

<i>Title:</i> NEON User Guide to Aquatic Macroinvertebrate Collection (NEON.DP1.20120)	<i>Date:</i> 12/17/2017
<i>Author:</i> Tanya Chesney	<i>Revision:</i> A

NEON USER GUIDE TO AQUATIC MACROINVERTEBRATE COLLECTION (NEON.DP1.20120)

PREPARED BY	ORGANIZATION	DATE
Tanya Chesney	DPS	12/17/2017
Stephanie Parker	AQU	12/17/2017
Caren Scott	AQU	12/17/2017

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CHANGE RECORD

REVISION	DATE	DESCRIPTION OF CHANGE
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1 DESCRIPTION

1.1 Purpose

This document provides an overview of the data included in this NEON Level 1 data product, the quality controlled product generated from raw Level 0 data, and associated metadata. In the NEON data products framework, the raw data collected in the field, for example, the macroinvertebrate stage of development are considered the lowest level (Level 0). Raw data that have been quality checked via the steps detailed herein, as well as simple metrics that emerge from the raw data are considered Level 1 data products.

The text herein provides a discussion of measurement theory and implementation, data product provenance, quality assurance and control methods used, and approximations and/or assumptions made during L1 data creation.

1.2 Scope

This document describes the steps needed to generate the L1 data product Macroinvertebrate collection and associated metadata from input data. This document also provides details relevant to the publication of the data products via the NEON data portal, with additional detail available in the file, NEON Data Variables for Aquatic Macroinvertebrate Collection (NEON.DP1.20120) (AD[04]), provided in the download package for this data product.

This document describes the process for ingesting and performing automated quality assurance and control procedures on the data collected in the field pertaining to AOS Protocol and Procedure: Aquatic Macroinvertebrate Sampling (AD[06]). The raw data that are processed in this document are detailed in the file, NEON Raw Data Validation for Aquatic Macroinvertebrate Collection (NEON.DP0.20120) (AD[03]), provided in the download package for this data product. Please note that raw data products (denoted by ‘DP0’) may not always have the same numbers (e.g., ‘20120’) as the corresponding L1 data product.

2 RELATED DOCUMENTS

2.1 Associated Documents

AD[01]	NEON.DOC.000001	NEON Observatory Design (NOD) Requirements
AD[02]	NEON.DOC.002652	NEON Level 1, Level 2 and Level 3 Data Products Catalog
AD[03]	NEON.DP0.20120.001_dataValidation.csv	NEON Raw Data Validation for Aquatic Macroinvertebrate Collection (NEON.DP0.20120)
AD[04]	NEON.DP1.20120.001_variables.csv	NEON Data Variables for Aquatic Macroinvertebrate Collection (NEON.DP1.20120)
AD[05]	NEON.DOC.001152	NEON Aquatic Sampling Strategy
AD[06]	NEON.DOC.003046	AOS Protocol and Procedure: Aquatic Macroinvertebrate Sampling
AD[07]	NEON.DOC.000008	NEON Acronym List
AD[08]	NEON.DOC.000243	NEON Glossary of Terms
AD[09]	OS_Generic_Transitions.pdf	NEON Algorithm Theoretical Basis Document: OS Generic Transitions
AD[10]	Nicl Language.pdf	NEON's Ingest Conversion Language (NICL) specifications

3 DATA PRODUCT DESCRIPTION

Aquatic macroinvertebrate-related data products include taxonomy, abundance, morphometrics, and give information related to the NEON Grand Challenge area of Biodiversity as well as additional data about the macroinvertebrate community in streams and lakes. These data can be used to assess the health of aquatic ecosystems. Macroinvertebrates are sampled three times per year at each NEON aquatic site (AD[05]). Sampling dates are based on a combination of variables, including hydrology in streams or ice on/ice off dates in lakes, accumulated degree days (temperature), and riparian greenness (phenology). Samples are collected by field personnel, preserved in the field, and sent to expert taxonomists for identification. For additional information see sampling design NEON Aquatic Sampling Strategy (AD[05]) and protocol AOS Protocol and Procedure: Aquatic Macroinvertebrate Sampling (AD[06]).

Data are organized into tables for field data collected by NEON technicians and external lab data returned by the expert taxonomy lab(s). Field data contains metadata on sample time, location, type of habitat and substratum, and the type of sampler used, which determines the benthic area sampled. The lab data includes subsampling information, taxonomic analysis, count and size class data. Lab data are corrected for subsampling, however the data user must use both the lab data and the field data to calculate counts per benthic area of habitat if a quantitative result is desired. See Section 3.9 for suggested calculations.

3.1 Spatial Sampling Design

Benthic invertebrates at NEON aquatic sites (Figure 1) are sampled using a percent-based macrohabitat approach (after Moulton et al. 2002). Habitats sampled focus on riffles, runs, pools, and step pools depending on the percent cover of each habitat within each 1 km-long NEON Aquatic wadeable stream site (NOTE: some NEON sites may be less than 1 km due to permitting restrictions), and benthic-pelagic and littoral samples in lakes and non-wadeable streams. Five samples are collected in the dominant habitat type (wadeable stream) or littoral area (lake and non-wadeable stream), and three samples are collected in the second-most dominant habitat type (wadeable stream) or pelagic area (lakes and non-wadeable stream) for a total of eight samples on a given sampling date at a site.

Samplers used for macroinvertebrate collection are designed to work by disturbing the benthic sediments and catching invertebrates in an attached net or container, while delineating the benthic area sampled for a quantitative result. The sampler type chosen differs depending on the water depth, velocity, and substratum type in the chosen habitat (Hauer and Resh 2006). The collection method may differ depending on the habitat and substrate being sampled, however all samples are collected from the surface of the natural substratum in each habitat using a quantitative sampling method. See AOS Protocol and Procedure: Aquatic Macroinvertebrate Sampling (AD[06]) for additional details on sampling strategy and SOPs.

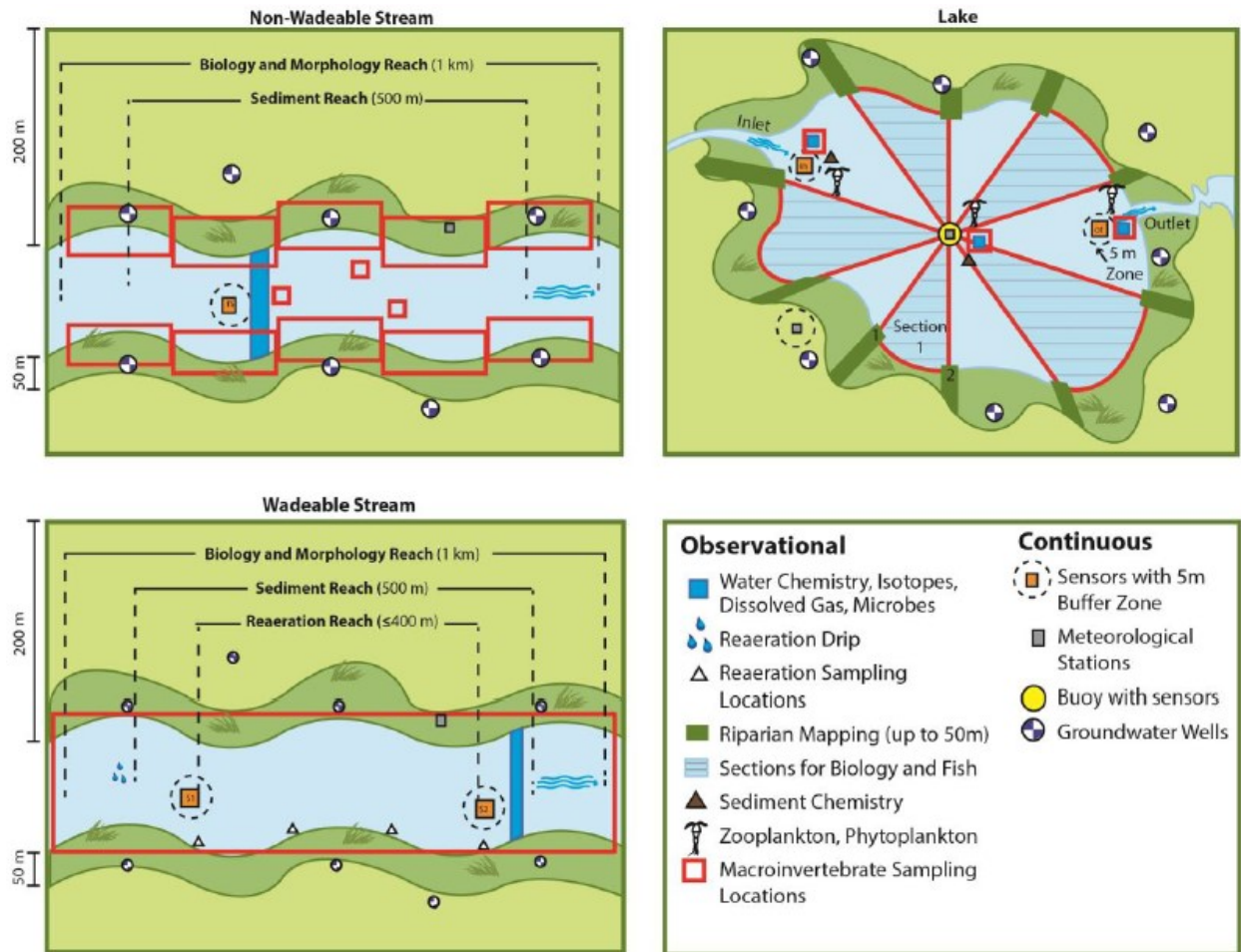


Figure 1: Generic aquatic site layouts (non-wadeable streams/ rivers, wadeable streams, and lakes) with macroinvertebrate sampling locations in red.

3.2 Temporal Sampling Design

Macroinvertebrate sampling occurs three times per year. Timing of sampling is site-specific and determined based on historical hydrological and meteorological data. Sample bout 1 is an early-season date, representing a period of rapid biomass accumulation after winter, typically prior to leaf out or ice-off where applicable. Sample bout 2 targets mid-summer baseflow conditions and sample bout 3 represents the late growing season (typically autumn) during leaf-fall where applicable. These dates differ on a site-by-site basis, but should always occur at, or near, baseflow conditions within the watershed. Sampling does not occur directly following a flood in wadeable streams (defined as $>1.5 \times$ base flow; Biggs et al. 1999). Should such a flood event occur on or prior to a target collection date, sampling is delayed 3 days-1 week (maximum 2 weeks, dependent on field schedule) to allow for invertebrates to recolonize the substratum (c.f. Brooks and Boulton 1991, Matthaei et al. 1996). Data collection for this data product occurs within one day

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per bout at a given site. See NEON Aquatic Sampling Strategy (AD[05]), AOS Protocol and Procedure: Aquatic Macroinvertebrate Sampling (AD[06]) for additional details.

3.3 Variables Reported

All variables reported from the field or laboratory technician (L0 data) are listed in the file, NEON Raw Data Validation for Aquatic Macroinvertebrate Collection (NEON.DP0.20120) (AD[03]). All variables reported in the published data (L1 data) are also provided separately in the file, NEON Data Variables for Aquatic Macroinvertebrate Collection (NEON.DP1.20120) (AD[04]).

Field names have been standardized with Darwin Core terms (<http://rs.tdwg.org/dwc/>; accessed 4 August 2017), the Global Biodiversity Information Facility vocabularies (<http://rs.gbif.org/vocabulary/gbif/>; accessed 16 February 2014), the VegCore data dictionary (<https://projects.nceas.ucsb.edu/nceas/projects/bien/wiki/VegCore>; accessed 16 February 2014), where applicable. NEON Aquatic Observation System (AOS) spatial data employs the World Geodetic System 1984 (WGS84) for its fundamental reference datum and Earth Gravitational Model 96 (EGM96) for its reference gravitational ellipsoid. Latitudes and longitudes are denoted in decimal notation to six decimal places, with longitudes indicated as negative west of the Greenwich meridian.

Some variables described in this document may be for NEON internal use only and will not appear in downloaded data.

3.4 Temporal Resolution and Extent

The finest temporal resolution that macroinvertebrate data will be tracked is per sampling day. All 8 samples are collected within a single day at a particular site. A suite of other biological sampling occurs at the site during the same ~30 day bout. Three sampling bouts occur per site per year.

The finest resolution at which temporal data are reported is at **collectDate**, the date and time of day when the samples were collected in the field.

The NEON Data Portal provides data in monthly files for query and download efficiency. Queries including any part of a month will return data from the entire month. Code to stack files across months is available here: <https://github.com/NEONScience/NEON-utilities>

3.5 Spatial Resolution and Extent

Each macroinvertebrate sample represents a patch of stream bottom within the 1 km permitted wadeable or non-wadeable stream reach, or permitted lake area, and contains multiple individuals. The exact location (latitude and longitude) of each sample is not tracked as it is intended to represent the overall habitat. The **locationID** reported in wadeable streams represents a mid-point in the permitted reach, plus coordinate uncertainty surrounding that point. In lakes and

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non-wadeable streams, some samples are collected near monumented locations associated with a more-specific **locationID**. Sampling locations are tracked by latitude and longitude and include an indication of **coordinateUncertainty**.

Up to two different habitats are sampled at each site to account for the variability or patchiness among habitats. Overall, this results in a spatial hierarchy of:

locationID (finest spatial resolution, ID of location within site) -> siteID (ID of NEON site) -> domainID (ID of a NEON domain)

3.6 Associated Data Streams

A subset of the macroinvertebrate field collection data are related to Macroinvertebrate DNA Barcode (NEON.DP1.20126) samples collected at the same time and location, related samples share the same **parentSampleID**.

Macroinvertebrate collection data are also loosely related to Aquatic General Field Metadata collected on the same sampling day (NEON.DOC.001646). Data for Aquatic General Field Metadata are available in the NEON data product “Gauge Height” (DP1.20267.001). These data products are linked through the **siteID** field and local date in the NEON Data Publication Workbook for AOS Macroinvertebrate Collection (AD[04]).

3.7 Product Instances

At each aquatic site, there will be up to 24 samples collected per year (8 samples per bout). Each sample generates multiple records from the external lab on a per taxon, per size class basis.

3.8 Data Relationships

For each record collected in `inv_fieldData` a number of child records may be created. In the event that sampling is impractical (e.g., the location is dry, ice covered, etc.), there will be no child records. If a **sampleID** is recorded in `inv_fieldData`, there will be one corresponding record in `inv_perSample` (sample sorting and subsampling at the external lab). Each **sampleID** record in `inv_fieldData` may have multiple child records in `inv_taxonomyRaw` and `inv_taxonomyProcessed`, one record for each **scientificName** and **sizeClass** combination. A record from `inv_fieldData` may have multiple or no records in `inv_perVial`, as that table represents individuals removed from the final archived sample and placed in the external lab’s in-house reference collection, records in this table are opportunistic and are organized by **sampleID** and **scientificName**. Duplicates and/or missing data may exist where protocol and/or data entry aberrations have occurred; users should check data carefully for anomalies before joining tables.

`inv_fieldData.csv` - > One record is created for each sample collected in the field, creating a **sampleID** which is linked to all subsequent tables. This table also indicates the field conditions,

including **habitatType**, **samplerType**, **substratumSizeClass**, and sample depth if applicable (e.g., lake and non-wadeable sites).

inv_perSample.csv - > One record (**sampleID**) is created for each sample processed at the expert taxonomy lab. Each sample is sorted (removing macroinvertebrates from organic and inorganic material) and may be subsampled if the number of macroinvertebrates in the sample appears to exceed 300. Data from this table are linked to the fieldData and subsequent taxon data through the **sampleIDs** in each table.

inv_taxonomyRaw.csv - > One record is created for each taxonomic group identified in a sample created in inv_fieldData. Taxonomic identifications are made to the lowest practical taxonomic level (typically genus or species). The taxonomic nomenclature in this file reflects the verbatim identifications provided by the external taxonomist and may contain synonyms. Data are linked to the fieldData and perSample tables through the **sampleIDs** in each table. Records in this table are unique by the combination of **sampleID**, **scientificName**, **morphospeciesID**, **sizeClass**, **immatureSpecimen**, **indeterminateSpecies**, and **identificationQualifier**.

inv_taxonomyProcessed.csv - > One record is created for each taxonomic group identified in a sample created in inv_fieldData. Taxonomic identifications are made to the lowest practical taxonomic level (typically genus or species). The taxonomic nomenclature in this file has been standardized and desynonymized according to NEON's master taxonomy for macroinvertebrates and zooplankton. Data are linked to the fieldData and perSample tables through the **sampleIDs** in each table. Records in this table are unique by the combination of **sampleID**, **scientificName**, **morphospeciesID**, **sizeClass**, **immatureSpecimen**, **indeterminateSpecies**, and **identificationQualifier**.

inv_perVial.csv - > One record is created for each taxonomic group removed from the final archive vial from the sample created in inv_fieldData. Individuals are removed from the archived sample to be kept at the expert taxonomy lab as part of the reference collection. Data are linked to the fieldData, perSample, and perTaxon tables through the **sampleIDs** in each table. Records in this table are unique by the combination of **sampleID** and **scientificName**. Individual organisms documented in inv_taxonomyRaw are returned to a single vial per **sampleID** for archiving. Any individuals removed from that vial to be used by the external lab for the reference collection are documented in the inv_perVial table. The reference collection is housed at the external facility for the life of the contract, and is organized by **domainID** and **scientificName**. The **referenceCount** field indicates the number of organisms that have been removed from the inv_taxonomyRaw vial to be archived.

3.9 Special Considerations

The macroinvertebrate taxonomic counts per size class come from an external lab, in the field **estimatedTotalCount**. This field is corrected for subsampling at the external lab, but is NOT corrected for benthic area. Data users will need to refer to the **benthicArea** presented in the inv_fieldData table and apply this correction to get the number of organisms per stream, lake,

or river bottom. All taxon records from a sample should be summed and divided by the **benthicArea** prior to reporting the total abundance per m².

$$macroinvertebrateAbundancePerM_i^2 = \frac{\sum_{i=1}^n inv_taxonomyProcessed.estimatedTotalCount_i}{inv_fieldData.benthicArea_i} \quad (1)$$

Where ‘i’ is a unique **sampleID**

See the external lab SOP (referenced in `inv_perSample`) for calculations applied to the data by the external laboratory.

4 DATA QUALITY

4.1 Data Entry Constraint and Validation

Many quality control measures are implemented at the point of data entry within a mobile data entry application or web user interface (UI). For example, data formats are constrained and data values controlled through the provision of dropdown options, which reduces the number of processing steps necessary to prepare the raw data for publication. The field data entry workflow for collecting macroinvertebrate field data is diagrammed in Figure 2.

An additional set of constraints are implemented during the process of ingest into the NEON database. The product-specific data constraint and validation requirements built into data entry applications and database ingest are described in the document NEON Raw Data Validation for Aquatic Macroinvertebrate Collection (NEON.DP0.20120) (AD[03]), provided with every download of this data product. Contained within this file is a field named ‘entryValidationRulesForm’, which describes syntactically the validation rules for each field built into the data entry application. Data entry constraints are described in NiCl syntax in the validation file provided with every data download, and the NiCl language is described in NEON’s Ingest Conversion Language (NICL) specifications ([AD[10]).

4.2 Automated Data Processing Steps

Following data entry into a mobile application or web user interface, the steps used to process the data through to publication on the NEON Data Portal are detailed in the NEON Algorithm Theoretical Basis Document: OS Generic Transitions (AD[09]).

4.3 Data Revision

All data are provisional until a numbered version is released; the first release of a static version of NEON data, annotated with a globally unique identifier, is planned to take place in 2020. During

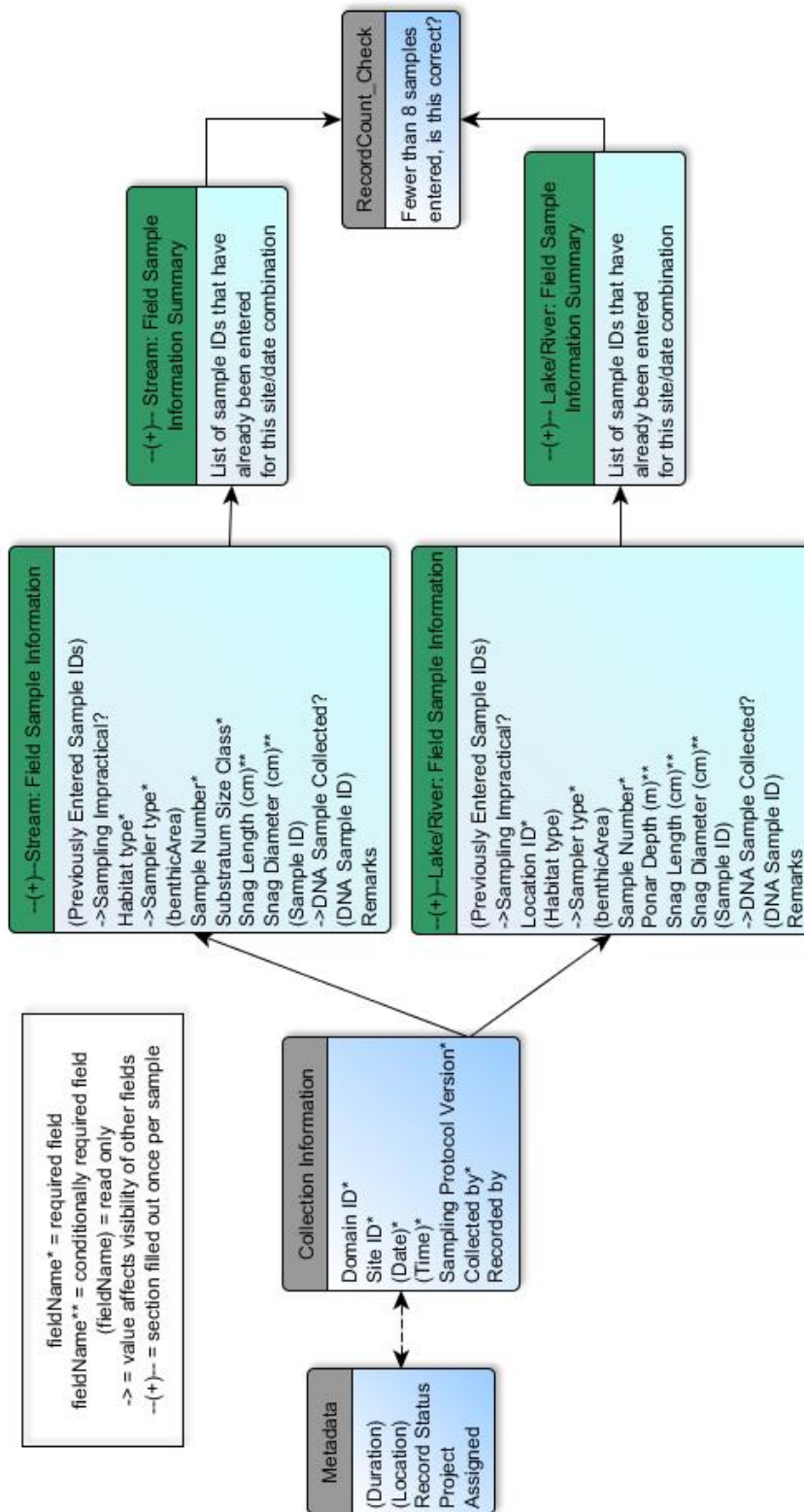


Figure 2: Schematic of the applications used by field technicians to enter macroinvertebrate field data

the provisional period, QA/QC is an active process, as opposed to a discrete activity performed once, and records are updated on a rolling basis as a result of scheduled tests or feedback from data users. The Change Log section of the data product readme, provided with every data download, contains a history of major known errors and revisions.

4.4 Quality Flagging

The **dataQF** field in each data record is a quality flag for known errors applying to the record. Please see the table below for an explanation of **dataQF** codes specific to this product.

fieldName	value	definition
dataQF	legacyData	Data recorded using a paper-based workflow that did not implement the full suite of quality control features associated with the interactive digital workflow

4.5 Analytical Facility Data Quality

Data analyses conducted on macroinvertebrate community data conform to the current data quality standards used by practitioners. Ten percent of all samples are quality checked for taxonomic difference between two taxonomists at the external facility. These records are indicated by the fields **qcChecked**, **qcEnumerationDifference**, and **qcTaxonomicDifference** indicating Percent Difference in Enumeration (PDE) and Percent Taxonomic Difference (PTD) (Stribling et al. 2008). Details on the calculations of these fields can be found in the external lab SOP.

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