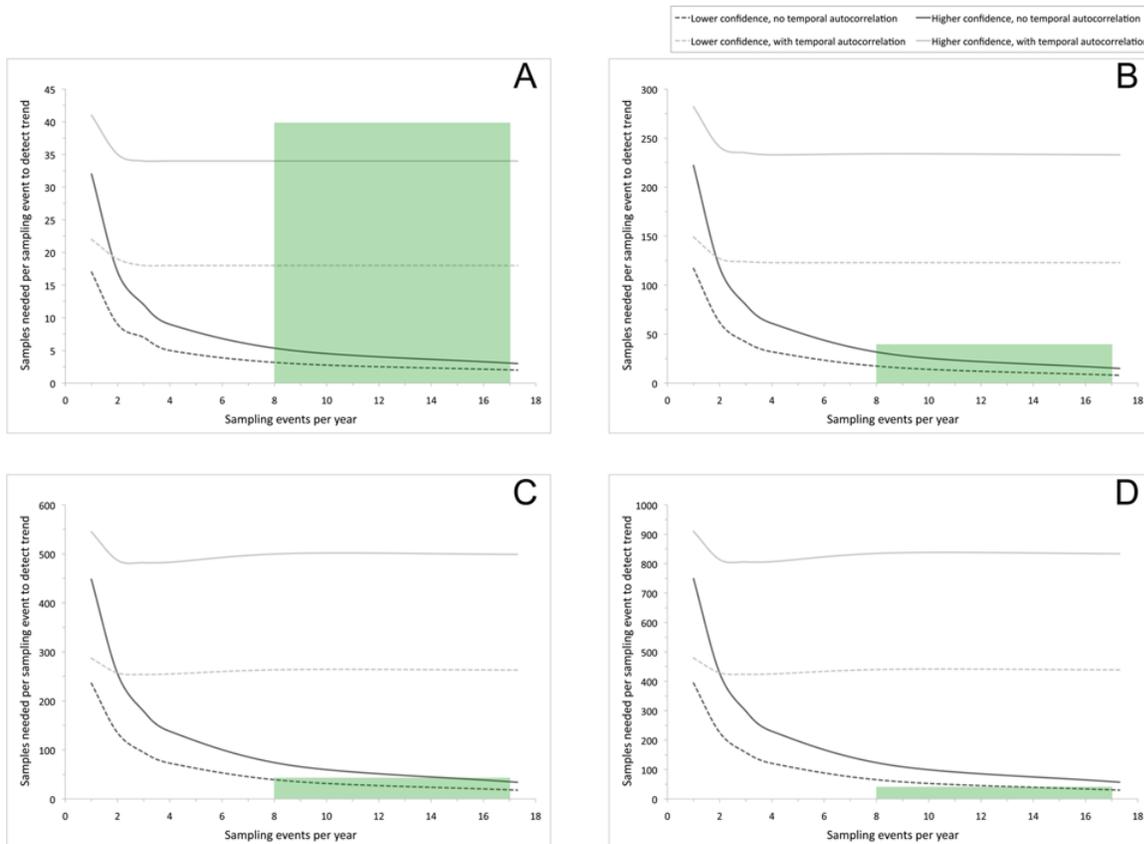


Figure 4. Results of power analyses for a common tick-borne pathogen (prevalence=15%) at a core (i.e., 25 years, A) and a gradient (i.e., 10 years, B) site and a rare tick-borne pathogen (prevalence=5%) at a core (C) and gradient (D) site.



(D) site. Each analysis includes results for all combinations of low ($\alpha=0.1$, power=0.8) and high ($\alpha=0.05$, power=0.9) confidence and with and without a temporal autocorrelation term (e^{-1} at a half year). Green boxes indicate levels of sampling effort (event frequency and number samples/event to be tested for pathogens) specified in the design.

Evaluation of power to detect pathogen(s)

The design has a ~100% probability of detecting a common pathogen and an 87% probability of detecting a rare pathogen when 40 or more ticks per species are tested per sampling event (**Figure 5A**). For very rare pathogens (1% prevalence) the probability is 33%. When analytical sampling effort is reduced by 50% (20 ticks per species tested per sampling event), these probabilities fall to 96%, 64%, and 18%, respectively. To achieve a 50% probability of detecting a pathogen, four, 14, and 68 ticks must be tested under scenarios when the pathogen is common, rare, or very rare, respectively (**Figure 5B**). These sample sizes must be increased to 10, 31, and 160 ticks to achieve an 80% detection probability.

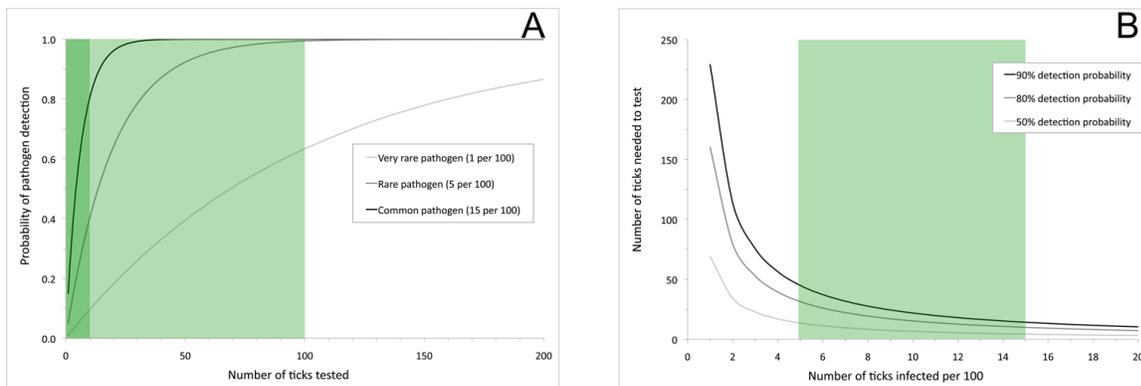


Figure 5. (A) Relationship between the number of samples tested for pathogens and the probability of detecting tick-borne pathogens. Green boxes indicated upper and lower estimates of number of samples to be tested per sampling event. (B) Minimum number of samples required for specified detection probabilities at various levels of infection prevalence for tick-borne pathogens. Green box indicates estimated prevalence of rare and common pathogens at NEON sites.

6.1.4 Logistics and Adaptability

To promote high temporal resolution time series data in the face of potential financial and logistic constraints, the current sampling strategy prioritizes intensive within-site sampling. In the event that field sampling effort must be reduced, a relatively high level of sampling effort will be maintained at NEON core sites, and effort reduced at gradient sites, to preserve the longer time series data associated with the former. If analytical cost savings are required, field sampling efforts specified in the sampling designs (e.g., frequency of sampling events, number of plots per site) will be maintained and cost savings realized through reductions in the number of samples that are tested for pathogens. Foregoing the testing of all samples collected during selected sampling events, rather than reducing the number of samples tested for every event, will facilitate data comparability by maintaining more consistent levels of analytical sampling effort and associated uncertainty through time. Remaining samples (i.e., untested or residual) can be archived for processing at a later date, either by NEON or other members of the research community.



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6.2 Sampling Design for Mosquito-Borne Pathogen Sampling

Mosquito sampling will broadly target all members of the family Culicidae. For mosquito-borne pathogen testing, samples will be screened for infection by arboviruses (Calisher 1994). Mosquitoes in the genera *Aedes* and *Culex* are of particular interest because they collectively transmit a variety of viral pathogens (Moore et al. 1993, Turell et al. 2005). Mosquitoes of other species will be tested for pathogens as resources permit and sample sizes warrant. Female mosquitoes, pooled by species, will be tested for the presence of alphaviruses, bunyaviruses, and flaviviruses using PCR-based and/or Vero cell culture methods. Some or all virus-positive pools will be subjected to additional pathogen-specific tests to identify the pathogen(s) present within them. Additional details of the design and implementation of NEON mosquito sampling can be found in the NEON TOS Science Design for Mosquito Abundance, Diversity and Phenology (AD[07]) and in TOS Protocol and Procedure: Mosquito Sampling (AD[04]).

6.2.1 Sampling Methods

6.2.1.1 Sample Collection in the Field

Mosquito sampling will be conducted using CDC miniature CO₂ light traps (Sudia and Chamberlain 1962). Among commonly used mosquito traps this type arguably collects the greatest diversity of mosquitoes across a broad range of environmental conditions and is regularly deployed as part of arbovirus surveillance (Service 1993). As such, use of this trap for NEON sampling should facilitate the integration of NEON mosquito data with similar data collected by many public health and mosquito control agencies. CDC miniature CO₂ light traps attract mosquitoes through the emission of light and release of carbon dioxide (e.g., from sublimating dry ice or compressed gas cylinders), the latter being a component of vertebrate exhalation that female mosquitoes use to locate hosts. Mosquitoes attracted to a trap are drawn into a specimen catch cup by the trap's fan.

6.2.1.2 Sampling Event

An event of mosquito sampling will involve the deployment of one dry-ice baited CDC miniature CO₂ light trap at each of 10 sampling plots per site. During a sampling event, each trap will be deployed continuously for roughly 40 hours beginning at dusk on the first day. Traps will be checked (collected mosquitoes retrieved, dry ice replenished) at dawn following both nights of deployment and at dusk on the second day. This results in each sampling event being split into three trapping periods: two trap nights and the intervening day. In 2018, the second night of sampling was dropped to reduce sampling costs. Data will be evaluated after this sampling year to assess whether the change resulted in total captures falling below the minimum sample size of 1,000 *Aedes* and *Culex* females for pathogen analyses at sites that have historically exceeded the threshold. Traps will be hung at a height of roughly two meters and baited with approximately 1.5kg of dry ice during each trapping period. Light bulbs will be turned off during deployment to reduce bycatch and conserve battery life. Additional details about mosquito sampling can be found in the NEON TOS Science Design for Mosquito Abundance, Diversity, and Phenology (AD[07]).



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Following their collection from traps, sampling personnel will transport catch cups containing mosquitoes to a NEON domain lab in portable coolers containing dry ice. At the domain lab, mosquitoes will be flash frozen and transferred into labeled vials. Sample vials will be stored at -80°C at the domain lab until they are sent to one or more external facilities for mosquito taxonomic identification and pathogen testing.

6.2.1.3 Taxonomic Identification of Samples

Taxonomic identification will be based on visual examination of external morphology (e.g., Darsie and Ward 1981) with some confirmation using DNA barcoding (Gibson et al. 2012). From among mosquitoes collected during each trapping period, NEON will either identify a set proportion of the total catch or a fixed number of mosquitoes. This decision will ultimately be based on catch rates observed during the first few years of sampling. The initial plan involves the latter approach: up to 200 mosquitoes per trapping period will be identified and enumerated by species and sex (e.g., number of females of species A collected at plot B within site C during night one of sampling event D). When more than 200 mosquitoes are collected in a trapping period, a representative subsample of ~200 individuals will be identified. All processing will be conducted in a manner that maintains the cold chain and prevents freeze/thaw cycles that could compromise the quality of samples for pathogen testing. Following identification, mosquitoes will be combined by species, sex, site, and event (e.g., all females of species A collected at site C during sampling event D) for pathogen testing. Any remaining unidentified mosquitoes will be bulked at the site/event level. See the NEON TOS Science Design for Mosquito Abundance, Diversity, and Phenology (AD[07]) for more details on taxonomic identification, DNA barcoding, preservation of pinned NEON mosquito samples, and handling and archive of bycatch.

6.2.1.4 Pathogen Testing

A subset of the species-identified female mosquitoes collected during each event of sampling will be tested for pathogens. Mosquitoes will be tested in pools of 10-50 individuals grouped by species at the site level (catches combined across all trapping periods and plots sampled during the event). Actual sample sizes for pathogen testing will depend on collection success and analytical costs. Because the prevalence of arboviruses in mosquitoes is generally very low (e.g., 1-5 infected individuals per 1,000 individuals, Andreadis et al. 2004, Gu and Novak 2004, Gu et al. 2008, Kwan et al. 2010), it is desirable to maximize the number of mosquitoes tested for pathogens. In instances where only a portion of mosquitoes collected during a sampling event are identified, NEON may pursue the identification and removal of additional individuals of target vector species from the unidentified bulk lot to increase analytical sample sizes for pathogen testing. This would occur at the end of the sampling season. Target species will include known vectors of arboviral pathogens within the NEON purview divided into 3 tiers (Moore et al. 1993, Turell et al. 2005):

1. Species in the genus *Aedes* including *Ae. aegypti*, *Ae. albopictus*, and *Ae. triseriatus*. These species collectively transmit dengue viruses and La Crosse encephalitis virus.



2. Species in the genus *Culex* including *Cx. pipiens*, *Cx. p. quinquefasciatus*, *Cx. tarsalis*, *Cx. salinarius*, *Cx. nigripalpus*, and *Cx. restuans*. These species are important vectors of West Nile virus and St. Louis encephalitis virus.
3. Mosquitoes of other taxa including *Culiseta melanura* and *Coquillettidia perturbans* (vectors of eastern equine encephalitis virus) or confirmed vectors of arboviral pathogens within NEON domains.

For any target species, a set of up to 1,000 individual mosquitoes per species per site per year will be slated for pathogen testing of arboviruses within the families Bunyaviridae, Alphaviridae, and Flaviviridae. This quantity of individuals is informed by the power analyses described in section 'Power Analyses Evaluating Sampling Frequencies' (illustrated in **Figure 4**). The highest priority for analysis is given to Tier 1 species (esp. *Aedes aegypti* and *Aedes albopictus*), followed by Tier 2 species and then Tier 3 species. To be eligible for pathogen testing, a species must have a minimum of 100 (Tier 1 species) or 200 (Tiers 2 & 3 species) individuals present at a site in a year that are available for testing. Pools of 10 to 50 individuals will be created from aggregations of female mosquitoes from the same species and bout of collection. All identified mosquito samples are either used for pathogen testing, are pinned as voucher specimens, or archived at -80 C.

After selection, each mosquito pool will first be tested for the presence of alphaviruses, bunyaviruses, and flaviviruses. This can be accomplished using various methods (or combinations thereof) including Vero cell screening, RT-PCR using specific or general (i.e., broadly-reactive, family specific) primers, and melt curve assays for viral RNA (Earley et al. 1967, Kuno et al. 1996, Kuno 1998, Lanciotti et al. 2000, Sanchez-Seco et al. 2001, Nasci et al. 2002, Naze et al. 2009). Because West Nile virus and dengue viruses are flaviviral pathogens of particular interest, any flavivirus-positive pools will be further assayed with pathogen-specific tests to identify the particular pathogen(s) present within them. Alphavirus- and bunyavirus-positive pools will be subjected to similar secondary tests as resources allow.

6.2.1.5 Archive

A small number of species-identified but untested mosquitoes will be used to develop a mosquito DNA barcode library and pointed for archiving. Remaining species-identified and untested mosquitoes, all or a subset of unidentified mosquitoes, and products generated by mosquito-borne pathogen testing (e.g., genomic extractions, RT-PCR products) will be archived in vials in existing facilities.

6.2.2 Spatial Distribution of Sampling

As described in the general introduction and in the NEON TOS Science Design for Spatial Sampling (AD[02]), a stratified random approach will be used to select 10 mosquito sampling plots within the dominant vegetation types ($\geq 5\%$ total cover) at each site. The number of plots per type will be proportional to the percent cover of each type. Plot locations will additionally be constrained to fall within 30m of roads because of a need to reduce sampling-associated travel times to and from plots. Once a set of long-term field season sampling plots is selected, NEON will maintain a fixed plots design,



sampling those plots irrespective of changes in the presence or abundance of mosquitoes and/or associated pathogens.

Evaluation of power to detect prevalence trends

In the absence of temporal autocorrelation, the design can detect an annual prevalence increase of 2.0% for a common pathogen at a core site (i.e., over 25 years) with higher confidence ($\alpha=0.05$, power=0.9) (Figure 6A), and an annual increase of 1.5% with lower confidence ($\alpha=0.1$, power=0.8). Assuming one sampling event every two weeks at core sites, a minimum of eight and 30 pools must be tested per event to detect annual prevalence increases of 1.0% and 0.5%, respectively, with lower confidence. When the pathogen is rare at a core site an annual increase of 3.0% can be detected with lower confidence. At gradient sites (i.e., over 10 years) the design cannot detect an annual prevalence increase of $\leq 5.0\%$ when the pathogen is common or rare (Figure 6B).

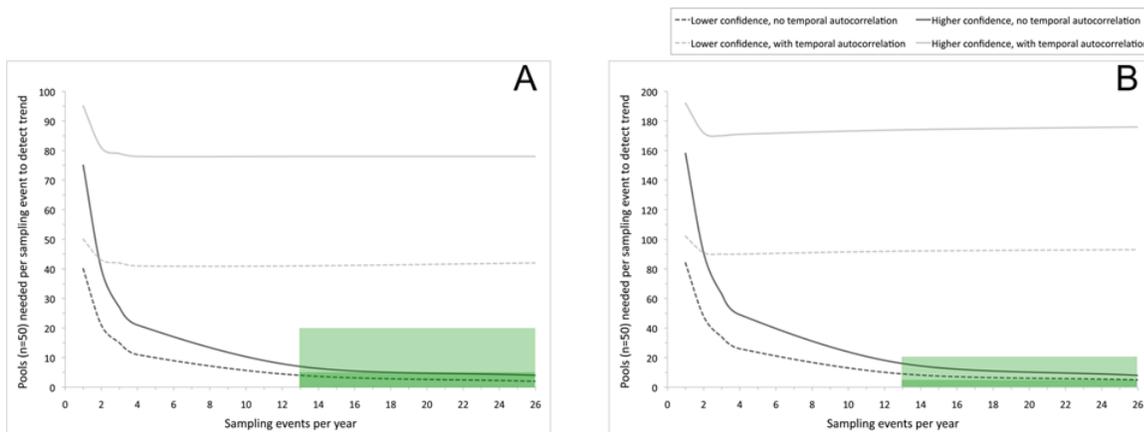


Figure 6. Results of power analyses for common mosquito-borne pathogens (prevalence=0.5%) at a core (i.e., 25 years, A) and a gradient (i.e., 10 years, B) site. Each analysis includes results for all combinations of low ($\alpha=0.1$, power=0.8) and high ($\alpha=0.05$, power=0.9) confidence and with and without a temporal autocorrelation term (e-1 at a half year). Green boxes indicate levels of sampling effort (event frequency and number samples/event to be tested for pathogens) specified in the design.

6.2.3 Temporal Distribution of Sampling

Sampling will be conducted at a frequency of one sampling event every two weeks at core sites and every four weeks at gradient sites. These frequencies will be maintained for the lifespan of each site irrespective of sampling success. Sampling will occur year-round as long as mosquitoes are being collected (see below for description of off season sampling), but a given sampling event will only be performed if the mean daily high temperature for the five days prior to the first day of the event is above the specified temperature threshold. Prior to 2017, this threshold was set at $\geq 4^{\circ}\text{C}$, as this temperature is a conservative threshold below which mosquitoes are generally not active (e.g., Bailey et al. 1965, Corbet and Danks 1973, Almeida and Gorla 1995, Ciota et al. 2011). Given the field staff reports of numerous zero-capture bouts, this threshold was re-evaluated using capture data collected to date in

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2017 and increased to $\geq 10^{\circ}\text{C}$. See the NEON TOS Science Design for Mosquito Abundance, Diversity, and Phenology (AD[07]) for additional details on the analysis and justification for the increase.

Mosquito sampling at some higher latitude NEON sites can be discontinued for part of each year when environmental conditions are unfavorable for mosquito activity. At these sites, a program of off-season sampling will be implemented to empirically detect the onset and conclusion of annual mosquito activity cycles. Within a NEON domain, off-season sampling will commence following three consecutive zero-catch sampling events at the core site. Off-season sampling will involve weekly deployment of one CDC miniature CO₂ light trap at each of three sampling plots for a single night of trapping at the core site. Sampling will transition back to the regular season plan (10 sampling plots with events every two or four weeks) following the collection of at least one mosquito during an off-season sampling event. See the NEON TOS Science Design for Mosquito Abundance, Diversity, and Phenology (AD[07]) for additional details on off-season mosquito sampling.

6.2.3.1 Power Analyses Evaluating Sampling Frequencies

Power analyses associated with mosquito-borne pathogen sampling are complicated by the tremendous spatiotemporal variability in mosquito abundance and associated difficulty in estimating the catch size for a “typical” sampling event. Based on current cost and budget estimates we anticipate sufficient funding for the taxonomic identification of up to 600 mosquitoes per trap per sampling event (up to 200 in each of the three trapping periods per event). Given the plan to deploy one trap at each of ten sampling plots at a site during an event, this yields a maximum of 6,000 taxonomically identified mosquitoes per site/event combination. Because catches during the diurnal portion of sampling events are likely to be relatively small, and nocturnal catches will not consistently exceed 200 mosquitoes per trap/trapping event at most sites, we assume this maximum can be reduced to a more conservative value of 2,000, the majority of which will be females. This catch success seems reasonable at sites where mosquitoes are abundant but could regularly be lower by an order of magnitude or more at sites where they are rare. The number of resulting analytical pools of 50 that can be generated from these 200-2,000 mosquitoes will vary depending on the relative abundance of species at a site. For power analyses we assumed sample size thresholds of 15-20 pools per species/sampling event combination on the high end and 1-4 pools on the low end. Year-zero prevalence values were set to 0.1% (rare pathogen) and 0.5% (common pathogen) based on typical rates of arboviral infection in mosquitoes (Andreadis et al. 2004, Gu and Novak 2004, Gu et al. 2008, Kwan et al. 2010). In addition to evaluating the power associated with sampling event frequencies of every two and four weeks, frequencies of one, two, three and four times per year were evaluated to generate power curves.

Evaluation of power to detect pathogen(s)

Sampling has high power to detect the presence of pathogens at Observatory sites. The design has a >90% probability of detecting a common pathogen when 450 or more mosquitoes (nine or more pools of 50) are tested (**Figure 7A**). This probability is between 22% and 63% when analytical sample sizes are lower (between one and four pools of 50, respectively). Power is lower for rare pathogens: the detection



probability exceeds 50% when roughly 685 mosquitoes are tested, and probabilities are between 5% and 18% when analytical sample sizes are very low (one and four pools of 50, respectively). For reference, a minimum of 1,609 and 2,301 mosquitoes would need to be tested to detect a rare pathogen with 80% and 90% confidence, respectively (**Figure 7B**). For a common pathogen these values are 321 and 459. Fifty percent detection probabilities require the testing of 693 mosquitoes when the pathogen is rare and 138 mosquitoes when the pathogen is common. Detection would be more likely during epizootics when arboviral infection prevalence is higher (e.g., 20-25 infected mosquitoes per thousand). In these scenarios, 50%, 80% and 90% detection probabilities require testing a minimum of 34, 80, and 114 mosquitoes, respectively.

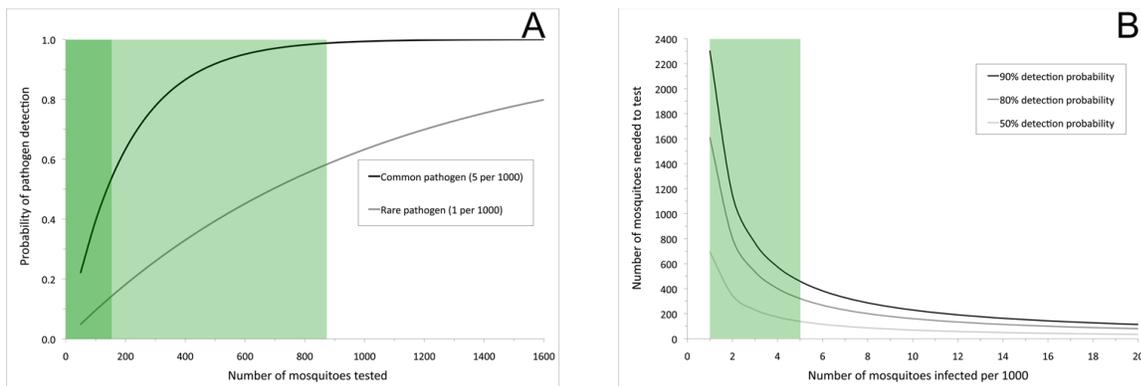


Figure 7. (A) Relationship between the number of samples tested for pathogens and the probability of detecting mosquito-borne pathogens. Green boxes indicated upper and lower estimates of number of samples to be tested per sampling event. (B) Minimum number of samples required for specified detection probabilities at various levels of infection prevalence for mosquito-borne pathogens. Green box indicates estimated prevalence of rare and common pathogens at NEON sites.

6.2.4 Logistics and Adaptability

In the event that sampling effort must be reduced, the sampling design (e.g., frequency of sampling events, number of plots per site) should be maintained, and cost savings realized through reductions in the number of samples that are tested for pathogens. Foregoing the testing of all samples collected during selected sampling events, rather than reducing the number of samples tested for every event, will facilitate data comparability by maintaining more consistent levels of analytical sampling effort and associated uncertainty through time. Remaining samples (i.e., untested or residual) can be archived for processing at a later date, either by NEON or through agreements with other members of the research community.

The CDC miniature CO₂ light trap has known sampling biases and limitations, at least two of which are relevant for mosquito-borne pathogen sampling. First, these traps are relatively ineffective at sampling the gravid or previously blood-fed mosquitoes that are the preferred targets for pathogen testing. Because gravid female mosquitoes are previously blood fed, their inclusion in testing pools enhances the likelihood of detecting pathogens when they are present at a site. Gravid traps represent an attractive



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supplement to CDC miniature CO₂ light traps because they target gravid mosquitoes through baiting with fetid water associated with oviposition sites (Reiter 1983). Gravid female mosquitoes of foul water-breeding species seeking a site to lay their eggs approach the trap and are sucked into a specimen catch cup by the trap's fan. While the use of gravid traps would increase mosquito catch rates (especially for gravid females) and the likelihood of detecting pathogens, the logistic challenges associated with standardizing and transporting the fetid water bait greatly complicate the use of gravid traps for NEON mosquito sampling. Additionally, the efficacy of these traps is often relatively low in rural or wildland settings where NEON sites are located. Another alternative and more easily standardized method involves resting box traps (Williams and Gingrich 2007). These resting shelters offer cool, shaded environments that many mosquitoes seek out during daylight hours (Burkett-Cadena et al. 2008). Mosquitoes alighting on an interior surface of a resting trap can be collected by vacuum aspiration or by a fan assembly integrated into the trap (Panella et al. 2011). For some mosquitoes, these traps may represent a relatively inexpensive and easily standardizable method of collecting gravid females (Komar et al. 1995). When catch rates are deemed insufficient, NEON will initially pursue deployment of additional CDC miniature CO₂ light traps to increase trapping success. The deployment of resting or gravid traps alongside CDC miniature CO₂ light traps may be considered as resources allow and as issues related to methodological standardization can be satisfactorily addressed.

The second notable limitation of the CDC miniature CO₂ light trap is that it does not effectively sample mosquitoes of certain species. Notable among these are species in the genus *Aedes* that are important vectors of multiple arboviruses including dengue viruses (Hoel et al. 2009). The BG sentinel trap represents an alternative trapping option for addressing this taxonomic sampling deficiency (Krockel et al. 2006, Pialoux et al. 2007, Meeraus et al. 2008). After several years of data collection at sites where *Ae. aegypti* and *Ae. albopictus* mosquitoes (the principle vectors of dengue viruses) are known or suspected to be present, NEON will evaluate whether sampling utilizing CDC miniature CO₂ light traps has adequately sampled these species. If capture rates are low, NEON will evaluate the feasibility of deploying BG sentinel traps alongside CDC miniature CO₂ light traps to collect these species for use in associated pathogen testing.

6.3 Sampling Design for Rodent-Borne Pathogen Sampling

Sampling for rodent-borne pathogens will primarily target rodents in the family Cricetidae. Associated species are often abundant and ecologically important members of rodent communities and individuals tend to be physiological tolerant of blood sample collection by commonly used methods. Species in the genus *Peromyscus*, especially *P. maniculatus* (deer mouse) and *P. leucopus* (white-footed mouse) are of particular interest since they are broadly distributed, often present at high abundance, and are known reservoirs for hantaviruses. All blood samples will be tested using enzyme-linked immunosorbent assay (ELISA) tests to detect antibodies reactive against hantaviruses and, if resources permit, against arenaviruses. Blood samples will be collected from individuals of other rodent taxa when sampling does not significantly increase their morbidity or mortality. The number of attached larval and nymphal ticks will also be recorded. As mentioned in the tick and tick-borne pathogen sampling design section,



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removal of ticks from captured small mammals is not currently planned. This decision reflects the need to limit handling time in the face of other prioritized data and sample collection requirements. Data on tick burdens will be used to estimate the additional handling time that would be required to remove ticks from sampled rodents. These estimates will allow the feasibility of incorporating tick collection into the NEON small mammal sampling protocol to be evaluated. Additional details of the design and implementation of NEON small mammal sampling can be found in the NEON TOS Science Design for Small Mammal Abundance and Diversity (AD[08]) and in TOS Protocol and Procedure: Small Mammal Sampling (AD[05]).

6.3.1 Sampling Methods

6.3.1.1 Sample Collection in the Field

Blood samples to be used for pathogen testing can be collected using a variety of methods. In selecting one or more methods for use by NEON, important considerations include taxonomic breadth of applicability, effects on sampled animals, need for associated anesthesia, volume of resulting samples, and required personal protective equipment (Mills et al. 1995). Methods vary in the degree to which they are restricted for use with animals of particular taxa due to anatomical incompatibility or physiological sensitivity. Related to this, some methods require the use of anesthesia, which increases handling time and may have deleterious health effects for individuals of particular species and/or under certain conditions (e.g., extreme temperatures or when the dosage of anesthetic is difficult to control) (Kosek et al. 1972). Any sampling-related increase in morbidity and mortality of sampled animals has the potential to bias data generated through NEON small mammal mark/recapture sampling and should be avoided. Finally, sampling methods differ in the volume of the blood that they generate because of variation in the size of and pressure in the blood vessel(s) involved. Given the desire to archive NEON samples for additional analyses by other members of the research community, larger sample volumes are preferable. Taken together, the ideal sampling method could be used on animals of a wide range of taxa, generate samples of relatively large volume, and have minimal deleterious impacts on sampled animals and risk for sampling personnel.

Based on these considerations, NEON initially conducted blood collection using either the retroorbital method or the submandibular method. The retroorbital method has been commonly used by field ecologists studying rodent/hantavirus interactions and is considered by many to be the preferred technique for blood collection (Joslin 2009, Auffray et al. 2011, Sikes et al. 2011). A microhematocrit tube is inserted behind one of the rodent's eyes and used to puncture the retroorbital sinus, a highly vascularized region at the back of the orbit (**Figure 8A**). The procedure can be applied humanely, yields a large volume of blood relative to other methods, and does not require the use of needles or lancets that may cause injury to animals and sampling personnel during handling. Field studies have shown that when used properly, the retroorbital method does not increase handling mortality or decrease recapture rates of sampled animals (Swann et al. 1997, Parmenter et al. 1998). While these results have been produced even without the use of anesthesia (Douglass et al. 2000), sampled animals are generally anesthetized via inhalation of isoflurane prior to blood collection. This precaution reduces the risks of



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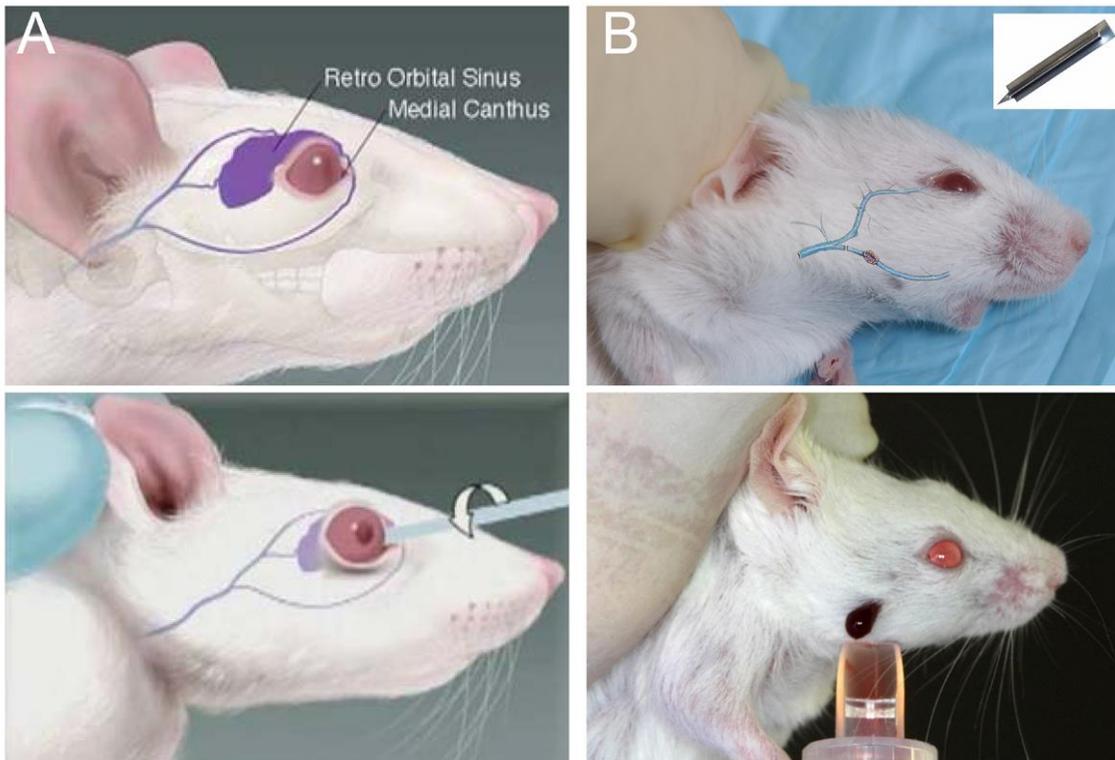
injury to animals and of bites or scratches to sampling personnel. Anesthetization was included in the NEON retroorbital bleeding protocol, as blood samples may be collected by sampling personnel with little if any prior experience handling small mammals.

Given concerns about deleterious effects of anesthesia on sampled animals, and the anticipation of growing resistance among institutional animal care and use committees to authorizing use of the retroorbital method, NEON also pursued use of the submandibular method. The submandibular method involves using a disposable metal lancet to puncture the submandibular vein that runs below and behind a rodent's mandible (**Figure 8B**). The method yields blood samples of variable but generally comparable volume to those generated by the retroorbital method but can be applied with little or no need for anesthesia. The submandibular method has proven highly effective with laboratory mice (Golde et al. 2005) but, to date, has been used relatively rarely in field studies. Given the relative success of this method during its initial years of sampling, NEON has adopted it as the only allowable method for blood collection.

Other methods of blood collection, including tail and saphenous vein bleeding methods (Hem et al. 1998, Abatan et al. 2008) are not as attractive as the retroorbital and submandibular methods because they generate samples of relatively small volumes – much less than the 20 microliters required for a hantavirus ELISA assay.

To minimize increases in sampling-related morbidity and mortality, NEON will only collect blood from animals that meet five criteria. First, the animal must be a member of a taxonomic group for which the sampling does not cause significant stress or injury (i.e., not a member of the family Heteromyidae). For both the retroorbital and submandibular methods, this includes most species of rodents in the family Cricetidae (e.g., *Peromyscus* spp.) and other taxa (i.e., Dipodids, Murids) for which the methods are anatomically suitable and their application (including use of anesthesia) is not unduly stressful. Second, NEON must have a permit to handle and collect blood from individuals of the species in question. Third, the animal must weigh at least ten grams. Fourth, the animal should appear to be in good health and not show signs of pronounced or physically debilitating injury (e.g., blindness in or damage to one or both eyes, one or more broken or deformed limbs). Finally, the animal can only be bled once during any given sampling event. Upon capture, an animal meeting these criteria will be anesthetized (if needed) and bled.

Methods for capturing small mammals are described in the NEON TOS Science Design for Small Mammal Abundance and Diversity (AD[08]) and TOS Protocol and Procedure: Small Mammal Sampling (AD[05]). In the field, blood samples will be collected into labeled cryovials and immediately frozen on dry ice in portable coolers. Coolers will be transported to a NEON domain lab, where cryovials will be stored at -80°C until they are sent to one or more external facilities for pathogen testing.



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Figure 8. Diagrams illustrating retroorbital (A) and submandibular (B) bleeding methods.

In 2012, rodent-borne pathogen sampling was prototyped at Rocky Mountain National Park in NEON Domain 10. The goals of the exercise were to test the feasibility of lab-based training in blood collection and field sampling. Training of field staff in the retroorbital bleeding method, many of whom had no prior experience handling rodents or collecting blood samples, was conducted during a one-day, hands-on course at the animal care facility of Colorado State University (Fort Collins, CO). Experiences during field sampling and resulting data demonstrated the feasibility of the proposed design and sampling protocol. Small mammal abundance and diversity sampling was conducted monthly on ten 100-trap grids for four months. Coincident with this activity, blood samples were collected from a total of 272 rodents (**Table 2**). These samples were sent to and analyzed by the lab of Dr. Brian Hjelle at the University of New Mexico. Results of strip immunoblot assay tests indicated that, of the four rodent species tested, evidence of hantavirus (presumably Sin Nombre virus) infection was only apparent in deer mice populations, which had an estimated infection prevalence of 10.3% at the site level and across the sampling period. Given the relative ease of the submandibular method as compared to the retro-orbital technique, coupled with the subsequent years of demonstrated success by field staff (Appendix B), the preliminary results of this prototype have been supported through time.



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Table 2. Results of rodent-borne pathogen sampling in Rocky Mountain National Park (NEON Domain 10, spring and summer 2012).

Species	# individuals tested	# individuals positive	Seroprevalence (%)
Long-tailed vole (<i>Microtus longicaudus</i>)	16	0	0.0
Montane vole (<i>Microtus montanus</i>)	8	0	0.0
Southern red-backed vole (<i>Myodes gapperi</i>)	15	0	0.0
Deer mouse (<i>Peromyscus maniculatus</i>)	233	24	10.3
	272	24	8.8

Many rodents play important roles as intermediate hosts for ticks and reservoirs for tick-borne pathogens. Because of the very large size of many rodent populations and the fact that individuals of particular species are highly competent pathogen reservoirs, rodents make significant contributions to the demography of ticks and the epizootiology of tick-borne pathogens (LoGiudice et al. 2003, Ostfeld et al. 2006). In recognition of these relationships there is strong interest in quantifying and collecting larval and nymphal ticks attached to rodents captured as part of NEON small mammal sampling. Nevertheless, removal of ticks from captured small mammals is not currently planned. This decision is driven by the need to limit handling time in the face of other prioritized data and sample collection requirements (e.g., length and weight measurements, blood collection). If time permits during processing of captured small mammals, NEON field staff will inspect and count attached larval and nymphal ticks. This approach has been shown to generate reliable estimates of absolute tick burdens for some cricetid and sciurid rodents (Brunner and Ostfeld 2008). Protocol details associated with any observational sampling of ticks on small mammals will be captured in TOS Protocol and Procedure: Small Mammal Sampling (AD[05]).

6.3.1.2 Pathogen Testing

All collected blood samples will be tested for serum antibodies reactive against hantaviruses and arenaviruses (the latter only if resources permit) using ELISAs. Test results will indicate past exposure to or infection by pathogens of interest (Elgh et al. 1997). External analytical facilities will report the sensitivity and specificity of their testing methods to NEON whenever possible. Samples in which select agents have been identified will be handled in accordance with state and/or federal regulations.

6.3.1.3 Archive

Blood samples remaining after NEON pathogen testing will be archived. Archiving plans, including details on accessing samples for private investigation are detailed in the NEON Bioarchive Concept of Operations (AD[09]).



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6.3.2 Spatial Distribution of Sampling

A stratified random approach will be used to select three to eight small mammal sampling grids within the dominant vegetation types ($\geq 5\%$ total cover) at each site (NEON TOS Science Design for Spatial Sampling; AD[02]). The number of plots per type will be proportional to the percent cover of each type. Plot locations will additionally be constrained to fall within 350m of roads because of a need to facilitate deployment and retrieval of sampling equipment.

During the first year of sampling, blood samples for use in pathogen testing will be collected from animals captured on up to three of the sampling grids at each site. Grids with the highest abundance of the rodent taxa targeted for blood collection will be selected, with a preference for grids in the most dominant vegetation type at each site. Following the first year of sampling, grid-level data on the abundance and diversity of small mammals collected during the first year of sampling will be used to select up to three long-term bleeding grids to be sampled beginning in the second year of sampling. Once a set of long-term bleed grids is selected, NEON will maintain a fixed plots design, sampling those grids irrespective of changes in the presence or abundance of rodents and/or associated pathogens.

6.3.3 Temporal Distribution of Sampling

Sampling events will occur six times per year at core sites and four time per year at gradient sites. The sampling frequency proposed for gradient sites represents a minimum and reflects a reduced frequency relative to the original design of 12 and 6 sampling events per year due to budget constraints. At core and gradient sites, respectively. Lower sampling frequencies have been shown to significantly increase error in estimates of both rodent abundance and prevalence of serum antibodies reactive against pathogens (Carver et al. 2010). Sampling will occur during any months when sampling personnel can access plots and weather conditions are safe for personnel and captured animals, with most sampling happening during the growing season.

6.3.3.1 Power Analyses Evaluating Sampling Frequencies

Evaluation of power to detect prevalence trends

For power analyses evaluating the ability of sampling and testing to detect long-term trends in prevalence of serum antibodies reactive against rodent-borne hantaviruses and potentially arenaviruses, year-zero antibody prevalence values were set to 2.5% (rare pathogen) or 10% (common pathogen) based on results of long-term studies of Sin Nombre virus infection in deer mice populations (Douglass et al. 2001, Calisher et al. 2007). In addition to evaluating the power associated with sampling event frequencies of every four and eight weeks, frequencies of one, two, three and four times per year were evaluated to generate power curves. Given an estimated capture success of 10-20% (based on data published in Thibault et al. (2011)) and an anticipated sampling effort of between one and three 100-trap trapping grids per site, between 30 and 60 rodents are likely to be sampled per site/sampling event combination. Based on this, a conservative per-event sample size threshold of 40 rodents bled and



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tested for antibodies reactive against pathogens was used. Thus, a sampling frequency was deemed adequate if 40 or fewer samples per event were sufficient to detect an antibody prevalence trend of a given magnitude.

When power analyses were run with a term depicting temporal autocorrelation in the antibody prevalence data, the proposed design was only capable of detecting annual prevalence increases of $\leq 1.0\%$ for a common pathogen at a core site (i.e., over 25 years). In this scenario, the design can detect an annual increase of 1.0% with higher confidence ($\alpha=0.05$, power=0.9) and an increase of 0.75% with lower confidence ($\alpha=0.1$, power=0.8) (**Figure 9A**). In the absence of temporal autocorrelation in the data, the design can detect annual increases of 0.5% and 0.25% with high confidence, but the latter only at a sampling frequency of one event every four weeks. Similarly, for a rare pathogen at a core site, the design can detect annual increases of 0.75% and 0.5% with high confidence, but the latter only at the one event every four weeks sampling frequency (**Figure 9B**). In contrast, the ability of the design to detect annual prevalence increases of $<1.0\%$ at gradient sites (i.e., over 10 years) is considerably lower. When the pathogen is common the design can detect an annual prevalence increase of 1.0% with higher confidence, but only at the highest sampling frequency (one event every four weeks) (**Figure 9C**). When the pathogen is rare the design cannot detect an annual increase of $<1.0\%$ irrespective of the level of confidence or sampling frequency (Figure 9D). While the power of rodent-borne pathogen sampling to detect interannual infection trends for rare pathogens is limited, the probability of detecting these pathogens when they are present at a site is still reasonably high. For example, using the same methods as employed for similar mosquito-borne pathogen calculations, the design has a 64% probability of detecting a rare rodent-borne pathogen (2.5% antibody prevalence) in a particular sampling event when 40 rodents are tested. At a mean prevalence of 4% that probability rises to 80%.

Evaluation of power to detect pathogen(s)

In terms of simply detecting antibodies reactive against hantaviruses or arenaviruses (i.e., presence/absence of infection) in a sampled reservoir population, the design has a 99% probability of detecting a common pathogen and a 64% probability of detecting a rare pathogen when 40 or more rodents per species are tested per sampling event (**Figure 10A**). That probability falls to 33% when infection is very rare (1% prevalence). When analytical sampling effort is reduced to 20 rodents tested per sampling event, these probabilities fall to 88%, 40% and 18%, respectively. A 50% probability of detecting infection requires testing 7, 29, or 60 rodents per sampling bout, respectively (**Figure 10B**). These sample sizes must be increased to 15, 67, and 160 rodents to raise the detection probability to 80%.

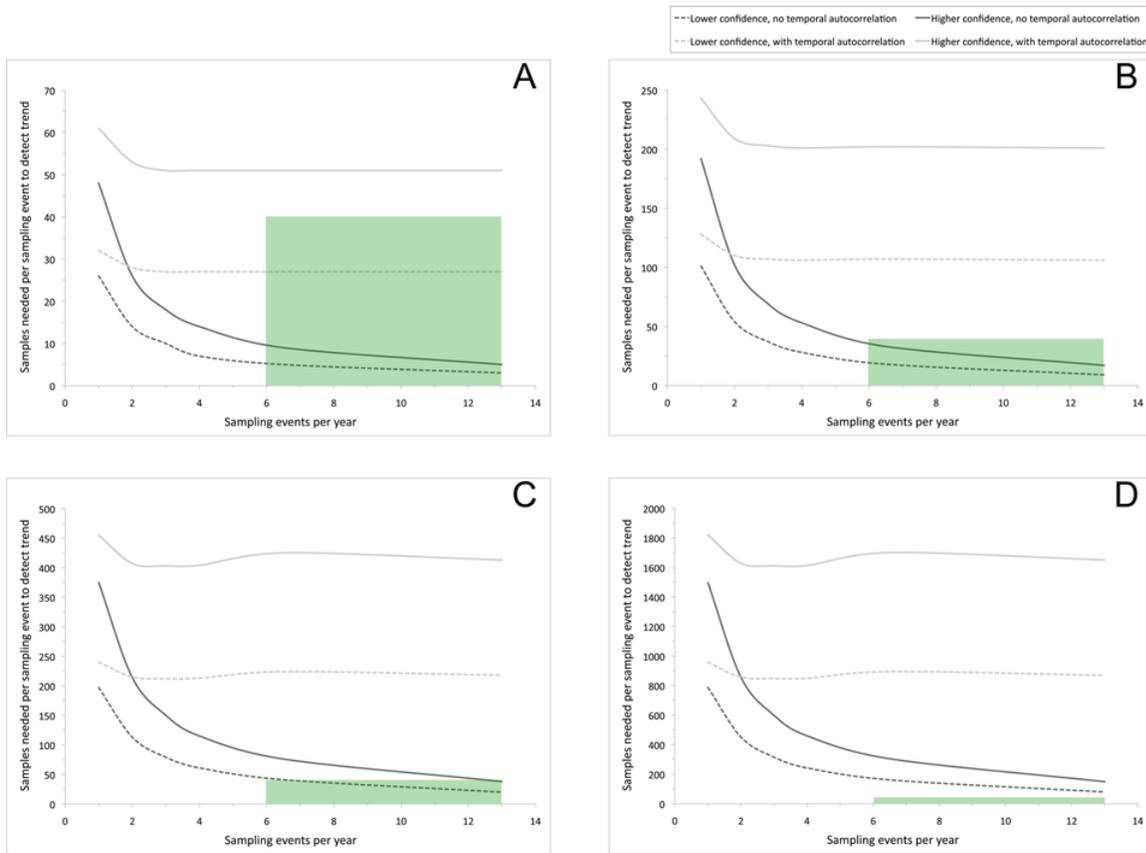


Figure 9. Results of power analyses for a common rodent-borne pathogen (prevalence=10%) at a core (i.e., 25 years, A) and gradient (i.e., 10 years, B) site, and a rare rodent-borne pathogen (prevalence=2.5%) at a core (C) and gradient (D) site. Each analysis includes results for all combinations of low ($\alpha=0.1$, power=0.8) and high ($\alpha=0.05$, power=0.9) confidence and with and without a temporal autocorrelation term (e^{-1} at a half year). Green boxes indicate levels of sampling effort (event frequency and number samples/event to be tested for pathogens) specified in the design.

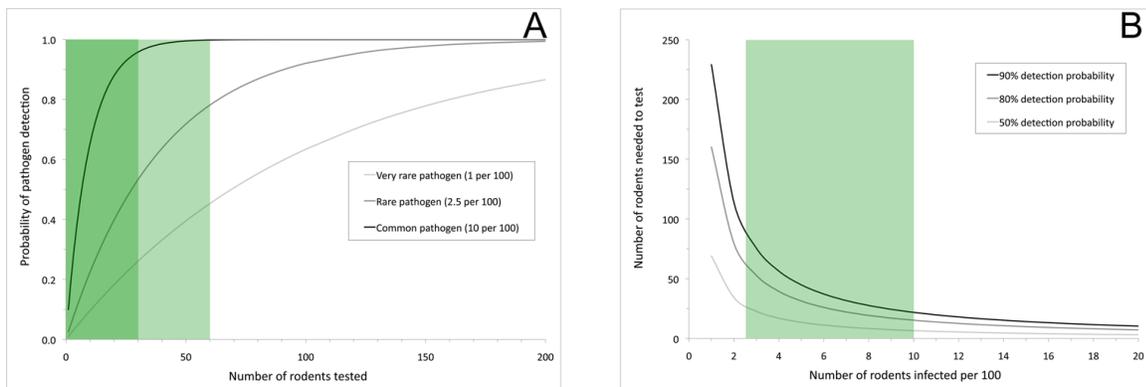


Figure 10. (A) Relationship between the number of samples tested for pathogens and the probability of detecting rodent-borne pathogens. Green boxes indicated upper and lower estimates of number of samples to be tested per sampling event. (B) Minimum number of samples required for specified detection probabilities at various levels of



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infection prevalence for rodent-borne pathogens. Green box indicates estimated prevalence of rare and common pathogens at NEON sites.

6.3.4 Logistics and Adaptability

In the event that sampling effort must be reduced, the sampling design (e.g., frequency of sampling events, number of plots per site) should be maintained, and cost savings realized through reductions in the number of samples that are tested for pathogens. Foregoing the testing of all samples collected during selected sampling events, rather than reducing the number of samples tested for every event, will facilitate data comparability by maintaining more consistent levels of analytical sampling effort and associated uncertainty through time. Remaining samples (i.e., untested or residual) can be archived for processing at a later date, either by NEON or through agreements with other members of the research community.

As an alternative to blood-based testing, many rodent-borne viral pathogens (including hantaviruses and arenaviruses) can be detected in fecal samples using PCR-based methods (Phan et al. 2011). This approach is attractive for a number of reasons. The collection of fecal samples does not require anesthetization and is minimally invasive for sampled rodents. Additionally, use of fecal samples would allow for the sampling of rodents that could not be bled due to their small size. This would expand rodent-borne pathogen sampling to include younger rodents as well as adults of species that are anatomically and/or physiologically not amenable to blood collection. Among these are shrews, known reservoirs of hantaviruses (Arai et al. 2008) that typically experience high and often fatal levels of stress associated with handling and blood sample collection. PCR-based analyses of tissue (including fecal) samples for rodent-borne pathogens are currently much more expensive than enzyme-based tests and are therefore less attractive as fewer samples can be tested for a given level of funding. Additionally, it is not always possible to collect fecal samples on demand. NEON may consider the use of PCR-based tests in the future if/when associated costs of PCR-based testing decline. NEON will collect (but not analyze) fecal samples from captured rodents and store/archive these in a manner that will preserve them for use in such pathogen testing (frozen at -80C).

Data on tick burdens collected as part of small mammal sampling could be used to estimate the additional handling time that would be required to remove ticks from sampled small mammals. These estimates would allow the feasibility of incorporating tick collection into the NEON small mammal sampling protocol to be evaluated.



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7 REFERENCES

- Abatan, O. I., K. B. Welch, and J. A. Nemzek. 2008. Evaluation of saphenous venipuncture and modified tail-clip blood collection in mice. *Journal of the American Association for Laboratory Animal Science* **47**:8-15.
- Aguirre, A. A., R. S. Ostfeld, and P. Daszak, editors. 2012. *New Directions in Conservation Medicine: Applied Cases of Ecological Health*. Oxford University Press.
- Alexander, H. M. 2010. Disease in natural plant populations, communities, and ecosystems: Insights into ecological and evolutionary processes. *Plant Disease* **94**:492-503.
- Allan, B. F., H. P. Dutra, L. S. Goessling, K. Barnett, J. M. Chase, R. J. Marquis, G. Pang, G. A. Storch, R. E. Thach, and J. L. Orrock. 2010. Invasive honeysuckle eradication reduces tick-borne disease risk by altering host dynamics. *Proceedings of the National Academy of Sciences of the United States of America* **107**:18523-18527.
- Almeida, F. F. L. and D. E. Gorla. 1995. Daily pattern of flight activity of *Aedes albifasciatus* in central Argentina. *Memorias Do Instituto Oswaldo Cruz* **90**:639-644.
- Altizer, S., A. Dobson, P. Hosseini, P. Hudson, M. Pascual, and P. Rohani. 2006. Seasonality and the dynamics of infectious diseases. *Ecology Letters* **9**:467-484.
- Altizer, S., R. S. Ostfeld, P. T. Johnson, S. Kutz, and C. D. Harvell. 2013. Climate change and infectious diseases: from evidence to a predictive framework. *Science* **341**:514-519.
- Anderson, P. K., A. A. Cunningham, N. G. Patel, F. J. Morales, P. R. Epstein, and P. Daszak. 2004. Emerging infectious diseases of plants: pathogen pollution, climate change and agrotechnology drivers. *Trends in Ecology & Evolution* **19**:535-544.
- Andreadis, T. G., J. F. Anderson, C. R. Vossbrinck, and A. J. Main. 2004. Epidemiology of West Nile virus in Connecticut: A five-year analysis of mosquito data 1999-2003. *Vector-Borne and Zoonotic Diseases* **4**:360-378.
- Arai, S., S. N. Bennett, L. Sumibcay, J. A. Cook, J. W. Song, A. Hope, C. Parmenter, V. R. Nerurkar, T. L. Yates, and R. Yanagihara. 2008. Short report: Phylogenetically distinct hantaviruses in the masked shrew (*Sorex cinereus*) and dusky shrew (*Sorex monticolus*) in the United States. *American Journal of Tropical Medicine and Hygiene* **78**:348-351.
- Atlas, R., C. Rubin, S. Maloy, P. Daszak, R. Colwell, and B. Hyde. 2010. One Health - Attaining optimal health for people, animals, and the environment. *Microbe* **5**:383-389.
- Auffray, J.-C., K. Blasdel, F. Bordes, M. Chabé, K. Chaisiri, N. Charbonnel, Y. Chaval, J. Claude, J.-F. Cosson, E. Dei-Cas, M. Desquesnes, G. Dobigny, B. Douangboupha, M. Galan, V. Haukisalmi, H. Henttonen, V. Herbreteau, J.-P. Hugot, T. Jiyipong, A. Latinne, J. Michaux, C. Milocco, S. Morand, M. Pagès, D. Phoophitpong, P. Pumhom, A. R. Salvador, S. Soonchan, Y. Suputtamongkol, S.



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- Waengsothorn, W. Duangdao, and A. Xuéreb, editors. 2011. *Protocols for Field and Laboratory Rodent Studies*. Kasetsart University Press.
- Bailey, S., B. Hoffmann, and D. Eliason. 1965. Flight and dispersal of the mosquito *Culex tarsalis coquillett* in the Sacramento Valley of California. University of California.
- Barbour, A. G. and D. Fish. 1993. The biological and social phenomenon of Lyme disease. *Science* **260**:1610-1616.
- Bhatt, S., P. W. Gething, O. J. Brady, J. P. Messina, A. W. Farlow, C. L. Moyes, J. M. Drake, J. S. Brownstein, A. G. Hoen, O. Sankoh, M. F. Myers, D. B. George, T. Jaenisch, G. R. W. Wint, C. P. Simmons, T. W. Scott, J. J. Farrar, and S. I. Hay. 2013. The global distribution and burden of dengue. *Nature* **496**:504-507.
- Binder, S., A. M. Levitt, J. J. Sacks, and J. M. Hughes. 1999. Emerging infectious diseases: Public health issues for the 21st century. *Science* **284**:1311-1313.
- Brault, A. C. and W. K. Reisen. 2013. Environmental perturbations that influence arboviral host range: insights into emergence mechanisms. Pages 57-75 in S. K. Singh, editor. *Viral Infections and Global Change*. John Wiley and Sons, Inc., New York, NY.
- Brooks, D. R. and E. P. Hoberg. 2013. The emerging infectious diseases crisis and pathogen pollution. Pages 213-229 in C. Rhode, editor. *The Balance of Nature and Human Impact*. Cambridge University Press.
- Brown, J. A., M. M. Salehi, M. Moradi, B. Panahbehagh, and D. R. Smith. 2013. Adaptive survey designs for sampling rare and clustered populations. *Mathematics and Computers in Simulation* **93**:108-116.
- Brown, K. and D. Gilfoyle, editors. 2010. *Healing the Herds: Disease, Livestock Economies, and the Globalization of Veterinary Medicine*. Ohio University Press, Athens, OH.
- Brunner, J. L. and R. S. Ostfeld. 2008. Multiple causes of variable tick burdens on small-mammal hosts. *Ecology* **89**:2259-2272.
- Burkett-Cadena, N. D., M. D. Eubanks, and T. R. Unnasch. 2008. Preference of female mosquitoes for natural and artificial resting sites. *Journal of the American Mosquito Control Association* **24**:228-235.
- Calisher, C. H. 1994. Medically important arboviruses of the United States and Canada. *Clinical Microbiology Reviews* **7**:89-116.
- Calisher, C. H., K. D. Wagoner, B. R. Amman, J. J. Root, R. J. Douglass, A. J. Kuenzi, K. D. Abbott, C. Parmenter, T. L. Yates, T. G. Ksiazek, B. J. Beaty, and J. N. Mills. 2007. Demographic factors associated with prevalence of antibody to Sin Nombre virus in deer mice in the western United States. *Journal of Wildlife Diseases* **43**:1-11.



- Caminade, C., J. M. Medlock, E. Ducheyne, K. M. McIntyre, S. Leach, M. Baylis, and A. P. Morse. 2012. Suitability of European climate for the Asian tiger mosquito *Aedes albopictus*: recent trends and future scenarios. *Journal of the Royal Society Interface* **9**:2708-2717.
- Carver, S., J. N. Mills, A. Kuenzi, T. Flietstra, and R. Douglass. 2010. Sampling frequency differentially influences interpretation of zoonotic pathogen and host dynamics: Sin Nombre virus and deer mice. *Vector-Borne and Zoonotic Diseases* **10**:575-583.
- Centers for Disease Control and Prevention. 2008. Surveillance for Lyme disease - United States, 1992-2006. *Morbidity and Mortality Weekly Report* **57**:1-12.
- Centers for Disease Control and Prevention. 2010. Summary of notifiable diseases - United States, 2010. *Morbidity and Mortality Weekly Report* **59**:1-116.
- Centers for Disease Control and Prevention. 2012a. Babesiosis surveillance - 18 states, 2011. *Morbidity and Mortality Weekly Report* **61**:505-509.
- Centers for Disease Control and Prevention. 2012b. Hantavirus pulmonary syndrome in visitors to a national park - Yosemite valley, California, 2012. *Morbidity and Mortality Weekly Report* **61**:952.
- Childs, J. E. and C. D. Paddock. 2003. The ascendancy of *Amblyomma americanum* as a vector of pathogens affecting humans in the United States. *Annual Review of Entomology* **48**:307-337.
- Ciota, A. T., C. L. Drummond, J. Drobnack, M. A. Ruby, L. D. Kramer, and G. D. Ebel. 2011. Emergence of *Culex pipiens* from overwintering hibernacula. *Journal of the American Mosquito Control Association* **27**:21-29.
- Clark, D. D. 1995. Lower temperature limits for activity of several Ixodid ticks (Acarl, Ixodidae) - Effects of body size and rate of temperature change. *Journal of Medical Entomology* **32**:449-452.
- Clark, K. L., J. H. Oliver, J. M. Grego, A. M. James, L. A. Durden, and C. W. Banks. 2001. Host associations of ticks parasitizing rodents at *Borrelia burgdorferi* enzootic sites in South Carolina. *Journal of Parasitology* **87**:1379-1386.
- Clark, K. L., J. H. Oliver, D. B. McKechnie, and D. C. Williams. 1998. Distribution, abundance, and seasonal activities of ticks collected from rodents and vegetation in South Carolina. *Journal of Vector Ecology* **23**:89-105.
- Clay, C. A., E. M. Lehmer, S. S. Jeor, and M. D. Dearing. 2009. Sin Nombre Virus and rodent species diversity: A test of the dilution and amplification hypotheses. *Plos One* **4**.
- Cleaveland, S., M. K. Laurenson, and L. H. Taylor. 2001. Diseases of humans and their domestic mammals: pathogen characteristics, host range and the risk of emergence. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* **356**:991-999.
- Coakley, S. M., H. Scherm, and S. Chakraborty. 1999. Climate change and plant disease management. *Annual Review of Phytopathology* **37**:399-426.



Title: TOS Science Design for Vectors and Pathogens		Date: 04/06/2022
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- Coker, R., J. Rushton, S. Mounier-Jack, E. Karimuribo, P. Lutumba, D. Kambarage, D. U. Pfeiffer, K. Stark, and M. Rweyemamu. 2011. Towards a conceptual framework to support one-health research for policy on emerging zoonoses. *Lancet Infectious Diseases* **11**:326-331.
- Cooley, R. A. and G. M. Kohls. 1944. The genus *Amblyomma* in the United States. *Journal of Parasitology* **30**:77-111.
- Cooley, R. A. and G. M. Kohls. 1945. The genus *Ixodes* in North America. *National Institutes of Health Bulletin* **184**.
- Corbet, P. S. and H. V. Danks. 1973. Seasonal emergence and activity of mosquitoes (Diptera - Culicidae) in a high Arctic locality. *Canadian Entomologist* **105**:837-872.
- Crowl, T. A., T. O. Crist, R. R. Parmenter, G. Belovsky, and A. E. Lugo. 2008. The spread of invasive species and infectious disease as drivers of ecosystem change. *Frontiers in Ecology and the Environment* **6**:238-246.
- Darsie, R. F. and R. A. Ward. 1981. Identification and geographical distribution of the mosquitoes of North America, North of Mexico. *Mosquito Systematics Supplement* **1**:1-313.
- Daszak, P., A. A. Cunningham, and A. D. Hyatt. 2000. Wildlife ecology - Emerging infectious diseases of wildlife - Threats to biodiversity and human health. *Science* **287**:443-449.
- Daszak, P., A. A. Cunningham, and A. D. Hyatt. 2001. Anthropogenic environmental change and the emergence of infectious diseases in wildlife. *Acta Tropica* **78**:103-116.
- Daszak, P., G. M. Tabor, A. M. Kilpatrick, J. Epstein, and R. Plowright. 2004. Conservation medicine and a new agenda for emerging diseases. *Annals of the New York Academy of Sciences* **1026**:1-11.
- Daubney, R., J. R. Hudson, and P. C. Garnham. 1931. Enzootic hepatitis or Rift Valley fever. An undescribed virus disease of sheep cattle and man from East Africa. *Journal of Pathology and Bacteriology* **34**:545-579.
- Dister, S. W., D. Fish, S. M. Bros, D. H. Frank, and B. L. Wood. 1997. Landscape characterization of peridomestic risk for Lyme disease using satellite imagery. *American Journal of Tropical Medicine and Hygiene* **57**:687-692.
- Diuk-Wasser, M. A., A. G. Hoen, P. Cislo, R. Brinkerhoff, S. A. Hamer, M. Rowland, R. Cortinas, G. Vourc'h, F. Melton, G. J. Hickling, J. I. Tsao, J. Bunikis, A. G. Barbour, U. Kitron, J. Piesman, and D. Fish. 2012. Human risk of infection with *Borrelia burgdorferi*, the Lyme disease agent, in eastern United States. *American Journal of Tropical Medicine and Hygiene* **86**:320-327.
- Dobson, A. and J. Foufopoulos. 2001. Emerging infectious pathogens of wildlife. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* **356**:1001-1012.
- Dobson, A. and B. Grenfell, editors. 1995. *Ecology of Infectious Disease in Natural Populations*. Cambridge University Press, Cambridge, UK.



Title: TOS Science Design for Vectors and Pathogens		Date: 04/06/2022
NEON Doc. #: NEON.DOC.000911	Author: Y. Springer	Revision: C

- Dobson, A. P. and E. R. Carper. 1996. Infectious diseases and human population history - Throughout history the establishment of disease has been a side effect of the growth of civilization. *Bioscience* **46**:115-126.
- Donahue, J. G., J. Piesman, and A. Spielman. 1987. Reservoir competence of white footed mice for Lyme disease spirochetes. *American Journal of Tropical Medicine and Hygiene* **36**:92-96.
- Doudier, B., J. Olano, P. Parola, and P. Brouqui. 2010. Factors contributing to emergence of *Ehrlichia* and *Anaplasma* spp. as human pathogens. *Veterinary Parasitology* **167**:149-154.
- Douglass, R. J., A. J. Kuenzi, T. Wilson, and R. C. Van Horne. 2000. Effects of bleeding nonanesthetized wild rodents on handling mortality and subsequent recapture. *Journal of Wildlife Diseases* **36**:700-704.
- Douglass, R. J., W. J. Semmens, S. J. Matlock-Cooley, and A. J. Kuenzi. 2006. Deer mouse movements in peridomestic and sylvan settings in relation to Sin Nombre virus antibody prevalence. *Journal of Wildlife Diseases* **42**:813-818.
- Douglass, R. J., T. Wilson, W. J. Semmens, S. N. Zanto, C. W. Bond, R. C. Van Horn, and J. N. Mills. 2001. Longitudinal studies of Sin Nombre virus in deer mouse-dominated ecosystems of Montana. *American Journal of Tropical Medicine and Hygiene* **65**:33-41.
- Duffy, D. C. and S. R. Campbell. 1994. Ambient air temperature as a predictor of activity of adult *Ixodes scapularis* (Acari, Ixodidae). *Journal of Medical Entomology* **31**:178-180.
- Earley, E., P. H. Peralta, and K. M. Johnson. 1967. A plaque neutralization method for arboviruses. *Proceedings of the Society for Experimental Biology and Medicine* **125**:741-&.
- Eisen, L., R. J. Eisen, and R. S. Lane. 2004. The roles of birds, lizards, and rodents as hosts for the western black legged tick *Ixodes pacificus*. *Journal of Vector Ecology* **29**:295-308.
- Eisen, L. and C. G. Moore. 2013. *Aedes (Stegomyia) aegypti* in the continental United States: A vector at the cool margin of its geographic range. *Journal of Medical Entomology* **50**:467-478.
- Elgh, F., A. Lundkvist, O. A. Alexeyev, H. Stenlund, T. Avsic-Zupanc, B. Hjelle, H. W. Lee, K. J. Smith, R. Vainionpaa, D. Wiger, G. Wadell, and P. Juto. 1997. Serological diagnosis of hantavirus infections by an enzyme-linked immunosorbent assay based on detection of immunoglobulin G and M responses to recombinant nucleocapsid proteins of five viral serotypes. *Journal of Clinical Microbiology* **35**:1122-1130.
- Epstein, P. R., H. F. Diaz, S. Elias, G. Grabherr, N. E. Graham, W. J. M. Martens, E. Mosley-Thompson, and J. Susskind. 1998. Biological and physical signs of climate change: Focus on mosquito-borne diseases. *Bulletin of the American Meteorological Society* **79**:409-417.
- Estrada-Pena, A. 2009. Tick-borne pathogens, transmission rates and climate change. *Frontiers in Bioscience* **14**:2674-U2780.



Title: TOS Science Design for Vectors and Pathogens		Date: 04/06/2022
NEON Doc. #: NEON.DOC.000911	Author: Y. Springer	Revision: C

- Falco, R. C. and D. Fish. 1989. The use of carbon dioxide-baited tick traps for sampling *Ixodes dammini* (Acari, Ixodidae). *Acarologia* **30**:29-33.
- Falco, R. C. and D. Fish. 1992. A comparison of methods for sampling the deer tick, *Ixodes dammini*, in a Lyme disease endemic area. *Experimental & Applied Acarology* **14**:165-173.
- Farrington, C. P. 1992. Estimating prevalence by group testing using generalized linear models. *Statistics in Medicine* **11**:1591-1597.
- Fisman, D. N. 2007. Seasonality of infectious diseases. Pages 127-143 *Annual Review of Public Health*.
- Gage, K. L., T. R. Burkot, R. J. Eisen, and E. B. Hayes. 2008. Climate and Vectorborne Diseases. *American Journal of Preventive Medicine* **35**:436-450.
- Gage, K. L. and M. Y. Kosoy. 2005. Natural history of plague: Perspectives from more than a century of research. *Annual Review of Entomology* **50**:505-528.
- Garcia, R. 1962. Carbon dioxide as an attractant for certain ticks (Acarina: Argasidae and Ixodidae). *Annals of the Entomological Society of America* **55**:605-606.
- Garnett, G. P. and E. C. Holmes. 1996. The ecology of emergent infectious disease - Infectious diseases pose an ever-emerging threat to humanity. *Bioscience* **46**:127-135.
- Gatewood, A. G., K. A. Liebman, G. Vourc'h, J. Bunikis, S. A. Hamer, R. Cortinas, F. Melton, P. Cislo, U. Kitron, and J. Tsao. 2009. Climate and tick seasonality are predictors of *Borrelia burgdorferi* genotype distribution. *Applied and Environmental Microbiology* **75**:2476-2483.
- Gibson, C. M., R. H. Kao, K. K. Blevins, and P. D. Travers. 2012. Integrative taxonomy for continental-scale terrestrial insect observations. *Plos One* **7**:e37528.
- Ginsberg, H. S. and C. P. Ewing. 1989. Comparison of flagging, walking, trapping, and collecting from hosts as sampling methods for northern deer ticks, *Ixodes dammini*, and lone star ticks, *Amblyomma americanum* (Acari, Ixodidae). *Experimental and Applied Acarology* **7**:313-322.
- Ginsberg, H. S., E. Zhioua, S. Mitra, J. Fischer, P. A. Buckley, F. Verret, H. B. Underwood, and F. G. Buckley. 2004. Woodland type and spatial distribution of nymphal *Ixodes scapularis* (Acari : Ixodidae). *Environmental Entomology* **33**:1266-1273.
- Golde, W. T., P. Gollobin, and L. L. Rodriguez. 2005. A rapid, simple, and humane method for submandibular bleeding of mice using a lancet. *Lab Animal* **34**:39-43.
- Gomez, A. and E. Nichols. 2013. Neglected wild life: Parasitic biodiversity as a conservation target. *International Journal for Parasitology: Parasites and Wildlife* **2**:222-227.
- Gould, E. A. and S. Higgs. 2009. Impact of climate change and other factors on emerging arbovirus diseases. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **103**:109-121.
- Gray, J. 1998. Review: The ecology of ticks transmitting Lyme borreliosis. *Experimental & Applied Acarology* **22**: 249. <https://doi.org/10.1023/A:1006070416135>.



- Gu, W. D. and R. J. Novak. 2004. Short report: Detection probability of arbovirus infection in mosquito populations. *American Journal of Tropical Medicine and Hygiene* **71**:636-638.
- Gu, W. D., T. R. Unnasch, C. R. Katholi, R. Lampman, and R. J. Novak. 2008. Fundamental issues in mosquito surveillance for arboviral transmission. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **102**:817-822.
- Gubler, D. J. 1998. Resurgent vector-borne diseases as a global health problem. *Emerging Infectious Diseases* **4**:442-450.
- Gubler, D. J., P. Reiter, K. L. Ebi, W. Yap, R. Nasci, and J. A. Patz. 2001. Climate variability and change in the United States: Potential impacts on vector- and rodent-borne diseases. *Environmental Health Perspectives* **109**:223-233.
- Harvell, C. D., K. Kim, J. M. Burkholder, R. R. Colwell, P. R. Epstein, D. J. Grimes, E. E. Hofmann, E. K. Lipp, A. Osterhaus, R. M. Overstreet, J. W. Porter, G. W. Smith, and G. R. Vasta. 1999. Review: Marine ecology - Emerging marine diseases - Climate links and anthropogenic factors. *Science* **285**:1505-1510.
- Harvell, C. D., C. E. Mitchell, J. R. Ward, S. Altizer, A. P. Dobson, R. S. Ostfeld, and M. D. Samuel. 2002. Ecology - Climate warming and disease risks for terrestrial and marine biota. *Science* **296**:2158-2162.
- Hatcher, M. J., J. T. Dick, and A. M. Dunn. 2014. Parasites that change predator or prey behaviour can have keystone effects on community composition. *Biology letters* **10**:20130879.
- Hebert, P. D. N., A. Cywinska, S. L. Ball, and J. R. DeWaard. 2003. Biological identifications through DNA barcodes. *Proceedings of the Royal Society B-Biological Sciences* **270**:313-321.
- Hem, A., A. J. Smith, and P. Solberg. 1998. Saphenous vein puncture for blood sampling of the mouse, rat, hamster, gerbil, guinea pig, ferret and mink. *Laboratory Animals* **32**:364-368.
- Hoel, D. F., D. L. Kline, and S. A. Allan. 2009. Evaluation of six mosquito traps for collection of *Aedes albopictus* and associated mosquito species in a suburban setting in north central Florida. *Journal of the American Mosquito Control Association* **25**:47-57.
- Huchon, D., O. Madsen, M. Sibbald, K. Ament, M. J. Stanhope, F. Catzeflis, W. W. de Jong, and E. J. P. Douzery. 2002. Rodent phylogeny and a timescale for the evolution of glires: Evidence from an extensive taxon sampling using three nuclear genes. *Molecular Biology and Evolution* **19**:1053-1065.
- Hudson, P., A. Rizzoli, B. Grenfell, H. Heesterbeek, and A. Dobson, editors. 2002. *The Ecology of Wildlife Diseases*. Oxford University Press, Oxford, UK.
- Hudson, P. J., A. P. Dobson, and K. D. Lafferty. 2006. Is a healthy ecosystem one that is rich in parasites? *Trends in Ecology & Evolution* **21**:381-385.



- Jones, C. J. and U. D. Kitron. 2000. Populations of *Ixodes scapularis* (Acari : Ixodidae) are modulated by drought at a Lyme disease focus in Illinois. *Journal of Medical Entomology* **37**:408-415.
- Jones, K. E., N. G. Patel, M. A. Levy, A. Storeygard, D. Balk, J. L. Gittleman, and P. Daszak. 2008. Global trends in emerging infectious diseases. *Nature* **451**:990-994.
- Joslin, J. 2009. Blood collection techniques in exotic small mammals. *Journal of Exotic Pet Medicine* **18**:117-139.
- Kao, R. H., C. M. Gibson, R. E. Gallery, C. L. Meier, D. T. Barnett, K. M. Docherty, K. K. Blevins, P. D. Travers, E. Azuaje, Y. P. Springer, K. M. Thibault, V. J. McKenzie, M. Keller, L. F. Alves, E. L. S. Hinckley, J. Parnell, and D. Schimel. 2012. NEON terrestrial field observations: designing continental-scale, standardized sampling. *Ecosphere* **3**.
- Kaplan, B., L. Kahn, and T. Monath. 2009. 'One Health - One Medicine': Linking human, animal and environmental health. *Veterinaria Italiana* **45**:1-8.
- Keesing, F., L. K. Belden, P. Daszak, A. Dobson, C. D. Harvell, R. D. Holt, P. Hudson, A. Jolles, K. E. Jones, C. E. Mitchell, S. S. Myers, T. Bogich, and R. S. Ostfeld. 2010. Impacts of biodiversity on the emergence and transmission of infectious diseases. *Nature* **468**:647-652.
- Keirans, J. E. and T. R. Litwak. 1989. Pictorial key to the adults of hard ticks, family Ixodidae (Ixodida, Ixodoidea), east of the Mississippi river. *Journal of Medical Entomology* **26**:435-448.
- Keller, M., D. S. Schimel, W. W. Hargrove, and F. M. Hoffman. 2008. A continental strategy for the National Ecological Observatory Network. *Frontiers in Ecology and the Environment* **6**:282-284.
- Kinzer, D. R., S. M. Presley, and J. A. Hair. 1990. Comparative efficiency of flagging and carbon dioxide-baited sticky traps for collecting the lone star tick, *Amblyomma americanum* (Acarina, Ixodidae). *Journal of Medical Entomology* **27**:750-755.
- Kollars, T. M., J. H. Oliver, L. A. Durden, and P. G. Kollars. 2000. Host associations and seasonal activity of *Amblyomma americanum* (Acari : Ixodidae) in Missouri. *Journal of Parasitology* **86**:1156-1159.
- Komar, N., R. J. Pollack, and A. Spielman. 1995. A nestable fiber pot for sampling resting mosquitoes. *Journal of the American Mosquito Control Association* **11**:463-467.
- Kosek, J. C., R. I. Mazze, and M. J. Cousins. 1972. The morphology and pathogenesis of nephrotoxicity following methoxyflurane (penthrane) anesthesia. An experimental model in rats. *Laboratory investigation; a journal of technical methods and pathology* **27**:575.
- Kovats, R. S., D. H. Campbell-Lendrum, A. J. McMichael, A. Woodward, and J. S. Cox. 2001. Early effects of climate change: do they include changes in vector-borne disease? *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* **356**:1057-1068.
- Krockel, U., A. Rose, A. E. Eiras, and M. Geier. 2006. New tools for surveillance of adult yellow fever mosquitoes: comparison of trap catches with human landing rates in an urban environment. *Journal of the American Mosquito Control Association* **22**:229-238.



- Kuenzi, A. J., R. J. Douglass, D. White, C. W. Bond, and J. N. Mills. 2001. Antibody to Sin Nombre virus in rodents associated with peridomestic habitats in west central Montana. *American Journal of Tropical Medicine and Hygiene* **64**:137-146.
- Kuno, G. 1998. Universal diagnostic RT-PCR protocol for arboviruses. *Journal of Virological Methods* **72**:27-41.
- Kuno, G., C. J. Mitchell, G. J. J. Chang, and G. C. Smith. 1996. Detecting bunyaviruses of the Bunyamwera and California serogroups by a PCR technique. *Journal of Clinical Microbiology* **34**:1184-1188.
- Kwan, J. L., S. Klueh, M. B. Madon, and W. K. Reisen. 2010. West Nile virus emergence and persistence in Los Angeles, California, 2003-2008. *American Journal of Tropical Medicine and Hygiene* **83**:400-412.
- LaDeau, S. L., A. M. Kilpatrick, and P. P. Marra. 2007. West Nile virus emergence and large-scale declines of North American bird populations. *Nature* **447**:710-U713.
- Lafferty, K. D. 2009. The ecology of climate change and infectious diseases. *Ecology* **90**:888-900.
- Lanciotti, R. S., A. J. Kerst, R. S. Nasci, M. S. Godsey, C. J. Mitchell, H. M. Savage, N. Komar, N. A. Panella, B. C. Allen, K. E. Volpe, B. S. Davis, and J. T. Roehrig. 2000. Rapid detection of West Nile virus from human clinical specimens, field-collected mosquitoes, and avian samples by a TaqMan reverse transcriptase-PCR assay. *Journal of Clinical Microbiology* **38**:4066-4071.
- Lardeux, F. J., R. H. Tejerina, V. Quispe, and T. K. Chavez. 2008. A physiological time analysis of the duration of the gonotrophic cycle of *Anopheles pseudopunctipennis* and its implications for malaria transmission in Bolivia. *Malaria Journal* **7**.
- Leger, E., G. Vourc'h, L. Vial, C. Chevillon, and K. D. McCoy. 2013. Changing distributions of ticks: causes and consequences. *Experimental and Applied Acarology* **59**:219-244.
- Levi, T., F. Keesing, K. Oggenfuss, R. S. Ostfeld. 2015. Accelerated phenology of blacklegged ticks under climate warming. *Phil. Trans. R. Soc. B* **370** 20130556; DOI: 10.1098/rstb.2013.0556.
- LoGiudice, K., R. S. Ostfeld, K. A. Schmidt, and F. Keesing. 2003. The ecology of infectious disease: Effects of host diversity and community composition on Lyme disease risk. *Proceedings of the National Academy of Sciences of the United States of America* **100**:567-571.
- Lounibos, L. P. 2002. Invasions by insect vectors of human disease. *Annual Review of Entomology* **47**:233-266.
- Luckhart, S., G. R. Mullen, L. A. Durden, and J. C. Wright. 1992. *Borrelia* sp. in ticks recovered from white tailed deer in Alabama. *Journal of Wildlife Diseases* **28**:449-452.
- Luis, A. D., R. J. Douglass, J. N. Mills, and O. N. Bjørnstad. 2010. The effect of seasonality, density and climate on the population dynamics of Montana deer mice, important reservoir hosts for Sin Nombre hantavirus. *Journal of Animal Ecology* **79**:462-470.
- MacNeil, A., T. G. Ksiazek, and P. E. Rollin. 2011. Hantavirus pulmonary syndrome, United States, 1993-2009. *Emerging Infectious Diseases* **17**:1195-1201.



Title: TOS Science Design for Vectors and Pathogens		Date: 04/06/2022
NEON Doc. #: NEON.DOC.000911	Author: Y. Springer	Revision: C

- Marano, N., P. M. Arguin, and M. Pappaioanou. 2007. Impact of globalization and animal trade on infectious disease ecology. *Emerging Infectious Diseases* **13**:1807-1809.
- Matisoo-Smith, E., R. M. Roberts, G. J. Irwin, J. S. Allen, D. Penny, and D. M. Lambert. 1998. Patterns of prehistoric human mobility in Polynesia indicated by mtDNA from the Pacific rat. *Proceedings of the National Academy of Sciences of the United States of America* **95**:15145-15150.
- McNeill, J. R. 2010. *Mosquito Empires: Ecology and War in the Greater Caribbean, 1620-1914*. Cambridge University Press, Cambridge.
- Mead, P. S., C. Nelson, A. Hinckley, S. Hook, K. Kugeler, A. Perea, and B. Beard. 2013. Estimating the public health burden of Lyme disease in the United States. *International Conference on Lyme Borreliosis and Other Tick-Borne Diseases*, Boston, MA.
- Medlock, J. M., K. M. Hansford, A. Bormane, M. Derdakova, A. Estrada-Pena, J. C. George, I. Golovljova, T. G. T. Jaenson, J. K. Jensen, P. M. Jensen, M. Kazimirova, J. A. Oteo, A. Papa, K. Pfister, O. Plantard, S. E. Randolph, A. Rizzoli, M. M. Santos-Silva, H. Sprong, L. Vial, G. Hendrickx, H. Zeller, and W. Van Bortel. 2013. Driving forces for changes in geographical distribution of *Ixodes ricinus* ticks in Europe. *Parasites & Vectors* **6**:1-11.
- Meeraus, W. H., J. S. Armistead, and J. R. Arias. 2008. Field comparison of novel and gold standard traps for collecting *Aedes albopictus* in Northern Virginia. *Journal of the American Mosquito Control Association* **24**:244-248.
- Meerburg, B. G., G. R. Singleton, and A. Kijlstra. 2009. Rodent-borne diseases and their risks for public health. *Critical Reviews in Microbiology* **35**:221-270.
- Mills, J. N. 2006. Biodiversity loss and emerging infectious disease: An example from the rodent-borne hemorrhagic fevers. *Biodiversity* **7**:9-17.
- Mills, J. N., B. R. Amman, and G. E. Glass. 2010. Ecology of hantaviruses and their hosts in North America. *Vector-Borne and Zoonotic Diseases* **10**:563-574.
- Mills, J. N., J. E. Childs, T. G. Ksiazek, C. J. Peters, and W. M. Velleca. 1995. Methods for trapping and sampling small mammals for virologic testing. U.S. Department of Health and Human Services, Center for Disease Control and Prevention, Atlanta, GA.
- Milne, A. 1943. The comparison of sheep tick populations (*Ixodes ricinus* L). *Annals of Applied Biology* **30**:240-250.
- Mitchell, C. E. and A. G. Power. 2003. Release of invasive plants from fungal and viral pathogens. *Nature* **421**:625-627.
- Moore, C. G., C. J. Mitchell, R. S. Nasci, T. F. Tsai, C. H. Calisher, A. A. Marfin, P. S. Moore, and D. J. Gubler. 1993. Guidelines for arbovirus surveillance programs in the United States. Department of Health and Human Services. Division of Vector-Borne Infectious Diseases.



Title: TOS Science Design for Vectors and Pathogens		Date: 04/06/2022
NEON Doc. #: NEON.DOC.000911	Author: Y. Springer	Revision: C

- Mordecai, E. A., K. P. Paaijmans, L. R. Johnson, C. Balzer, T. Ben-Horin, E. Moor, A. McNally, S. Pawar, S. J. Ryan, T. C. Smith, and K. D. Lafferty. 2013. Optimal temperature for malaria transmission is dramatically lower than previously predicted. *Ecology Letters* **16**:22-30.
- Morens, D. M., G. K. Folkers, and A. S. Fauci. 2004. The challenge of emerging and re-emerging infectious diseases. *Nature* **430**:242-249.
- Morris, C. D. 1989. Eastern Equine encephalitis. Pages 1-20 *in* T. P. Monath, editor. The arboviruses: epidemiology and ecology. Boca Raton, Fla, CRC Press.
- Mukherjee, N., L. Beati, M. Sellers, L. Burton, S. Adamson, R. G. Robbins, F. Moore, and S. Karim. 2014. Importation of exotic ticks and tick-borne spotted fever group rickettsiae into the United States by migrating songbirds. *Ticks and tick-borne diseases* **5**:127-134.
- Murray, C. J. L., L. C. Rosenfeld, S. S. Lim, K. G. Andrews, K. J. Foreman, D. Haring, N. Fullman, M. Naghavi, R. Lozano, and A. D. Lopez. 2012. Global malaria mortality between 1980 and 2010: a systematic analysis. *Lancet* **379**:413-431.
- Myers, S. S., L. Gaffikin, C. D. Golden, R. S. Ostfeld, K. H. Redford, T. H. Ricketts, W. R. Turner, and S. A. Osofsky. 2013. Human health impacts of ecosystem alternations. *Proceedings of the National Academy of Sciences of the United States of America* **110**:18753-18760.
- Nasci, R. S., K. L. Gottfried, K. L. Burkhalter, V. L. Kulasekera, A. J. Lambert, R. S. Lanciotti, A. R. Hunt, and J. R. Ryan. 2002. Comparison of Vero cell plaque assay, TaqMan (R) reverse transcriptase polymerase chain reaction RNA assay, and VecTest (TM) antigen assay for detection of West Nile virus in field-collected mosquitoes. *Journal of the American Mosquito Control Association* **18**:294-300.
- National Research Council. 2001. Grand challenges in environmental sciences. National Academy Press, Washington, D.C.
- National Research Council. 2003. NEON - Addressing the nation's environmental challenges. National Academies Press, Washington, D.C.
- Naze, F., K. Le Roux, A. Schuffenecker, H. Zeller, F. Staikowsky, P. Grivard, A. Michault, and P. Laurent. 2009. Simultaneous detection and quantitation of chikungunya, dengue and West Nile viruses by multiplex RT-PCR assays and dengue virus typing using high resolution melting. *Journal of Virological Methods* **162**:1-7.
- Needham, G. R. and P. D. Teel. 1991. Off-host physiological ecology of ixodid ticks. *Annual Review of Entomology* **36**:659-681.
- Newhouse, V. F., R. W. Chamberlin, J. G. Johnston Jr, and W. D. Sudia. 1966. Use of dry ice to increase mosquito catches of the CDC miniature light trap. *Mosquito News* **26**:30-35.
- Nichol, S. T., C. F. Spiropoulou, S. Morzunov, P. E. Rollin, T. G. Ksiazek, H. Feldmann, A. Sanchez, J. Childs, S. Zaki, and C. J. Peters. 1993. Genetic identification of a hantavirus associated with an outbreak of acute respiratory illness. *Science* **262**:914-917.



- Ogden, N. H., M. Bigras-Poulin, C. J. O'Callaghan, I. K. Barker, K. Kurtenbach, L. R. Lindsay, and D. F. Charron. 2007. Vector seasonality, host infection dynamics and fitness of pathogens transmitted by the tick *Ixodes scapularis*. *Parasitology* **134**:209-227.
- Ogden, N. H., S. Mechai, and G. Margos. 2013. Changing geographic ranges of ticks and tick-borne pathogens: drivers, mechanisms and consequences for pathogen diversity. *Frontiers in Cellular and Infection Microbiology* **3**.
- Ostfeld, R. S., C. D. Canham, K. Oggenfuss, R. J. Winchcombe, and F. Keesing. 2006. Climate, deer, rodents, and acorns as determinants of variation in Lyme-disease risk. *Plos Biology* **4**:1058-1068.
- Ostfeld, R. S., C. G. Jones, and J. O. Wolff. 1996. Of mice and mast. *Bioscience* **46**:323-330.
- Paaijmans, K. P., S. Blanford, B. H. K. Chan, and M. B. Thomas. 2012. Warmer temperatures reduce the vectorial capacity of malaria mosquitoes. *Biology Letters* **8**:465-468.
- Paddock, C. D. and M. J. Yabsley. 2007. Ecological havoc, the rise of white-tailed deer, and the emergence of *Amblyomma americanum* - associated zoonoses in the United States. Pages 289-324 *Wildlife and Emerging Zoonotic Diseases: The Biology, Circumstances and Consequences of Cross-Species Transmission*. Springer Berlin, Heidelberg.
- Panella, N. A., R. J. Kent Crockett, B. J. Biggerstaff, and N. Komar. 2011. The Centers for Disease Control and Prevention resting trap: A novel device for collecting resting mosquitoes. *Journal of the American Mosquito Control Association* **27**:323-325.
- Parmenter, C. A., T. L. Yates, R. R. Parmenter, J. N. Mills, J. E. Childs, M. L. Campbell, J. L. Dunnum, and J. Milner. 1998. Small mammal survival and trapability in mark-recapture monitoring programs for hantavirus. *Journal of Wildlife Diseases* **34**:1-12.
- Patz, J. A., E. C. Confalonieri, F. P. Amerasinghe, K. B. Chua, P. Daszak, A. D. Hyatt, D. Molyneux, M. Thomson, L. Yameogo, M. M. Lazaro, P. Vasconcelos, Y. Rubio-Palis, D. Campbell-Lendrum, T. Jaenisch, H. Mahamat, C. Mutero, D. Waltner-Toews, and C. Whiteman. 2005. Human health: Ecosystem regulation of infectious diseases. Pages 391-415 in R. Hassan, R. Scholes, and N. Ash, editors. *Millennium Ecosystem Assessment - Ecosystems and Human Well-being: Current State and Trends*. Island Press, Washington, DC.
- Patz, J. A., P. Daszak, G. M. Tabor, A. A. Aguirre, M. Pearl, J. Epstein, N. D. Wolfe, A. M. Kilpatrick, J. Fofopoulos, D. Molyneux, D. J. Bradley, and working group on land use change. 2004. Unhealthy landscapes: Policy recommendations on land use change and infectious disease emergence. *Environmental Health Perspectives* **112**:1092-1098.
- Patz, J. A., T. K. Graczyk, N. Geller, and A. Y. Vittor. 2000a. Effects of environmental change on emerging parasitic diseases. *International Journal for Parasitology* **30**:1395-1405.
- Patz, J. A., M. A. McGeehin, S. M. Bernard, K. L. Ebi, P. R. Epstein, A. Grambsch, D. J. Gubler, P. Reither, I. Romieu, and J. B. Rose. 2000b. The potential health impacts of climate variability and change for



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- the United States: Executive summary of the report of the health sector of the US national assessment. *Environmental Health Perspectives* **108**:367.
- Paweska, J. T. and P. J. van Vuren. 2014. Rift Valley Fever virus: A virus with potential for global emergence. Pages 169-200 *in* N. Johnson, editor. *Role of Animals in Emerging Viral Diseases*. Elsevier, London.
- Perry, B., D. Grace, and K. Sones. 2013. Current drivers and future directions of global livestock disease dynamics. *Proceedings of the National Academy of Sciences of the United States of America* **110**:20871-20877.
- Pfuntner, A. P. 1979. A modified CO₂-baited miniature surveillance trap. *Bulletin of the Society of Vector Ecology* **4**:31-35.
- Phan, T. G., B. Kapusinszky, C. L. Wang, R. K. Rose, H. L. Lipton, and E. L. Delwart. 2011. The fecal viral flora of wild rodents. *Plos Pathogens* **7**:e1002218.
- Pialoux, G., B. A. Gauzere, S. Jaureguiberry, and M. Strobel. 2007. Chikungunya, an epidemic arbovirolosis. *Lancet Infectious Diseases* **7**:319-327.
- Piesman, J., T. N. Mather, J. G. Donahue, J. Levine, J. D. Campbell, S. J. Karakashian, and A. Spielman. 1986. Comparative prevalence of *Babesia microti* and *Borrelia burgdorferi* in four populations of *Ixodes dammini* in eastern Massachusetts. *Acta Tropica* **43**:263-270.
- Pons, J., T. G. Barraclough, J. Gomez-Zurita, A. Cardoso, D. P. Duran, S. Hazell, S. Kamoun, W. D. Sumlin, and A. P. Vogler. 2006. Sequence-based species delimitation for the DNA taxonomy of undescribed insects. *Systematic Biology* **55**:595-609.
- R Core Team. 2013. R: A language and environment for statistical computing. *in* R Foundation for Statistical Computing, editor., Vienna, Austria.
- Randolph, S. E. 2001. The shifting landscape of tick-borne zoonoses: Tick-borne encephalitis and Lyme borreliosis in Europe. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* **356**:1045-1056.
- Reeves, W. C., J. L. Hardy, W. K. Reisen, and M. M. Milby. 1994. Potential effect of global warming on mosquito-borne arboviruses. *Journal of Medical Entomology* **31**:323-332.
- Reisen, W. K., Y. Fang, and V. M. Martinez. 2006. Effects of temperature on the transmission of West Nile virus by *Culex tarsalis* (Diptera : Culicidae). *Journal of Medical Entomology* **43**:309-317.
- Reiter, P. 1983. A portable, battery powered trap for collecting gravid *Culex* mosquitoes. *Mosquito News* **43**:496-498.
- Reiter, P. 2001. Climate change and mosquito-borne disease. *Environmental Health Perspectives* **109**:141-161.
- Rock, M., B. J. Buntain, J. M. Hatfield, and B. Hallgrimsson. 2009. Animal-human connections, "one health," and the syndemic approach to prevention. *Social Science & Medicine* **68**:991-995.



- Rohr, J. R., A. P. Dobson, P. T. J. Johnson, A. M. Kilpatrick, S. H. Paull, T. R. Raffel, D. Ruiz-Moreno, and M. B. Thomas. 2011. Frontiers in climate change-disease research. *Trends in Ecology & Evolution* **26**:270-277.
- Rosenberg, R., M. A. Johansson, A. M. Powers, and B. R. Miller. 2013. Search strategy has influenced the discovery rate of human viruses. *Proceedings of the National Academy of Sciences of the United States of America* **110**:13961-13964.
- Rulison, E. L., I. Kuczaj, G. Pang, G. J. Hickling, J. I. Tsao, and H. S. Ginsberg. 2013. Flagging versus dragging as sampling methods for nymphal I (Acari: Ixodidae). *Journal of Vector Ecology* **38**:163-167.
- Sanchez-Seco, M. P., D. Rosario, E. Quiroz, G. Guzman, and A. Tenorio. 2001. A generic nested-RT-PCR followed by sequencing for detection and identification of members of the alphavirus genus. *Journal of Virological Methods* **95**:153-161.
- Sauer, J. R. and J. A. Hair, editors. 1986. *Morphology, Physiology, and Behavioral Biology of Ticks*. Ellis Horwood Limited, Chichester.
- Savage, H. M., M. S. Godsey, A. Lambert, N. A. Panella, K. L. Burkhalter, J. R. Harmon, R. R. Lash, D. C. Ashley, and W. L. Nicholson. 2013. First detection of Heartland virus (Bunyaviridae: Phlebovirus) from field collected arthropods. *American Journal of Tropical Medicine and Hygiene* **89**:445-452.
- Schimel, D., W. Hargrove, F. Hoffman, and J. MacMahon. 2007. NEON: a hierarchically designed national ecological network. *Frontiers in Ecology and the Environment* **5**:59-59.
- Schimel, D. S., M. Keller, S. Berukoff, R. H. Kao, H. Loescher, H. Powell, T. Kampe, D. Moore, W. Gram, D. T. Barnett, R. Gallery, C. Gibson, K. Goodman, C. L. Meier, S. Parker, L. Pitelka, Y. P. Springer, K. Thibault, and R. Utz. 2011. 2011 science strategy, enabling continental-scale ecological forecasting. National Ecological Observatory Network, Boulder, CO.
- Scott, T. W. and S. C. Weaver. 1989. Eastern Equine encephalomyelitis virus: Epidemiology and evolution of mosquito transmission. *Advances in Virus Research* **37**:277-328.
- Service, M. W. 1993. *Mosquito Ecology: Field Sampling Methods*. Elsevier Applied Science, London.
- Shope, R. E. 1992. Impacts of global climate change on human health: Spread of infectious disease. Pages 363-370 in S. K. Majumdar, L. S. Kalkstein, B. M. Yarnal, E. W. Miller, and L. M. Rosenfeld, editors. *Global Climate Change: Implications, Challenges and Mitigation Measures*. Pennsylvania Academy of Science, Easton.
- Sikes, R. S., W. L. Gannon, and M. Amer Soc. 2011. Guidelines of the American society of mammalogists for the use of wild mammals in research. *Journal of Mammalogy* **92**:235-253.
- Sonenshine, D. E., editor. 1994. *Ecological Dynamics of Tick-borne Zoonoses*. Oxford University Press, Oxford, UK.
- Sonenshine, D. E. and R. M. Roe. 2014. *Biology of Ticks*. Oxford University Press, Oxford, UK.



- Spach, D. H., W. C. Liles, G. L. Campbell, R. E. Quick, D. E. Anderson, and T. R. Fritsche. 1993. Tick-borne diseases in the United States. *New England Journal of Medicine* **329**:936-947.
- Springer, Y. P., L. Eisen, L. Beati, A. M. James, and R. J. Eisen. 2014. Spatial distribution of counties in the continental United States with records of occurrence of *Amblyomma americanum* (Ixodida: Ixodidae). *Journal of Medical Entomology* **51**:342-351.
- Stafford, K. C. 1994. Survival of immature *Ixodes scapularis* (Acari, Ixodidae) at different relative humidities. *Journal of Medical Entomology* **31**:310-314.
- Steere, A. C., S. E. Malawista, J. A. Hardin, S. Ruddy, P. W. Askenase, and W. A. Andiman. 1977. Erythema chronicum migrans and Lyme arthritis - Enlarging clinical spectrum. *Annals of Internal Medicine* **86**:685-698.
- Strange, R. N. and P. R. Scott. 2005. Plant disease: A threat to global food security. Pages 83-116 *Annual Review of Phytopathology*.
- Sudia, W. D. and R. W. Chamberlain. 1962. Battery-operated light trap, an improved model. *Mosquito News* **22**:126-129.
- Sutherst, R. W. 2004. Global change and human vulnerability to vector-borne diseases. *Clinical Microbiology Reviews* **17**:136-173.
- Swabe, J., editor. 1999. *Animals, Disease, and Human Society: Human-Animal Relations and the Rise of Veterinary Medicine*. Routledge, London.
- Swann, D. E., A. J. Kuenzi, M. L. Morrison, and S. DeStefano. 1997. Effects of sampling blood on survival of small mammals. *Journal of Mammalogy* **78**:908-913.
- Tatem, A. J., S. I. Hay, and D. J. Rogers. 2006. Global traffic and disease vector dispersal. *Proceedings of the National Academy of Sciences of the United States of America* **103**:6242-6247.
- Taylor, L. H., S. M. Latham, and M. E. J. Woolhouse. 2001. Risk factors for human disease emergence. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* **356**:983-989.
- Teel, P. D., H. R. Ketchum, D. E. Mock, R. E. Wright, and O. F. Strey. 2010. The gulf coast tick: A review of the life history, ecology, distribution, and emergence as an arthropod of medical and veterinary importance. *Journal of Medical Entomology* **47**:707-722.
- Thibault, K. M., S. R. Supp, M. Giffin, E. P. White, and S. K. M. Ernest. 2011. Species composition and abundance of mammalian communities. *Ecology* **92**:2316.
- Thompson, R., A. Lymbery, and A. Smith. 2010. Parasites, emerging disease and wildlife conservation. *International Journal for Parasitology* **40**:1163-1170.
- Thompson, S. K. 1990. Adaptive cluster sampling. *Journal of the American Statistical Association* **85**:1050-1059.



- Turell, M. J., D. J. Dohm, M. R. Sardelis, M. L. O Guinn, T. G. Andreadis, and J. A. Blow. 2005. An update on the potential of North American mosquitoes (Diptera : Culicidae) to transmit West Nile virus. *Journal of Medical Entomology* **42**:57-62.
- Vail, S. G. and G. Smith. 1998. Air temperature and relative humidity effects on behavioral activity of blacklegged tick (Acari : Ixodidae) nymphs in New Jersey. *Journal of Medical Entomology* **35**:1025-1028.
- van Riper, C., S. G. van Riper, M. L. Goff, and M. Laird. 1986. The epizootiology and ecological significance of malaria in Hawaiian land birds. *Ecological Monographs* **56**:327-344.
- Watts, D. M., D. S. Burke, B. A. Harrison, R. E. Whitmire, and A. Nisalak. 1987. Effect of temperature on the vector efficiency of *Aedes aegypti* for dengue-2 virus. *American Journal of Tropical Medicine and Hygiene* **36**:143-152.
- Weaver, S. C. and W. K. Reisen. 2010. Present and future arboviral threats. *Antiviral research* **85**:328-345.
- Weiss, R. A. and A. J. McMichael. 2004. Social and environmental risk factors in the emergence of infectious diseases. *Nature Medicine* **10**:S70-S76.
- Williams, G. M. and J. B. Gingrich. 2007. Comparison of light traps, gravid traps, and resting boxes for West Nile virus surveillance. *Journal of Vector Ecology* **32**:285-291.
- Wolfe, N. D., P. Daszak, A. M. Kilpatrick, and D. S. Burke. 2005. Bushmeat hunting deforestation, and prediction of zoonoses emergence. *Emerging Infectious Diseases* **11**:1822-1827.
- Wolfe, N. D., C. P. Dunavan, and J. Diamond. 2007. Origins of major human infectious diseases. *Nature* **447**:279-283.
- Woolhouse, M. E. J. and S. Gowtage-Sequeria. 2005. Host range and emerging and reemerging pathogens. *Emerging Infectious Diseases* **11**:1842-1847.
- Yates, T. L., J. N. Mills, C. A. Parmenter, T. G. Ksiazek, R. R. Parmenter, J. R. Vande Castle, C. H. Calisher, S. T. Nichol, K. D. Abbott, J. C. Young, M. L. Morrison, B. J. Beaty, J. L. Dunnum, R. J. Baker, J. Salazar-Bravo, and C. J. Peters. 2002. The ecology and evolutionary history of an emergent disease: Hantavirus pulmonary syndrome. *Bioscience* **52**:989-998.



APPENDIX A PRELIMINARY TICK DATA FROM NEON SITES

Table A1. Number of ticks captured by site in 2016 and 2017. NS indicates the site was not sampled.

Domain	Domain Name	Site ID	Site Name	Total Number of Ticks Captured	
				2016	2017
D01	Northeast	BART	Bartlett Experimental Forest	0	0
D01	Northeast	HARV	Harvard Forest	983	710
D02	Mid-Atlantic	BLAN	Blandy Experimental Farm	894	1,301
D02	Mid-Atlantic	SCBI	Smithsonian Conservation Biology Institute	3,132	2,464
D02	Mid-Atlantic	SERC	Smithsonian Environmental Research Center	4,266	3,884
D03	Southeast	DSNY	Disney Wilderness Preserve	4	4
D03	Southeast	JERC	Jones Ecological Research Center	1	0
D03	Southeast	OSBS	Ordway-Swisher Biological Station	4,988	4,427
D04	Atlantic Neotropical	GUAN	Guanica Forest	0	0
D04	Atlantic Neotropical	LAJA	Lajas Experimental Station	0	0
D05	Great Lakes	STEI	Steigerwaldt Land Services	10	36
D05	Great Lakes	TREE	Treehaven	1,081	1,138
D05	Great Lakes	UNDE	UNotre Dame Environmental Research Center	33	3
D06	Prairie Peninsula	KONA	Konza Prairie Biological Station - Agricultural	NS	29
D06	Prairie Peninsula	KONZ	Konza Prairie Biological Station	1,836	1,241
D06	Prairie Peninsula	UKFS	The University of Kansas Field Station	26,836	49,414
D07	Appalachians & Cumberland Plateau	GRSM	Great Smoky Mountains National Park	3	5
D07	Appalachians & Cumberland Plateau	MLBS	Mountain Lake Biological Station	NS	NS
D07	Appalachians & Cumberland Plateau	ORNL	Oak Ridge National Laboratory	7,018	6,423
D08	Ozarks Complex	DELA	Dead Lake	16	21
D08	Ozarks Complex	LENO	Choctaw National Wildlife Refuge (Lenoir)	1,468	1,321
D08	Ozarks Complex	TALL	Talladega National Forest	5,854	2,416
D09	Northern Plains	DCFS	Dakota Coteau Field School	0	17
D09	Northern Plains	NOGP	Northern Great Plains Research Laboratory	3	12
D09	Northern Plains	WOOD	Woodworth	56	204
D10	Central Plains	CPER	Central Plains Experimental Range	0	0
D10	Central Plains	RMNP	Rocky Mountain National Park, CASTNET	0	0
D10	Central Plains	STER	North Sterling, Co	0	0
D11	Southern Plains	CLBJ	CLBJ National Grassland	1	12
D11	Southern Plains	OAES	Oklahoma Agricultural Experimental Station	14	47
D12	Northern Rockies	YELL	Yellowstone National Park	NS	NS
D13	Southern Rockies & Colorado Plateau	MOAB	Moab	0	0
D13	Southern Rockies & Colorado Plateau	NIWO	Niwot Ridge Mountain Research Station	0	0
D14	Desert Southwest	JORN	Jornada LTER	0	0
D14	Desert Southwest	SRER	Santa Rita Experimental Range	0	0
D15	Great Basin	ONAQ	Onaqui	0	1



Domain	Domain Name	Site ID	Site Name	Total Number of Ticks Captured	
				2016	2017
D16	Pacific Northwest	ABBY	Abby Road	10	21
D16	Pacific Northwest	WREF	Wind River Experimental Forest	NS	NS
D17	Pacific Southwest	SJER	San Joaquin Experimental Range	0	0
D17	Pacific Southwest	SOAP	Soaproot Saddle	NS	NS
D17	Pacific Southwest	TEAK	Lower Teakettle	NS	NS
D18	Tundra	BARR	Barrow Environmental Observatory	NS	0
D18	Tundra	TOOL	Toolik Lake	NS	0
D19	Taiga	BONA	Caribou - Poker Creeks Research Watershed	NS	0
D19	Taiga	DEJU	Delta Junction	0	0
D19	Taiga	HEAL	Healy	0	0



APPENDIX B PRELIMINARY RODENT-BORNE PATHOGEN DATA FROM NEON SITES

Table B1. Number of rodent blood samples tested (total N) and the percentage of samples that tested positive for *Hantavirus* (% positive) by site by year. See Table A1 for key to site codes. NS indicates the site was not sampled.

siteID	value	2014	2015	2016	2017
ABBY	total N	NS	NS	19	62
	% positive	NS	NS	0	0
BART	total N	253	2	188	89
	% positive	0	0	5.85	0
BLAN	total N	3	61	30	20
	% positive	0	0	0	0
BONA	total N	NS	NS	NS	101
	% positive	NS	NS	NS	0
CLBJ	total N	NS	NS	71	202
	% positive	NS	NS	8.45	2.48
CPER	total N	36	1	23	7
	% positive	0	0	0	14.29
DCFS	total N	NS	NS	NS	13
	% positive	NS	NS	NS	0
DEJU	total N	NS	NS	4	94
	% positive	NS	NS	0	0
DELA	total N	NS	NS	26	65
	% positive	NS	NS	0	0
DSNY	total N	8	0	63	80
	% positive	0	0	0	0
GRSM	total N	NS	NS	25	0
	% positive	NS	NS	0	0
GUAN	total N	NS	NS	41	9
	% positive	NS	NS	0	0
HARV	total N	229	8	199	184
	% positive	0	0	2.01	0
HEAL	total N	NS	3	36	70
	% positive	NS	0	0	0
JERC	total N	181	0	146	98
	% positive	0	0	0.68	0
JORN	total N	NS	17	20	8
	% positive	NS	5.88	0	0
KONZ	total N	NS	179	456	105
	% positive	NS	0.56	1.1	1.9
LAJA	total N	NS	NS	100	0
	% positive	NS	NS	0	0
LENO	total N	NS	NS	20	18
	% positive	NS	NS	5	0
MOAB	total N	NS	NS	137	37
	% positive	NS	NS	9.49	8.11
NIWO	total N	NS	177	39	58
	% positive	NS	3.39	0	0
NOGP	total N	NS	NS	6	4



siteID	value	2014	2015	2016	2017
	% positive	NS	NS	0	0
OAES	total N	NS	62	131	7
	% positive	NS	0	3.82	0
ONAQ	total N	190	0	105	38
	% positive	2.63	0	8.57	2.63
ORNL	total N	37	2	50	0
	% positive	0	0	0	0
OSBS	total N	54	0	61	71
	% positive	0	0	0	0
RMNP	total N	NS	NS	NS	79
	% positive	NS	NS	NS	1.27
SCBI	total N	87	133	96	39
	% positive	9.2	9.02	8.33	7.69
SERC	total N	1	29	41	15
	% positive	0	6.9	9.76	0
SJER	total N	NS	NS	NS	64
	% positive	NS	NS	NS	0
SRER	total N	NS	NS	259	187
	% positive	NS	NS	1.54	0
STEI	total N	2	0	235	211
	% positive	0	0	0	0
STER	total N	48	0	48	151
	% positive	4.17	0	4.17	5.96
TALL	total N	89	0	62	116
	% positive	2.25	0	1.61	0.86
TOOL	total N	NS	NS	NS	2
	% positive	NS	NS	NS	0
TREE	total N	0	0	79	157
	% positive	0	0	0	0
UKFS	total N	1	162	272	57
	% positive	0	0.62	0.74	0
UNDE	total N	258	0	198	303
	% positive	0	0	0	0
WOOD	total N	533	0	27	51
	% positive	3.56	0	0	3.92



APPENDIX C CODE FOR POWER ANALYSES

R Code written by P. Duffy (Neptune Inc.) for NEON

Code for functions

```
# Function to compute the sample size for detecting a linear trend when the response
# is binomial for samples to be taken uniformly-spaced in time
# Result is the per-sampling-period sample size
# p0 = baseline (year 0) probability
# b1 = annual increase in log-odds [parameter of interest for inference]
# maxtime = total number of years of sampling,
# sampfreq = time interval between samples (in years), e.g. 1 = once per year, 0.5 = twice per year, etc
# timerange = time interval at which temporal correlation drops to exp(-1)
# note: if timerange = 0, then no temporal correlation is included
# sigLevel = significance level or type I error rate
# power = power (1 minus acceptable type II error rate) to detect trend at the level specified by b1
# pooled = logical. If True, multiple organisms are homogenized prior to analysis
# poolsize = number of organisms in the pool if pooled = T
```

```
binomialsampsize = function( p0, b1, maxtime, sampfreq, timerange=0,
                             sigLevel=0.05, power=0.9, pooled=TRUE,
                             poolsize=50){
```

```
# p0 is vector of baseline rates or probabilities
# b1 is vector of trend values
# sigLevel = significance level or type I error rate
# power = power (1 minus acceptable type II error rate) to detect trend at the level specified by b1
# maxtime is a vector of different maximum lengths of time (years)
# sampfreq is the number of sampling intervals per unit of time 1=once per year,
# 0.5=twice per year, 0.3333=three times per year, 0.25=four times per year,
# 0.07692=every 4 weeks, 0.03846=every 2 weeks)
# timerange is an exponential-decay correlation parameter
# pooled indicates whether the samples are pooled
# poolsize is the number of samples in each pool
# p0 = 0.1; b1 = 0.05; maxtime = 10; sampfreq = 0.5; timerange=0; sigLevel=0.05;
# power=0.9; pooled = TRUE; poolsize = 50
# Convert p0 to b0
if(!pooled){
  b0=log(p0/(1-p0))
}
if(pooled){ # See Farrington article for derivation.
  b0 = log((-1)*log(1-p0))
# An approximation of this for small p0 is b0 = log(p0)
}
# Precalculate re-used quantity
zpre=qnorm(1-sigLevel)+qnorm(power)
# Set up storage
```



```
narr=array(dim=c(length(sampfreq),length(maxtime),length(p0),length(b1)))
# Loop through different specified sampling frequencies
for(i in 1:length(sampfreq)){
# Loop through different specified time limits
for(j in 1:length(maxtime)){
# Set up sequence of sampling times
tm=seq(0,maxtime[j],sampfreq[i])
# Compute temporal correlation matrix
if( timerange==0){
cormat=diag(1,length(tm))
} else {
tmp=as.matrix(dist(tm,upper=TRUE,diag=TRUE))
cormat=exp(-tmp/timerange)
}
# Construct design matrix for regression
if(!pooled) xmat=cbind(1,tm)
if(pooled) xmat=cbind(log(poolsize),1,tm)
# Loop through different specified intercept terms
for(k in 1:length(p0)){
# Loop through different specified slop terms
for(m in 1:length(b1)){
# Compute regression curve
if(!pooled) regrFits=as.vector(xmat%%c(b0[k],b1[m]))
if(pooled) regrFits=as.vector(xmat%%c(1,b0[k],b1[m]))
# Convert to sampling mean
if(!pooled){
tmp = exp(regrFits)
p = tmp/(1+tmp)
}
if(pooled){
p=1-exp((-1)*exp(regrFits))
}
# Construct variance and standard deviation matrix
wmat=diag(p*(1-p))
wrootmat=diag(sqrt(p*(1-p)))
# Compute standard error of estimate based on
# sample size of 1
if(!pooled){
xtxinvsolve=solve(t(xmat)%%wmat%%xmat)
ses=xtxinvsolve%%t(xmat)%%wrootmat%%cormat%%wrootmat%%xmat%%xtxinvsolve
}
if(pooled){
xtxinvsolve=solve(t(xmat[-1])%%wmat%%xmat[-1])
ses=xtxinvsolve%%t(xmat[-1])%%wrootmat%%cormat%%wrootmat%%xmat[-1]%%xtxinvsolve
}
# Compute required sample size
```



```
narr[i,j,k,m] = ceiling((zpre/b1[m])^2*ses[2,2])
}
}
}
}
# Label output and return
dimnames(narr)=list(sampfreq,maxtime,p0,b1)
return(narr)
}
```

```
#####
```

```
# Function to compute the sample size for detecting a linear
# trend when the response is negative binomial for samples
# to be taken uniformly-spaced in time
# Result is the per-sampling-period number of "successes" to be sampled
# m0 = the mean under the baseline condition (year 0)
# p1 = annual percent increase (decrease) in the mean [parameter of
# interest for inference], related to slope: b1 = log( 1 + p1/100 )
# dispersion = dispersion parameter of negative binomial
# variance = mu + mu^2/dispersion [higher dispersion -> closer to poisson]
# maxtime = total number of years of sampling
# sampfreq = time interval between samples (in years)
# scale = scale parameter for negative binomial (poisson over-dispersal)
# timerange = time interval at which temporal correlation drops to exp(-1)
# if timerange=0, then no temporal correlation is included
# sigLevel = significance level or type I error rate
# power = power (1 minus acceptable type II error rate) to detect trend at the level specified by b1
```

```
negbinomialsampsize = function( m0, p1, dispersion, maxtime, sampfreq,
timerange=0, sigLevel=0.05, power=0.9 ){
# Precalculate re-used quantity
zpre=qnorm(1-sigLevel)+qnorm(power)
# Set up storage
narr=array(dim=c(length(sampfreq),length(maxtime),length(dispersion),
length(m0),length(p1)))
# Loop through different specified sampling frequencies
for(i in 1:length(sampfreq)){
# Loop through different specified time limits
for(j in 1:length(maxtime)){
# Set up sequence of sampling times
tm=seq(0,maxtime[j],sampfreq[i])
# Compute temporal correlation matrix
if(timerange==0){
cormat=diag(1,length(tm))
}else{
tmp=as.matrix(dist(tm,upper=TRUE,diag=TRUE))
```



```
cormat=exp(-tmp/timerange)
}
# Construct design matrix for regression
xmat=cbind(1,tm)
# Loop through different specified dispersion parameters
for(d in 1:length(dispersion)){
# Loop through different specified intercept terms
for(k in 1:length(m0)){
# Loop through different specified slop terms
for(m in 1:length(p1)){
# Compute regression curve
logmu=as.vector(xmat**%c(log(m0[k]),log(1+p1[m]/100)))
# Construct variance and standard deviation matrix
mu=exp(logmu)
wmat=diag(mu)
wrootmat=diag(sqrt(dispersion[d]*mu))
# Compute standard error of estimate based on
# sample size of 1
txxinv=solve(t(xmat)**%wmat**%xmat)
ses=txxinv**%t(xmat)**%wrootmat**%cormat**%wrootmat**%xmat**%txxinv
# Compute required sample size
narr[i,j,d,k,m]= ceiling((zpre/log(1+p1[m]/100))^2*ses[2,2])
}
}
}
}
}
# Label output and return
dimnames(narr) = list(sampfreq,maxtime,dispersion,m0,p1)
return(narr)
}
```

Additional code to add for tick-borne pathogens

```
# Code to generate tables
# csv files will be created in the current R directory
# Create tables

m0=c(2,6)
# for m0 we are assuming a max testing number of 100 and an associated conservative
# testing number of 40 and so for the starting #values an m0 of 2 translates to 5% and
# an m0 of 6 translates to 15%
p1=c(0.25,0.5,0.75,1)
# for p1 we are assuming annual prevalence increases of 0.25%, 0.5%, 0.75%, and 1%
maxtime=c(10,25)
sampfreq=c(1,0.5,0.3333,0.25,0.1154,0.05769)
```



```
nbin1s = negbinomialsampsize(m0,p1,2,maxtime,sampfreq)[,,1,,]
cat(", ",file="tick_negbi_notemp.csv")
for(i in 1:length(m0)){
cat("Baseline Mean = ",m0[i],", ",file="tick_negbi_notemp.csv",append=T)
for(j in 2:length(maxtime)){
cat(", ",file="tick_negbi_notemp.csv",append=T)
}
}
cat("\n",file="tick_negbi_notemp.csv",append=T)
for(i in 1:length(p1)){
cat(", ",file="tick_negbi_notemp.csv",append=T)
for(j in 1:length(m0)){
cat("Annual % Increase = ",p1[i],", ",file="tick_negbi_notemp.csv",append=T)
for(k in 2:length(maxtime)){
cat(", ",file="tick_negbi_notemp.csv",append=T)
}
}
cat("\n",file="tick_negbi_notemp.csv",append=T)
for(j in 1:length(m0)){
for(k in 1:length(maxtime)){
cat(maxtime[k],", ",file="tick_negbi_notemp.csv",append=T)
}
}
cat("\n",file="tick_negbi_notemp.csv",append=T)
for(m in 1:length(sampfreq)){
cat(sampfreq[m],", ",file="tick_negbi_notemp.csv",append=T)
for(j in 1:length(m0)){
for(k in 1:length(maxtime)){
cat(nbin1s[m,k,j,i],", ",file="tick_negbi_notemp.csv",append=T)
}
}
}
cat("\n",file="tick_negbi_notemp.csv",append=T)
}
}
```

```
m0=c(2,6)
# for m0 we are assuming a max testing number of 100 and an associated conservative testing number
of 40 and so for the starting values an m0 of 2 translates to 5% and an m0 of 6 translates to 15%
p1=c(0.25,0.5,0.75,1)
# for p1 we are assuming annual prevalence increases of 0.25%, 0.5%, 0.75%, and 1%
maxtime=c(10,25)
sampfreq=c(1,0.5,0.3333,0.25,0.1154,0.05769)
nbin1c = negbinomialsampsize(m0,p1,2,maxtime,sampfreq,timerange=0.5)[,,1,,]
cat(", ",file="tick_negbi_withtemp.csv")
for(i in 1:length(m0)){
cat("Baseline Mean = ",m0[i],", ",file="tick_negbi_withtemp.csv",append=T)
```



```
for(j in 2:length(maxtime)){
cat(", ",file="tick_negbi_withtemp.csv",append=T)
}
}
cat("\n",file="tick_negbi_withtemp.csv",append=T)
for(i in 1:length(p1)){
cat(", ",file="tick_negbi_withtemp.csv",append=T)
for(j in 1:length(m0)){
cat("Annual% Increase = ",p1[i],", ",file="tick_negbi_withtemp.csv",append=T)
for(k in 2:length(maxtime)){
cat(", ",file="tick_negbi_withtemp.csv",append=T)
}
}
cat("\n",file="tick_negbi_withtemp.csv",append=T)
for(j in 1:length(m0)){
for(k in 1:length(maxtime)){
cat(maxtime[k],", ",file="tick_negbi_withtemp.csv",append=T)
}
}
cat("\n",file="tick_negbi_withtemp.csv",append=T)
for(m in 1:length(sampfreq)){
cat(sampfreq[m],", ",file="tick_negbi_withtemp.csv",append=T)
for(j in 1:length(m0)){
for(k in 1:length(maxtime)){
cat(nbin1c[m,k,j,i],", ",file="tick_negbi_withtemp.csv",append=T)
}
}
}
cat("\n",file="tick_negbi_withtemp.csv",append=T)
}
}
```

Additional code to add for mosquito-borne pathogens

```
# Code to generate tables
# csv files will be created in the current R directory
# Create tables
p0=c(.001,.005,.01)
b1=c(.005,.01,.015,.02,.03,.04,.05)
maxtime=c(10,25)
sampfreq=c(1,0.5,0.3333,0.25,0.07692,0.03846)
binom = binomialsampsize(p0,b1,maxtime,sampfreq,timerange=0.5)
cat(", ",file="Mosquito_withtemp.csv")
for(i in 1:length(p0)){
cat("Baseline Probability = ",p0[i],", ",file="Mosquito_withtemp.csv",append=T)
for(j in 2:length(maxtime)){
cat(", ",file="Mosquito_withtemp.csv",append=T)
```



```
}  
}  
cat("\n",file="Mosquito_withtemp.csv",append=T)  
for(i in 1:length(b1)){  
  cat(", ",file="Mosquito_withtemp.csv",append=T)  
  for(j in 1:length(p0)){  
    cat("beta1 = ",b1[i],", ",file="Mosquito_withtemp.csv",append=T)  
    for(k in 2:length(maxtime)){  
      cat(", ",file="Mosquito_withtemp.csv",append=T)  
    }  
  }  
  cat("\n",file="Mosquito_withtemp.csv",append=T)  
  for(j in 1:length(p0)){  
    for(k in 1:length(maxtime)){  
      cat(maxtime[k],", ",file="Mosquito_withtemp.csv",append=T)  
    }  
  }  
  cat("\n",file="Mosquito_withtemp.csv",append=T)  
  for(m in 1:length(sampfreq)){  
    cat(sampfreq[m],", ",file="Mosquito_withtemp.csv",append=T)  
    for(j in 1:length(p0)){  
      for(k in 1:length(maxtime)){  
        cat(binom[m,k,j,i],", ",file="Mosquito_withtemp.csv",append=T)  
      }  
    }  
  }  
  cat("\n",file="Mosquito_withtemp.csv",append=T)  
}  
  
# Code to generate tables  
# csv files will be created in the current R directory  
# Create tables  
p0=c(.001,.005,.01)  
b1=c(.005,.01,.015,.02,.03,.04,.05)  
maxtime=c(10,25)  
sampfreq=c(1,0.5,0.3333,0.25,0.07692,0.03846)  
binom = binomialsampsize(p0,b1,maxtime,sampfreq,)  
cat(", ",file="Mosquito_notemp.csv")  
for(i in 1:length(p0)){  
  cat("Baseline Probability = ",p0[i],", ",file="Mosquito_notemp.csv",append=T)  
  for(j in 2:length(maxtime)){  
    cat(", ",file="Mosquito_notemp.csv",append=T)  
  }  
}  
cat("\n",file="Mosquito_notemp.csv",append=T)  
for(i in 1:length(b1)){
```



```

cat("",file="Mosquito_notemp.csv",append=T)
for(j in 1:length(p0)){
cat("beta1 = ",b1[j],"",file="Mosquito_notemp.csv",append=T)
for(k in 2:length(maxtime)){
cat("",file="Mosquito_notemp.csv",append=T)
}
}
cat("\n",file="Mosquito_notemp.csv",append=T)
for(j in 1:length(p0)){
for(k in 1:length(maxtime)){
cat(maxtime[k],"",file="Mosquito_notemp.csv",append=T)
}
}
cat("\n",file="Mosquito_notemp.csv",append=T)
for(m in 1:length(sampfreq)){
cat(sampfreq[m],"",file="Mosquito_notemp.csv",append=T)
for(j in 1:length(p0)){
for(k in 1:length(maxtime)){
cat(binom[m,k,j,i],"",file="Mosquito_notemp.csv",append=T)
}
}
}
cat("\n",file="Mosquito_notemp.csv",append=T)
}
}

```

Additional code to add for rodent-borne pathogens

```

m0=c(1,4)
# for m0 we are assuming a catch rate per bout of 40 animals and so for the starting values
# an m0 of 1 translates to 2.5% and an m0 # of 4 translates to 10%
p1=c(0.25,0.5,0.75,1)
# for p1 we are assuming annual prevalence increases of 0.25%, 0.5%, 0.75%, and 1%
maxtime=c(10,25)
sampfreq=c(1,0.5,0.3333,0.25,0.1539,0.07692)
nbin1s = negbinomialsampsize(m0,p1,2,maxtime,sampfreq)[,,1,,]
cat("",file="rodent_negbi_notemp.csv")
for(i in 1:length(m0)){
cat("Baseline Mean = ",m0[i],"",file="rodent_negbi_notemp.csv",append=T)
for(j in 2:length(maxtime)){
cat("",file="rodent_negbi_notemp.csv",append=T)
}
}
cat("\n",file="rodent_negbi_notemp.csv",append=T)
for(i in 1:length(p1)){
cat("",file="rodent_negbi_notemp.csv",append=T)
for(j in 1:length(m0)){

```



```

cat("Annual % Increase = ",p1[i],",",file="rodent_negbi_notemp.csv",append=T)
for(k in 2:length(maxtime)){
cat(", ",file="rodent_negbi_notemp.csv",append=T)
}
}
cat("\n",file="rodent_negbi_notemp.csv",append=T)
for(j in 1:length(m0)){
for(k in 1:length(maxtime)){
cat(maxtime[k],",",file="rodent_negbi_notemp.csv",append=T)
}
}
cat("\n",file="rodent_negbi_notemp.csv",append=T)
for(m in 1:length(sampfreq)){
cat(sampfreq[m],",",file="rodent_negbi_notemp.csv",append=T)
for(j in 1:length(m0)){
for(k in 1:length(maxtime)){
cat(nbin1s[m,k,j,i],",",file="rodent_negbi_notemp.csv",append=T)
}
}
}
cat("\n",file="rodent_negbi_notemp.csv",append=T)
}
}
m0=c(1,4)
# for m0 we are assuming a catch rate per bout of 40 animals and so for the
# starting values an m0 of 1 translates to 2.5% and an m0 of 4 translates to 10%
p1=c(0.25,0.5,0.75,1)
# for p1 we are assuming annual prevalence increases of 0.25%, 0.5%, 0.75%, and 1%
maxtime=c(10,25)
sampfreq=c(1,0.5,0.3333,0.25,0.1539,0.07692)
nbin1c = negbinomialsampsize(m0,p1,2,maxtime,sampfreq,timerange=0.5)[,1,,]
cat(", ",file="rodent_negbi_withtemp.csv")
for(i in 1:length(m0)){
cat("Baseline Mean = ",m0[i],",",file="rodent_negbi_withtemp.csv",append=T)
for(j in 2:length(maxtime)){
cat(", ",file="rodent_negbi_withtemp.csv",append=T)
}
}
cat("\n",file="rodent_negbi_withtemp.csv",append=T)
for(i in 1:length(p1)){
cat(", ",file="rodent_negbi_withtemp.csv",append=T)
for(j in 1:length(m0)){
cat("Annual % Increase = ",p1[i],",",file="rodent_negbi_withtemp.csv",append=T)
for(k in 2:length(maxtime)){
cat(", ",file="rodent_negbi_withtemp.csv",append=T)
}
}
}

```



```
cat("\n",file="rodent_negbi_withtemp.csv",append=T)
for(j in 1:length(m0)){
  for(k in 1:length(maxtime)){
    cat(maxtime[k],",",file="rodent_negbi_withtemp.csv",append=T)
  }
}
cat("\n",file="rodent_negbi_withtemp.csv",append=T)
for(m in 1:length(sampfreq)){
  cat(sampfreq[m],",",file="rodent_negbi_withtemp.csv",append=T)
  for(j in 1:length(m0)){
    for(k in 1:length(maxtime)){
      cat(nbin1c[m,k,j,i],",",file="rodent_negbi_withtemp.csv",append=T)
    }
  }
}
cat("\n",file="rodent_negbi_withtemp.csv",append=T)
}
}
```